

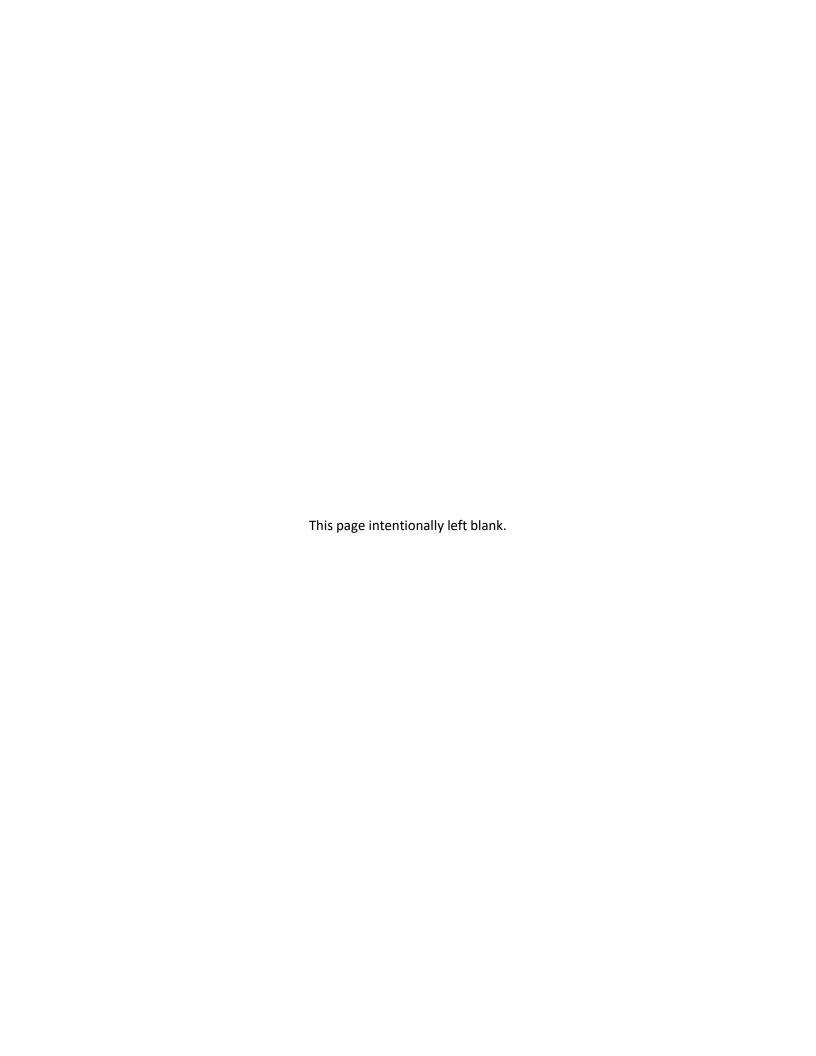
Naval Facilities Engineering Systems Command Northwest Silverdale, Washington

Final

Sampling and Analysis Plan Per- and Polyfluoroalkyl Substances Remedial Investigation Outlying Landing Field Coupeville

Naval Air Station Whidbey Island Oak Harbor, Washington

January 2022



SAP Worksheet #1—Title and Signature Page



Naval Facilities Engineering Systems Command Northwest Silverdale, Washington

Final

Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville

Naval Air Station Whidbey Island Oak Harbor, Washington

January 2022

Prepared for NAVFAC Northwest by CH2M HILL, Inc. Bellevue, Washington Contract N62470-16-D-9000 CTO N4425520F4343



SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUORO NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, W JANUARY 2022 PAGE 2	DALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE //ASHINGTON
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SAP Worksheet #1—Title and Signature Page (continued)

Approval Signatures:

TBD

Naval Facilities Engineering Systems Command Atlantic **Quality Assurance Officer**

Other Approval Signatures:

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CLUBB.KENDRA.R.1374441171 Date: 2022.01.25 11:39:13 -08'00'

PAGE 3

Kendra Clubb Naval Facilities Engineering Systems Command Northwest Remedial Project Manager

Digitally signed by CHAN **CHAN PONGKHAMSING** PONGKHAMSING Date: 2022.01.25 18:29:09

Chan Pongkhamsing United States Environmental Protection Agency Region 10 Remedial Project Manager

SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUORO, NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, W, JANUARY 2022 PAGE 4	ALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE ASHINGTON
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Executive Summary

The Department of the Navy (Navy), Naval Facilities Engineering Systems Command (NAVFAC) Northwest has contracted CH2M HILL, Inc. (CH2M) to conduct a Remedial Investigation (RI) specific to known or suspected releases of per- and polyfluoroalkyl substances (PFAS) to the environment at Outlying Landing Field (OLF) Coupeville in Coupeville, Washington in Island County. This Uniform Federal Policy-Sampling and Analysis Plan/ Quality Assurance Project Plan (SAP) describes the inspection activities, including sampling, to be conducted onbase at the Coupeville property and at selected off-base locations adjacent to OLF Coupeville. CH2M prepared this document under the NAVFAC Comprehensive Long-term Environmental Action — Navy 9000 Contract N62470-16-D-9000, Contract Task Order N4425520F4343, for submittal to NAVFAC Northwest and the United States Environmental Protection Agency (USEPA). CH2M prepared this SAP to help ensure that environmental data collected are scientifically sound, of known and documented quality, and suitable for intended uses.

OLF Coupeville is a military airfield associated with Naval Air Station (NAS) Whidbey Island. It was commissioned for use by the Navy in 1943 and provides support for day and night Field Carrier Landing Practice operations by the Navy for aircraft based out of NAS Whidbey Island. Such operations allow aviators and their crews to fly in patterns as well as practice touch-and-go, simulating carrier landings and take offs. During these practice runs, jet aircraft approach the runway and touch down, immediately taking off again and looping around the field to prepare for another landing and takeoff.

Investigations for PFAS at OLF Coupeville began in 2016. In accordance with Navy policy, off-base drinking water wells were sampled near OLF Coupeville from 2016-2017 (CH2M, 2019a; 2021b). Eight drinking water wells were identified to have perfluorooctane sulfonic acid (PFOS) and/or perfluorooctanoic acid (PFOA) above the USEPA lifetime health advisory. Two expedited Site Inspections (SIs) were conducted from 2016-2018 to better understand the aquifer system and identify the presence of PFAS on-base (CH2M, 2018a); and aquifer testing and groundwater modeling conducted to improve the understanding of aquifer properties and advance the conceptual model of the system (CH2M, 2018b). The results of these previous groundwater investigations at OLF Coupeville identified the presence of PFOS and/or PFOA in groundwater in six on-base groundwater monitoring wells at concentrations exceeding the USEPA lifetime health advisory¹. Additionally, samples collected from the Town of Coupeville's Keystone Well, a community drinking water well located off-Base to the west of OLF Coupeville, indicate that the Keystone Well temporarily exceeded the lifetime health advisory for PFOA and may exceed the advisory level in the future based on current and future pumping rates.

A preliminary assessment (PA) for OLF Coupeville was issued in November 2018 (CH2M, 2018c). The PA identified two on-Base potential release areas (referred to as potential source areas in the PA) of PFAS that warranted further investigation. The two on-base potential release areas were investigated during the 2019-2020 Supplemental SI (CH2M, 2021a). The Supplemental SI confirmed the presence of PFOA and/or PFOS above the USEPA groundwater SLs in several wells near and downgradient of the two potential release areas (formerly called PSAs). PFOA and PFOS concentrations in new monitoring wells indicated that the migration of PFAS at OLF is predominantly to the south-southwest with a vertically downward trajectory. However, the extent of the PFOA and PFOS impacts in groundwater are not consistent with low soil concentrations at depth near Building 2709 and Facilities 1, 2, and 11. PFOA and PFOS were also detected in monitoring wells west of the runway near the Keystone Well, and these concentrations are likely attributable to a source area near the northwestern end of the runway and may also be contributing to the PFAS concentrations in the Keystone Well. The RI sampling described herein seeks to delineate the horizontal and vertical extent of PFAS in soil, groundwater, and surface water (if

At the time of the Expedited SI, these concentrations were screened against the USEPA lifetime health advisory of 70 nanograms per liter (ng/L) each for PFOA and PFOS, and 70 ng/L for combined PFOA and PFOS. However, the USEPA lifetime health advisory is no longer used for Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) decision making. The Navy's current policy is to use the screening levels for soil and groundwater consistent with the Department of Defense (DoD) technical guidance for PFAS investigations (ASD, 2021) and EPA, 2021.

present) at OLF Coupeville and to address data gaps relating to PFAS transport pathways and suspected additional PFAS release areas. The objectives of the RI are as follows:

- Define the horizontal and vertical extent of PFAS at the on-base release areas (Building 2709 and Facilities 1, 2, and 11) and determine whether there is unacceptable risk to human health and ecological² receptors.
- Confirm the presence of an additional on-base release area west of the runway near the Keystone Well and, if
 identified, define the nature and extent of PFAS concentrations in this area and determine whether there is
 unacceptable risk to human health and ecological² receptors.
- Refine the data representative of groundwater flow and potential PFAS migration directions between the onbase release areas, the release area west of the runway, and impacted on-base groundwater monitoring wells and off-base drinking water wells with exceedances of the USEPA lifetime health advisory for PFOA and PFOS.
- Refine the data demonstrating temporal variability in PFAS concentrations in groundwater at OLF Coupeville.
- Determine whether PFAS are present in ditch surface soil and surface water (if surface water is present/observed), define the nature and extent of PFAS in ditch surface soil and surface water (if present), and determine whether there is unacceptable risk to human health and ecological² receptors.

It is anticipated that the RI will be accomplished during at least two field investigation phases, the first of which will include the on-base fieldwork described herein, with additional phases, including the off-Base field work to be described in an addendum to this SAP. If additional data is needed to accomplish the RI objectives, additional phases may be performed which will be described under future task order work plan documents. The Phase 1 investigation described in this SAP will include the following activities:

- Conduct surface and subsurface soil sampling at or near Building 2709 and Facilities 1, 2, and 11.
- Install an on-base shallow-zone groundwater monitoring well east of Building 2709.
- Collect depth-discrete groundwater grab samples and install an on-base shallow/intermediate groundwater monitoring well pair between the on-base release areas and on-base groundwater monitoring wells to the southwest where PFOA has been detected in exceedance of the SL.
- Conduct surface and subsurface soil sampling west of the runway near the Keystone Well.
- Collect depth-discrete groundwater grab samples and install on-base shallow/intermediate groundwater monitoring well pairs west of the runway near the Keystone Well.
- Collect depth-discrete groundwater grab samples and install on-base intermediate/deep groundwater monitoring well pairs near the southern end of the runway and the southwestern Base boundary.
- Collect paired surface water (if present) and ditch surface soil samples from on-base runway drainage ditches.
- Analyze all samples for the 18 PFAS listed in Method 537.1 via Liquid Chromatography Tandem Mass Spectrometry (USEPA, 2020a) in compliance with the DoD Quality Systems Manual (QSM) Version 5.3 Table B-15 or the most recent version of the DoD QSM for which Battelle has DoD Environmental Laboratory Accreditation Program (ELAP) certification.

PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for both the Human Health and Ecological Risk Assessments. Prior to conducting an ecological risk screening, the ecological screening values that will be used for comparison to site media will be reviewed based on current science and relevancy to site-specific receptors (e.g., relevant ecological receptors present). Ecological screening values selected for the screening will be presented in a technical memo or other documented form for stakeholder review and acceptance.

- Collect lithologic data from the newly installed groundwater monitoring wells and soil borings for further understanding of the overall conceptual site model for the site.
- Survey each newly installed groundwater monitoring well and soil boring location.

This SAP was developed in accordance with the following guidance documents:

- Guidance for Quality Assurance Project Plans (USEPA, 2002)
- Uniform Federal Policy for Quality Assurance Project Plans: Evaluating, Assessing, and Documenting Environmental Data Collection and Use Programs - Part 1: UFP-QAPP Manual (USEPA, 2005)
- Guidance on Systematic Planning Using the Data Quality Objectives Process (USEPA, 2006)
- Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/November 2020 Update (NAVFAC, 2020)

Additional samples of groundwater, soil, ditch surface soil and surface water (the media described herein) may be collected in locations where data gaps continue to exist to delineate PFAS to PALs or assess fate and transport pathways, consistent with stakeholder agreement and in accordance with procedures outlined in this SAP. Media not explicitly discussed in this SAP, such as drinking water, will not be sampled at this time or under this SAP.

This SAP consists of 37 worksheets specific to the scope of the RI. All tables are embedded within the worksheets. All figures are included at the end of the document. The scoping presentation provided to stakeholders in the December 2020 scoping session is included in **Appendix A**. Field standard operating procedures (SOPs) are included in **Appendix B**. The DoD-ELAP Accreditation letter is included in **Appendix C**. Laboratory SOPs are included in **Appendix D**.

The laboratory information cited in this SAP is specific to Battelle. If additional laboratory services are necessary to meet the project objectives, revised SAP worksheets will be submitted to NAVFAC and regulatory agencies (as appropriate) for approval and appended to this SAP.

SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON ANUARY 2022 PAGE 8	

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Appendixes

- A RI Scoping Session Presentation
- B Field Standard Operating Procedures
- C Department of Defense Environmental Laboratory Accreditation Program Accreditation Letter
- D Laboratory Standard Operating Procedures

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- 10-1 OLF Coupeville Conceptual Site Model
- 11-1 Problem Quality Objectives/Systematic Planning Process Statements
- 17-1 Sampling Strategy Table OLF Coupeville RI

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- 10-1 Base Location Map
- 10-2 Site Layout Map
- 10-3 Off-Base Drinking Water Sampling Area and Results
- 10-4 Cross Section Locations
- 10-5 Geologic Cross Section A-A'
- 10-6 Geologic Cross Section B-B'
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- 10-11 Groundwater Elevation Contour Map Intermediate Elevation Interval
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- 10-13 Summary of Soil PFAS Concentrations
- 10-14 PFOA Concentrations in Groundwater Shallow Aquifer Zone
- 10-15 PFOA Concentrations in Groundwater Intermediate Aquifer Zone
- 10-16 PFOA Concentrations in Groundwater Deep Aguifer Zone
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- 10-18 PFOS Concentrations in Groundwater Intermediate Aquifer Zone
- 10-19 PFOS Concentrations in Groundwater Deep Aquifer Zone
- 11-1 Decision Logic for Project Quality Objectives
- 11-2 Proposed RI Drilling and Sampling Locations

Acronyms and Abbreviations

± plus or minus% percent> greater than< less than

≤ less than or equal to °C degree(s) Celsius

μg/kg microgram(s) per kilogram

11Cl-PF3OUdS 11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid 9Cl-PF3ONS 9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid

ADONA 4,8-dioxa-3H-perfluoronanoic acid

AFFF aqueous film-forming foam

AM Activity Manager

AQM Activity Quality Manager
ASD Assistant Secretary of Defense

BERA Baseline Ecological Risk Assessment

bgs below ground surface

CA corrective action

CAS Chemical Abstracts Service
CCV continuing calibration verification

CERCLA Comprehensive Environmental Response, Compensation, and Liability Act

CH2M CH2M HILL, Inc.

CLEAN Comprehensive Long-term Environmental Action—Navy

CSM conceptual site model

DL detection limit

DoD Department of Defense

DV data validator

Ecology Washington Department of Ecology

EDD electronic data deliverable

ELAP Environmental Laboratory Accreditation Program

ESV ecological screening value

EtFOSAA n-ethyl perfluorooctanesulfonamidoacetic acid

FD field duplicate
FTL Field Team Leader

H&S health and safety

HDPE high-density polyethylene

HFPO-DA perfluoro-2-methyl-3-oxahexanoic acid

HQ hazard quotient

HSM Health and Safety Manager
HSP Health and Safety Plan

ICAL initial calibration ID identification

IDW investigation-derived waste ISC instrument sensitivity check

LC/MS/MS liquid chromatography – tandem mass spectrometer

LCS laboratory control sample
LCL lower confidence limit
LOD limit of detection
LOQ limit of quantitation

MD matrix duplicate

MeFOSAA n-methyl perfluorooctanesulfonamidoacetic acid

MPC measurement performance criteria

MS matrix spike

MSD matrix spike duplicate

N/A not applicable
NAS Naval Air Station

NAVD88 North American Vertical Datum of 1988

NAVFAC Naval Facilities Engineering Systems Command

Navy Department of the Navy

NEtFOSAA n-ethylperfluoro-1-octancesulfonamidoacetic acid

ng/L nanogram(s) per liter

NMeFOSAA n-methylperfluoro-1-octanesulfonamidoacetic acid

NTR Navy Technical Representative

OLF Outlying Landing Field

PA Preliminary Assessment

PAL project action limit

PC Project Chemist

PFAS per- and polyfluoroalkyl substances

PFBS perfluorobutane sulfonate **PFDA** perfluorodecanoic acid **PFDoA** perfluorododecanoic acid **PFPeA** perfluoropentanoic acid **PFHxA** perfluorohexanoic acid perfluoroheptanoic acid **PFHpA PFHxS** perfluorohexanesulfonic acid **PFNA** perfluorononanoic acid **PFOA** perfluorooctanoic acid

PFOS perfluorooctane sulfonic acid
PFTeDA perfluorotetradecanoic acid
PFTrDA perfluorotridecanoic acid
PFUnA perfluoroundecanoic acid
PID photoionization detector

PM Project Manager POC point of contact

PPR provisional peer-reviewed
PQL project quantitation limit
PQO Project Quality Objective

QA quality assurance

QAO Quality Assurance Officer

QC quality control

QSM Quality Systems Manual

QSM 5.3 Quality Systems Manual Version 5.3

RI Remedial Investigation
RPD relative percent difference
RPM Remedial Project Manager
RSL regional screening level

SAP Sampling and Analysis Plan

SI site inspection SL screening level

SLERA Screening-Level Ecological Risk Assessment

SME Subject Matter Expert

SOP standard operating procedure

SPE solid phase extraction

SSHO Site Safety and Health Officer

SSL soil screening level

STC Senior Technical Consultant

TAT turnaround time
TBD to be determined
TM Task Manager

UCL upper confidence limit

USEPA United States Environmental Protection Agency

AMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE IAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON
ANUARY 2022
AGE 14

SAP Worksheet #2—Sampling and Analysis Plan Identifying Information

Site Name/Number: Outlying Landing Field (OLF) Coupeville, Naval Air Station (NAS) Whidbey Island

Operable Unit: Not Applicable (N/A)

Contractor Name: CH2M HILL, Inc. (CH2M)

Contract Number: N62470-16-D-9000, Contract Task Order 4343

Contract Title: Comprehensive Long-term Environmental Action – Navy (CLEAN) Program 9000

Work Assignment: Remedial Investigation (RI) specific to releases of per- and polyfluoroalkyl substances

(PFAS) to the environment for Naval Facilities Engineering Systems Command (NAVFAC)

Northwest at the OLF in Coupeville, Washington

1. This Sampling and Analysis Plan (SAP) was prepared in accordance with the following guidance documents:

- Interim Final Guidance for Conducting Remedial Investigations and Feasibility Studies Under CERCLA (USEPA, 1988)
- Superfund Amendments and Reauthorization Act (USEPA, 1986)
- Guidance for Quality Assurance Project Plans (USEPA, 2002)
- Uniform Federal Policy for Quality Assurance Project Plans: Evaluating, Assessing, and Documenting Environmental Data Collection and Use Programs - Part 1: UFP-QAPP Manual (USEPA, 2005)
- Guidance on Systematic Planning Using the Data Quality Objectives Process (USEPA, 2006)
- Naval Facilities Engineering Command Sampling and Analysis Plans Update on the Uniform Federal Policy Quality Assurance Project Plans (UFP-QAPP) – Navy Sampling and Analysis Plans (SAP) Tiered Approach (Navy, 2011a)
- Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/November 2020 Update (NAVFAC, 2020)
- Department of the Navy Environmental Restoration Program Manual (Navy, 2018)
- 2. Identify regulatory Program: Comprehensive Environmental Response, Compensation, and Liability Act of 1980
- 3. This document is a project-specific SAP.
- 4. List dates of scoping sessions that were held:

Scoping Session	Date
Project Scoping Session with NAVFAC Northwest Remedial Project Manager (RPM), United States Environmental Protection Agency (USEPA), Washington Department of Ecology (Ecology), Washington Department of Health, Island County, and Town of Coupeville	December 10, 2020

SAP Worksheet #2—Sampling and Analysis Plan Identifying Information (continued)

5. List dates and titles of any SAP documents written for previous site work that are relevant to the current investigation:

Document	Date
Sampling and Analysis Plan, Investigation of Perfluorinated Compounds in Drinking Water, Ault Field and Outlying Landing Field Coupeville, Naval Air Station Whidbey Island, Oak Harbor and Coupeville, Washington (CH2M, 2017a)	January 2017
Sampling and Analysis Plan, Site Inspection for Perfluorinated Compounds in Groundwater, Outlying Landing Field Coupeville, NAS Whidbey Island, Coupeville, Washington (CH2M, 2017b)	January 2017
Sampling and Analysis Plan, Monitoring Well Installation, Aquifer Testing, Drinking Water Sampling, and Groundwater Sampling, Outlying Landing Field Coupeville, NAS Whidbey Island, Coupeville, Washington (CH2M, 2017c)	December 2017
Sampling and Analysis Plan, Supplemental Site Inspection, Outlying Landing Field Coupeville, NAS Whidbey Island, Oak Harbor, Washington (CH2M, 2019b)	August 2019
Sampling and Analysis Plan Addendum, Supplemental Site Inspection, Outlying Landing Field Coupeville, NAS Whidbey Island, Oak Harbor, Washington (CH2M, 2020a)	August 2020

6. List organizational partners (stakeholders) and identify the connection with lead organization:

Organization Partners/Stakeholders	Connection
NAVFAC Northwest – Kendra Clubb	RPM
NAVFAC Atlantic	Quality Assurance Officer (QAO)
USEPA Region 10 – Chan Pongkhamsing	Project Manager (PM)
Washington Department of Ecology – Mike Shaljian	Hydrogeologist - Toxics Cleanup Program

7. Lead organization:

Department of the Navy (Navy) – NAVFAC Northwest

- 8. If any required SAP elements and required information are not applicable to the project or are provided elsewhere, then note the omitted SAP elements and provide an explanation for their exclusion as follows:
 - Crosswalk table is excluded because all required information is provided in this SAP.

SAP Worksheet #3—Distribution List

Name of SAP Recipients	Title/Role	Organization	Telephone Number	Email Address or Mailing Address
Kendra Clubb	RPM/Task Order Contracting Officer's Representative	NAVFAC Northwest	(509) 999-6843	kendra.leibman@navy.mil
Charlie Escola	Navy Technical Representative (NTR)	NAVFAC Northwest	(503) 201-5020	charles.escola@navy.mil
Steve Skeehan	NTR	NAVFAC Northwest	(253) 279-0212	Steve.skeehan@navy.mil
To be determined (TBD)	NAVFAC QAO	NAVFAC Atlantic	TBD	TBD
Chan Pongkhamsing	PM	USEPA Region 10	(206) 553-1806	pongkhamsing.chan@epa.gov
Mike Shaljian	Hydrogeologist - Toxics Cleanup Program	Ecology	(360) 489-2753	mish461@ECY.WA.GOV
Joe Grogan	Utility Manager	Town of Coupeville	(360) 914-0314	utilities1@townofcoupeville.org
Jennifer Madsen	Activity Manager (AM), PM	CH2M	(360) 888-0281	jennifer.madsen@ch2m.com
Rachel Clennon	Task Manager (TM)	CH2M	(978) 387-8171	rachel.clennon@ch2m.com
Peter Lawson	Senior Technical Consultant (STC)	CH2M	(530) 949-0870	peter.lawson@ch2m.com
Paul Townley	Activity Quality Manager (AQM)	CH2M	(206) 295-6904	paul.townley@ch2m.com
Laura Cook	Subject Matter Expert (SME)	CH2M	(757) 671-6214	laura.cook@ch2m.com
Janna Staszak	Program SAP Quality Reviewer	CH2M	(757) 268-6136	janna.staszak@ch2m.com
Anita Dodson	Navy CLEAN Program Chemist/SAP Reviewer	CH2M	(757) 284-9208	anita.dodson@ch2m.com
Travis Pitts	Project Chemist (PC)	CH2M	(541) 768-3727	travis.pitts@ch2m.com
Doug Weaver	Data Validator (DV)	Environmental Data Services, Inc. (EDS)	(757) 564-0090	dweaver@end-data.com
TBD	Field Team Leader (FTL)	CH2M	TBD	TBD
TBD	Site Safety and Health Officer (SSHO)	CH2M	TBD	TBD
Jonathan Thorn	Laboratory PM	Battelle	(781) 681-5565	thorn@battelle.org

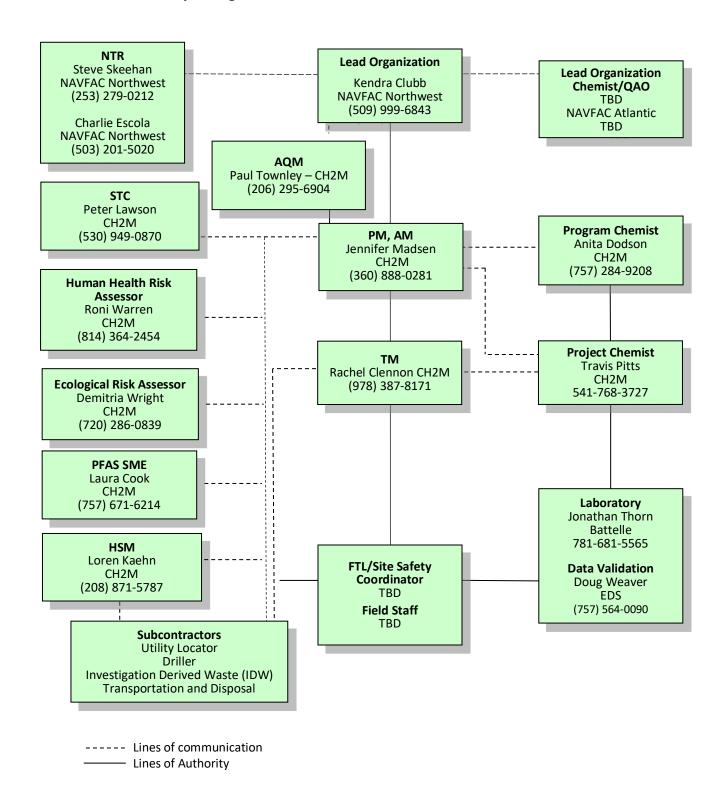
SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON
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SAP Worksheet #4—Project Personnel Sign-Off Sheet

Name	Organization/Title/Role	Telephone Number	Signature/Email Receipt	SAP Section Reviewed	Date SAP Read
Jennifer Madsen	CH2M/AM, PM	(360) 888-0281			
Peter Lawson	CH2M/STC	(530) 949-0870			
Paul Townley	CH2M/AQM	(206) 295-6904			
Laura Cook	CH2M/SME	(757) 671-6214			
Rachel Clennon	CH2M/Project TM	(978) 387-8171			
Anita Dodson	CH2M/Navy CLEAN Program Chemist/SAP Reviewer	(757) 284-9208			
Travis Pitts	CH2M/PC	(541) 768-3727			
TBD	CH2M/FTL	TBD			
TBD	CH2M/SSHO	TBD			
Jonathan Thorn	Battelle/Laboratory PM	(781) 681-5565			
Doug Weaver	Environmental Data Services, Inc. (EDS)/Data Validation PM	(757) 564-0090			

SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUORO NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, W JANUARY 2022 PAGE 20	PALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE VASHINGTON

SAP Worksheet #5—Project Organizational Chart



SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON JANUARY 2022 PAGE 22	

SAP Worksheet #6—Communication Pathways

Communication Drivers	Responsible Entity	Name	Phone Number	Procedure
Communication with Navy (lead agency)	NAVFAC Northwest NTR	Steve Skeehan	steve.skeehan@navy.mil (253) 279-0212	Primary point of contact (POC) for NAVFAC Northwest for the contractor during fieldwork; oversees fieldwork, provides base-specific information, provides coordination with NAS Whidbey Island, and can delegate communication to other internal POCs.
Communication with Navy (lead agency)	NAVFAC Northwest NTR	Charlie Escola	charles.escola@navy.mil (503) 201-5020	Primary POC for NAVFAC Northwest for the contractor during fieldwork; oversees fieldwork, provides base-specific information, provides coordination with NAS Whidbey Island, and can delegate communication to other internal POCs.
Communication with Navy (lead agency)	NAVFAC Northwest RPM	Kendra Clubb	kendra.leibman@navy.mil (509) 999-6843	Primary POC for NAVFAC Northwest; can delegate communication to other internal or external POCs. CH2M PM will notify the NTR, RPM, and NAVFAC Atlantic QAO by email or telephone call within 24 hours for changes affecting the scope or implementation of the SAP.
Communication with Navy (lead agency)	NAVFAC Atlantic QAO	TBD	TBD	Primary POC for NAVFAC LANT.
Communication regarding overall project status Communication regarding overall project status and implementation and primary POC with RPM and project team	AM, PM	Jennifer Madsen	jennifer.madsen@ch2m.com (360) 888-0281	Oversees project and will be informed of project status by the PM and TM. All data results will be communicated to appropriate team members following data receipt and review.
Communication regarding items specific to OLF tasks and primary POC for field team	CH2M TM	Rachel Clennon	rachel.clennon@ch2m.com (978) 387-8171	Oversees the investigation task and will be informed of task status by the FTL. If field changes are necessary, TM will work with the PM and RPM to prepare a Field Change Request to be submitted to the NTR and RPM and will communicate in-field changes to the team by email within 24 hours. All data results will be communicated to appropriate team members following data receipt and review.

SAP Worksheet #6—Communication Pathways (continued)

Communication Drivers	Responsible Entity	Name	Phone Number	Procedure
Technical communications for project implementation, and data interpretation	CH2M STC	Peter Lawson	peter.lawson@ch2m.com (530) 949-0870	Contact STC regarding questions/issues encountered in the field, input on data interpretation, as needed. The STC will have 24 hours to respond to technical field questions as necessary. Additionally, the STC will review the data as necessary prior to discussions with NAVFAC Northwest and reporting review.
Quality issues	CH2M AQM	Paul Townley	paul.townley@ch2m.com (206) 295-6904	Contact AQM regarding quality issues during project implementation. The AQM will report to the PM, RPM, and NTR.
Technical communications for project implementation, and data interpretation	CH2M SME	Laura Cook	laura.cook@ch2m.com (757) 671-6214	Contact SME regarding questions/issues encountered in the field, input on data interpretation, as needed. SME will have 24 hours to respond to technical field questions as necessary. Additionally, SME will review the data as necessary prior to Base and NAVFAC Northwest discussions and reporting review.
Health and safety (H&S)	CH2M Health and Safety Manager (HSM)	Loren Kaehn	loren.kaehn@ch2m.com (208) 871-5787	Responsible for generation of the Health and Safety Plan (HSP) and approval of the activity hazard analyses prior to the start of fieldwork. The PM will contact the HSM as needed regarding questions/issues encountered in the field.
H&S	CH2M SSHO	TBD	TBD	Responsible for the adherence of team members to the site safety requirements described in the HSP. Will report H&S incidents and near losses to the PM as soon as possible.
	CH2M PM	Jennifer Madsen	jennifer.madsen@ch2m.com (360) 888-0281	
Stop Work Order	СН2М ТМ	Rachel Clennon	rachel.clennon@ch2m.com (978) 387-8171	Any field member can immediately stop work if an unsafe condition that is immediately threatening to human health is observed. The field staff, FTL, or SSHO should notify the NTR,
	CH2M FTL/ SSHO	TBD	TBD	RPM, and the CH2M PM immediately. Ultimately, the FTL and PM can stop work for a period of time. NAVFAC Northwest can stop work at any time.
	Field Team Members	TBD	TBD	

SAP Worksheet #6—Communication Pathways (continued)

Communication Drivers	Responsible Entity	Name Phone Number		Procedure
SAP changes in field	FTL TBD TBD		TBD	Documentation of deviations from the SAP will be made in the field notes, and the PM will be notified immediately. The RPM and Navy QAO will be also be notified, and deviations will be made only with approval from the PM and approval of a field change request by the RPM.
Field changes/field progress reports	FTL	TBD	TBD	Documentation of field activities and SAP deviations (made with the approval of STC and/or QAO) in field notes; provide daily progress reports to PM.
Reporting laboratory data quality issues	Laboratory PM	Jonathan Thorn	thorn@battelle.org (781) 681-5565	All quality assurance (QA)/quality control (QC) issues with project field samples will be reported within 2 days to the PC by the laboratory.
Communication to Navy QAO regarding SAP changes	Navy CLEAN Program Chemist	Anita Dodson	anita.dodson@ch2m.com (757) 284-9208	Changes to the project that would prompt a SAP change that would require Navy QAO approval include the following: the addition of an analytical suite not previously included in the SAP, the addition of an environmental matrix not previously included in the SAP, laboratory accreditation to a new Department of Defense (DoD) Quality Systems Manual (QSM) version, inclusion of a new laboratory into the SAP for any reason, or updates to the conceptual site model (CSM) that prompt new DQOs. Updated laboratory limit of quantitation (LOQ), limit of detection (LOD), and detection limit (DL) values will not prompt a SAP update for Navy QAO approval unless those updates negatively affect the ability to meet project action limits (PALs).
Analytical corrective actions (CAs)	PC	Travis Pitts	travis.pitts@ch2m.com (541) 768-3727	Any CAs for field and analytical issues will be determined by the PC and reported to the PM within 4 hours. The PM will ensure SAP requirements are met by field staff for the duration of the project.

SAP Worksheet #6—Communication Pathways (continued)

Communication Drivers	Responsible Entity	Name	Phone Number	Procedure	
Data tracking from field collection to database upload Release of analytical data	PC	Travis Pitts	travis.pitts@ch2m.com (541) 768-3727	Tracks data from sample collection through database upload daily. No analytical data can be released until validation of the data is completed and has been approved by the PC. The PC will review analytical results within 24 hours of receipt for release to the PM. The PC will inform the Navy CLEAN Program Chemist, who will notify the Navy QAO and RPM of any laboratory issues that would prevent the project from meeting project quality objectives (PQOs) or would cause significant delay in the project schedule.	
Reporting data quality issues	DV	Doug Weaver	dweaver@end-data.com (757) 564-0090	The DV reviews and qualifies analytical data as necessary. The data along with a validation narrative are returned to the PC within 7 calendar days.	
Field CAs	FTL, AM, PM, and Project TM	FTL (TBD) Jennifer Madsen (AM/PM) Rachel Clennon (TM)	TBD jennifer.madsen@ch2m.com (360) 888-0281 rachel.clennon@ch2m.com (978) 387-8171	Field and analytical issues requiring CA will be determined by the FTL, TM and/or PM on an as-needed basis. The PM will ensure SAP requirements are met by field staff for the duration of the project. The FTL will notify the PM via phone of any need for CA within 4 hours. The PM will notify the NTR and RPM of any field issues that would negatively affect schedule or the ability to meet project data quality objectives.	

SAP Worksheet #7—Personnel Responsibilities Table

Name	Title/Role	Organizational Affiliation	Responsibilities
Kendra Clubb	RPM	NAVFAC Northwest	Oversees entire project for Navy; reviews and approves SAP and changes to SAP; provides base-specific information and coordinates with NAS Whidbey Island.
Charlie Escola	NTR	NAVFAC Northwest	Oversees fieldwork; provides base-specific information, and coordination with NAS Whidbey Island.
Steve Skeehan	NTR	NAVFAC Northwest	Oversees fieldwork; provides base-specific information, and coordination with NAS Whidbey Island.
TBD	NAVFAC QAO/Chemist	NAVFAC Atlantic	Provides QA oversight and reviews SAPs.
Jennifer Madsen	AM, PM	CH2M	Oversees and manages project activities.
Rachel Clennon	TM	CH2M	Oversees and manages all tasks associated with OLF.
Peter Lawson	STC	CH2M	Provides senior technical support for project approach and execution.
Paul Townley	AQM	CH2M	Provides QA oversight.
Laura Cook	SME	CH2M	Provides senior technical support for project approach and execution.
Janna Staszak	SAP Reviewer	CH2M	Reviews and approves the SAP and changes or makes revisions to the SAP.
Anita Dodson	Navy CLEAN Program Chemist/SAP Reviewer	CH2M	Provides SAP project delivery support, reviews and approves SAPs, and performs final data evaluation and QA oversight.
Travis Pitts	PC	CH2M	Data management: Performs data evaluation and QA oversight, is the POC with laboratory and validator for analytical issues.
Loren Kaehn	HSM	CH2M	Prepares HSP and manages H&S for all field activities.
Doug Weaver	DV	Environmental Data Services, Inc. (EDS)	Validates laboratory data from an analytical standpoint prior to data use.
TBD	FTL	CH2M	Coordinates all field activities and sampling.
TBD	Field Staff	CH2M	Conducts field activities.
Jonathan Thorn	Laboratory PM	Battelle	Manages samples tracking and maintains good communication with PC.
Zachary Willenberg	Laboratory QAO	Battelle	Responsible for audits, CA, and checks of QA performance within the laboratory.

SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON
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SAP Worksheet #8—Special Personnel Training Requirements Table

No specialized training beyond standard H&S training is required for this project.

SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE
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SAP Worksheet #9—Project Scoping Session Participants Sheet

Project Name: OLF Coupeville RI Site Name: OLF Coupeville

Projected Date(s) of Sampling: September to December 2021 | **Site Location:** Coupeville, Washington

PM: Jennifer Madsen/CH2M

Date of Session: Thursday, December 10, 2020

Scoping Session Purpose: To obtain consensus on overall objectives of the investigation at OLF Coupeville and discuss

proposed investigation scope.

Name	Title/Project Role	Affiliation	Phone Number	Email Address
Kendra Clubb	NAVFAC Northwest RPM	NAVFAC Northwest	(509) 999-6843	kendra.leibman@navy.mil
Melissa Gehrmann	NAS Whidbey Island Public Works Department Installation Environmental Program Director, Acting	NAVFAC Northwest	TBD	melissa.s.gehrmann1@navy.mil
Chan Ponghkamsing	USEPA Region 10 RPM	USEPA	(206) 553-1806	pongkhamsing.chan@epa.gov
Ted Repasky	Hydrogeologist	USEPA	TBD	repasky.ted@epa.gov
Doug Kelly	Hydrogeologist	Island County	(360) 678-7885	d.kelly@islandcountywa.gov
Mike Shaljian	Hydrogeologist	Ecology	(360) 489-2753	mish461@ecy.wa.gov
Steve Hulsman	SME	Washington Department of Health	TBD	steve.hulsman@dlh.wa.gov
Joe Grogan	Utility Superintendent	Town of Coupeville	(360) 914-0314	utilities1@townofcoupeville.org
Jennifer Madsen	PM/AM	CH2M	(360) 888-0281	jennifer.madsen@ch2m.com
Rachel Clennon	TM	CH2M	(978) 387-8171	rachel.clennon@ch2m.com
Peter Lawson	STC	CH2M	(530) 949-0870	peter.lawson@ch2m.com
Heather Perry	Hydrogeologist SME /Groundwater modeler	CH2M	(530) 355-1622	heather.perry@ch2m.com
Gerrit Gardner	SAP Lead Author	CH2M	(609) 769-7302	gerrit.gardner@ch2m.com

Comments

The scoping meeting consisted of two sessions conducted on December 10, 2020. During the first session, NAVFAC Northwest and CH2M presented an overview of the Supplemental Site Inspection (SI) and groundwater modeling results. During the second session, NAVFAC Northwest and CH2M presented preliminary locations for soil borings, new groundwater monitoring wells, and surface water (if present) and ditch surface soil sampling where data gaps persist. **Appendix A** provides the scoping presentation, which was shared with stakeholders during this session.

SAP Worksheet #9—Project Scoping Session Participants Sheet (continued)

USEPA expressed a concern that the preliminary number of soil borings may not be enough to generate a sufficient amount of data to properly evaluate the extent of PFAS in the soil at Building 2709 and Facilities 1, 2, and 11, or at the suspected release area near the Keystone Well and WI-CV-MW15/WI-CV-MW16 well cluster. NAVFAC Northwest acknowledged this concern and explained that the preliminary number of borings for the RI should be seen as an appropriate next step in the process and that the phased approach would allow for additional soil borings in the future.

Fieldwork for Phase 1 of the RI is expected to begin in fall 2021.

Consensus Decisions

USEPA, Ecology, Island County, and the Town of Coupeville agreed with the RI approach that NAVFAC Northwest and CH2M presented, including the RI objectives, implementation of separate investigation phases for on-base and off-base field tasks, and the media to be sampled (soil, groundwater, ditch surface soil, and surface water if present). The stakeholders provided concurrence with regard to general locations of soil borings near confirmed and potential PFAS release areas, proposed on- and off-base monitoring well locations, and proposed on- and off-base surface water and ditch surface soil sampling locations for Phases 1 and 2 of the investigation. Media sampled will be analyzed for PFAS via liquid chromatography – tandem mass spectrometer (LC/MS/MS) in compliance with DoD Quality Systems Manual Version 5.3 (QSM 5.3) Table B-15 or the most recent version of the DoD QSM for which the lab is accredited at the time of analysis.

Action Items

- Refine scope details and schedule for Phases 1 and 2 of the RI field investigation.
- Generate the project SAP, Waste Management Plan-Environmental Protection Plan, and Accident Prevention
 Plan-Site Safety and Health Plan. Note that the subsequent phases of the RI field investigation will be
 described in future addenda to this SAP document.
- Begin procurement of access agreements for off-base parcels for off-base phases of the RI field investigation.

SAP Worksheet #10—Conceptual Site Model

OLF Coupeville is located two miles southeast of the Town of Coupeville, Washington, in Island County (**Figure 10-1**). **Figure 10-2** presents the site layout of OLF Coupeville. **Table 10-1** presents the CSM for OLF Coupeville, NAS Whidbey Island, Coupeville, Washington.

Table 10-1. OLF Coupeville Conceptual Site Model

Table 10-1. OLF Coupeville Co	•				
Site Name	OLF Coupeville, NAS Whidbey Island, Coupeville, Washington				
Study Area Description	OLF Coupeville is a military airfield associated with NAS Whidbey Island. It was commissioned for use by the Navy in 1943 and provides support for day and night Field Carrier Landing Practice operations by the Navy for aircraft based out of NAS Whidbey Island. Such operations allow aviators and crew to fly in patterns as well as practice touch-and-go, simulating carrier landings and take offs. During these practice runs, jet aircraft approach the runway and touch down, immediately taking off again and looping around the field to prepare for another landing and takeoff.				
	The areas to be investigated consist of areas near the confirmed release areas of Building 2709 and Facilities 1, 2, and 11, on-base near the Keystone Well (west of the runway), on-base near the southern end of the runway and the southwestern Base boundary, and on-base drainage ditches near the runway. The generalized areas of investigation are outlined on Figure 10-2 .				
	Sampling of on-base Drinking Water Wells – September 2016				
	PFAS were first detected in the on-base supply well located in the southwestern portion of the OLF Coupeville near Building 2807 (Figure 10-2) during groundwater sampling activities conducted by the Navy in November September 2016 (Navy, 2014, 2016). The on-base wells were resampled in fall 2020; results indicated that concentrations of perfluorooctanoic acid (PFOA) at this location exceeded the USEPA lifetime health advisory of 70 nanograms per liter (ng/L).				
	Expedited SI – November 2016-March 2017				
	The detection of PFAS in the on-base supply well prompted initiation of an expedited SI at OLF Coupeville and an off-base drinking water investigation near OLF Coupeville. PFAS were detected in 8 of the 27 well locations installed, and PFOA and/or perfluorooctane sulfonic acid (PFOS) exceeded the SL¹ and the USEPA lifetime health advisory of 70 ng/L for combined PFOA and PFOS at four locations (CH2M, 2018a).				
	Second Expedited SI – December 2017-September 2018				
Study Area Investigation History	A second expedited SI was conducted in 2017 and 2018 near the Keystone Well to evaluate PFAS concentrations near the Keystone Well and the potential impacts that increased pumping of the Keystone Well proposed by the Town of Coupeville could have on groundwater conditions at and nearby OLF Coupeville. The expedited SI included installation of two new monitoring well pairs, an aquifer test at the Keystone Well, and development of a groundwater flow model to evaluate aquifer test data (CH2M, 2018b). Groundwater samples indicated that concentrations of PFOA exceeded the SL at three of the four newly installed well locations. The aquifer test indicated that water levels in several groundwater monitoring wells at OLF Coupeville were observed to fluctuate in response to pumping at the Keystone Well. The groundwater modeling indicated that increased production of the Keystone Well would likely expand the Keystone Well capture zone to cover much of the western portion of OLF Coupeville including several wells with PFAS concentrations exceeding the USEPA lifetime health advisory.				
	Preliminary Assessment – November 2018				
	A Preliminary Assessment (PA) for PFAS at OLF Coupeville, completed in November 2018, identified Building 2709 and Facilities 1, 2, and 11 as potential PFAS release areas (CH2M, 2018c).				

SAP Worksheet #10—Conceptual Site Model (continued)

Table 10-1. OLF Coupeville Conceptual Site Model

Supplemental SI - August 2019-May 2020

A Supplemental SI was conducted from August 2019 to May 2020 (CH2M, 2021a, forthcoming). Soil sampling confirmed the presence of PFAS release areas at Buildings 2709 and Facilities 1, 2, and 11. Groundwater samples indicated that concentrations of PFOS or PFOA exceeded the SLs at six new well locations, two near the release areas and four to the northwest, west and southwest of runway. Groundwater flow directions at OLF Coupeville suggest that PFAS detected in on-base monitoring wells southwest of Building 2709 and Facilities 1, 2, and 11 emanated from these release areas. Groundwater flow directions and elevated PFAS concentrations in monitoring wells near the northwestern end of the runway are likely associated with an additional, unidentified release area west of the runway, which has not yet been confirmed. Data collected during the Supplemental SI were used to update and refine the 2018 groundwater flow model and perform solute transport modeling.

Study Area Investigation History (continued)

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The Supplemental SI report (CH2M, 2021a) recommended additional investigation during the RI to evaluate the presence of an additional release area west of the runway, delineate the release areas associated with Building 2709 and Facilities 1, 2, and 11 (east of the runway), and investigate the connection between the on-base PFAS impacts with the elevated PFAS concentrations detected at off-base residential wells south of OLF Coupeville.

Off-base Drinking Water Investigation – November 2016-September 2020

From November 2016 through October 2017, the Navy sampled 112 drinking water wells near OLF Coupeville, including the Town of Coupeville's Keystone Well. Eight residential drinking water wells to the south of OLF Coupeville were found to have PFOA concentrations above the USEPA lifetime health advisory (**Figure 10-3**) (CH2M, 2018c, 2019a, 2020b). Biannual sampling of drinking water wells where PFAS were detected and drinking water wells adjacent to properties with PFAS exceedances of the USEPA lifetime health advisory was initiated by the Navy in 2017 (CH2M, 2021b; 2020b).

Release areas for on-base impacts to groundwater and off-base PFAS impacts to drinking water to be considered in this investigation are on-base locations of suspected releases of aqueous film-forming foam (AFFF) (referred to as potential source areas in the PA). At OLF Coupeville they include the following:

- Building 2709 (Crash Truck Shelter) fire trucks containing AFFF are stored and truck washing occurs; off-truck AFFF was stored at this location in the past.
- Facilities 1, 2, and 11 (Control Tower, Airfield Operations Building, and Potable Water Well Pump House) – PFAS have been detected in groundwater downgradient of these locations.

Release Areas

Groundwater and soil sampling results from the 2019-2020 Supplemental SI (CH2M, 2021a) are consistent with the identification of these facilities as potential release areas.

PFAS have been detected in groundwater samples collected from on-base wells, with concentrations exceeding the screening level (SL)¹ of 40 nanograms per liter (ng/L) for PFOA at 11 locations and concentrations exceeding the SL of 40 ng/L for PFOS at three locations (see Nature and Extent section below) (CH2M, 2018b, 2021a).

During the 2017-2018 Expedited SI (CH2M, 2018b) and 2019-2020 Supplemental SI (CH2M, 2021a, forthcoming), PFOA and PFOS were detected in groundwater collected from monitoring wells west of the runway. Based on data from the 2019-2020 Supplemental SI, these detections are suspected to be related to a separate release area and not to either of the release areas identified in the PA.

Table 10-1. OLF Coupeville Conceptual Site Model

		conceptual site Woder
Site	Physical Characteristics	OLF Coupeville is located on a broad plateau of Smith Prairie in central Whidbey Island at an elevation of approximately 195 feet North American Vertical Datum of 1988 (NAVD88). The paved runway is approximately 5,400 feet long and is bordered by grass maintained by mowing operations extending to the public roads (Navy, 1994). A runway safety area extends approximately 3,300 feet south of the runway footprint and is bordered by trees and residential parcels (Figure 10-2).
	Geology and Hydrogeology	Whidbey Island lies within the Puget Lowland, a topographic and structural depression between the Olympic Mountains and the Cascade Range. The geology of the area is heavily influenced by glacial advances and retreats. The geologic units on Whidbey Island thus consist of a sequence of Quaternary age (less than 2 million years old) glacial and interglacial deposits that may be more than 3,000 feet thick (USGS, 1982). The near-surface deposits are mostly glacial sediment of the most recent Fraser glaciation (10,000 to 20,000 years old). The glacial and post-glacial sediments make up most of the overburden units underlying the Base.
		Lithology observed in soil borings is consistent with the previous mapping by Polenz et al. (2005). Surficial geology at OLF Coupeville consists of the Partridge Gravel, which was deposited by glacial meltwaters and is composed of sand, gravel, and sand-gravel mixtures with minor interlayered silt and silty sand. Based on soil borings completed in 2017, the Partridge Gravel generally extends to depths of 180 to 200 feet below ground surface (bgs) at OLF Coupeville and is characterized by fine to medium sand with intermittent occurrences of gravel and laterally discontinuous layers of silt and clay, some of which can be up to 30 feet thick (CH2M, 2018a). Pleistocene deposits, including Vashon till, lie beneath the Partridge Gravel. In the vicinity of OLF Coupeville, these deposits consist of heterogeneous clay, claystone, and silt, and frequently contain organic material, such as plant material and peat.
		The subsurface lithology and hydrogeology at OLF Coupeville are depicted in cross section on Figure 10-4 (cross section locations) and Figures 10-5 through 10-10 (cross sections A-A' through F-F').
		The unconfined groundwater table at OLF Coupeville generally occurs within the Partridge Gravel between 120 and 140 feet bgs (approximately 50 to 70 feet NAVD88). Perched groundwater may be present above the water table controlled by local occurrences of low-permeability silt and clay layers. With depth, localized layers of silt and clay may promote semiconfined to confined aquifer conditions within the Partridge Gravel. Many local water supply wells are screened in the lower portion of the Partridge Gravel. These wells are typically screened below 150 feet bgs in transmissive sand and gravel.
		The April 2017 groundwater elevation study of 27 groundwater monitoring wells located within the OLF Coupeville boundary (CH2M, 2018a) indicated groundwater elevation fluctuations of up to 0.6 foot over a 48-hour monitoring period. The groundwater monitoring wells for this study were screened within three general elevation intervals, which were categorized based on their elevation relative to mean sea level: shallow (screened above 50 feet NAVD88), intermediate (screened 0 to 50 feet NAVD88), and deep (screened near or below sea level NAVD88). The shallow, intermediate, and deep elevation zone designations do not indicate three discrete aquifers or water-bearing zones. Rather, with the exception of some shallow wells possibly screened within localized areas of perched groundwater, the shallow, intermediate, and deep elevation zones are located within the single aquifer system within which most local water supply wells, including the Keystone Well (screened from 142 to 182 feet bgs), are screened.

Table 10-1. OLF Coupeville Conceptual Site Model

Table 10-1. OL	Table 10-1. OLF Coupeville Conceptual Site Model				
ology		The initial aquifer test for the Keystone Well had a yield of 302 gallons per minute with approximately 20 feet of drawdown over a 24-hour period and a transmissivity of 79,000 gallons per day per foot (Robinson Noble, Inc., 2008). During the 2017/2018 Keystone Well aquifer test, drawdown in response to pumping at the Keystone Well was observed at several on-base monitoring wells west of the OLF Coupeville Runway (CH2M, 2018b). Drawdown in response to pumping was not observed at monitoring wells south or east of WI-CV-MW14M. The current yield of the Keystone Well as operated by the Town of Coupeville is approximately 200 to 250 gallons per minute over a 21- to 23-hour period each day.			
	eology	Hydraulic heads near the western boundary of the Base (WI-CV-MW14M, WI-CV-MW15S/M, and WI-CV-MW16S/M) are somewhat depressed by pumping at the Keystone Well, which has induced a westward gradient in that part of the Base toward the Keystone Well. Groundwater flow conditions prior to the installation of the Keystone Well are not well understood, but it is likely that without the hydraulic stress from the pumping of the Keystone Well, groundwater would be expected to migrate to the south without a westerly component.			
Site Conditions (continued)	Conditions	Groundwater levels measured in groundwater monitoring wells screened in the shallow zone are encountered between 90 and 130 feet bgs, which may support the interpretation that some are screened in perched conditions that are laterally discontinuous across the Base. Static water levels in wells screened in the intermediate zone indicate semiconfined conditions, with hydrostatic heads rising 30 to 40 feet above the base of the silt/clay aquitard (where present). Groundwater elevation data and groundwater modeling studies indicate the presence of a groundwater mound (divide) centered in the northern portion of OLF Coupeville. This interpretation is supported by the Island County Water Resource Management Plan (Island County, 2005), which suggests that OLF Coupeville is located on a hydrogeologic divide, and groundwater is likely to be flowing radially away from OLF Coupeville. The dominant flow direction in the intermediate zone over the majority of OLF Coupeville is to the southwest, shifting to the south-southeast in the southern portion of the site. Groundwater flow in the deep zone is inferred to be predominantly to the south/southeast. Vertical gradients on-base at OLF Coupeville are predominantly downward (CH2M, 2018a, 2018b).			
		Groundwater contour maps have been generated for intermediate- and deep-screened monitoring well networks, included on Figures 10-11 and 10-12 , respectively. Groundwater levels measured in shallow-screened wells are highly variable, suggesting some wells are screened in perched conditions. As such, a groundwater contour map has not been developed for the shallow wells. Groundwater elevation data as well as groundwater modeling results (CH2M, 2018b, forthcoming) indicate the presence of a groundwater mound (divide) northeast of OLF Coupeville.			
Chemicals of Potential Cond	cern	18 PFAS (listed in Worksheet #15)			
		Groundwater			
Nature and Extent		PFOA concentrations are present above the SL in shallow zone groundwater at the release areas near Building 2709 and Facilities 1, 2, and 11 (Figure 10-14) ² and are attributed to leaching of PFAS from vadose zone soil in these areas. Downgradient of the release areas, PFOA is present above the SL in groundwater in the shallow and intermediate aquifer zones (Figure 10-14 and 10-15), including the base supply well adjacent to Building 2807, and present at concentrations below the SL in the deep aquifer zone (Figure 10-16). An inferred southwesterly groundwater flow direction would suggest PFAS emanated from the shallow release areas near Building 2709 and Facilities 1, 2, and 11 and migrated vertically deeper into the aquifer system as groundwater flowed downgradient to the south; this flow scenario provides evidence for potential off-base migration affecting off-base drinking water. However, there are data gaps as to where the highest concentrations of PFOA are located as the plume migrates downgradient. For instance, the PFOA concentration was lower than the SL at WI-CV-MW26D (Figure 10-16), the monitoring well closest to the southern site boundary in a downgradient direction from the release areas, yet several deep aquifer off-base residential wells exceed the USEPA lifetime health advisory further downgradient.			

Table 10-1. OLF Coupeville Conceptual Site Model

Low-level to non-detect PFOA concentrations within the shallow, intermediate, and deep aquifer zones to the north, east, and directly west of the release areas (**Figures 10-14** through **10-16**) indicate that the extent of impacts is defined in these directions, but the extent of impacts downgradient to the south and southwest is not yet defined.

PFOA is present at concentrations above the SL in groundwater in the intermediate aquifer zone west of the runway near the Keystone Well (**Figures 10-14** and **10-15**). These concentrations appear to represent a separate release area near the northwestern end of the runway that may also be the source of PFOA in the Keystone Well. Because there is limited PFOA data from the shallow and deep aquifer zones in this area (near and downgradient of the WI-CV-MW15 and WI-CV-MW16 well pairs), and limited PFOA data in general to the west and south, the horizontal and vertical extent of PFOA impacts associated with this release area is not yet defined.

PFOS concentrations are present above the SL in shallow groundwater at the release areas near Building 2709 and Facilities 1, 2, and 11 (Figure 10-17) and are attributed to leaching of PFAS from vadose zone soil in these areas. PFOS is not present above the SL in any of the three aquifer zones downgradient of these release areas (Figures 10-17 through 10-19). This suggests that the extent of PFOS impacts associated with these release areas may be limited to the shallow aquifer zone adjacent to the release areas, but additional delineation to the south and east will be necessary to confirm this extent.

PFOS is also present above the SL in the shallow aquifer zone at WI-CV-MW23S (downgradient of the WI-CV-MW15 and WI-CV-MW16 well pairs; see **Figures 10-17** and **10-18**). While low concentrations in intermediate and deep aquifer zone groundwater downgradient of this well suggest PFOS impacts may be limited to the area around WI-CV-MW23S (**Figures 10-18** and **10-19**), the extent of PFOS in this area has not been fully delineated.

Soil

Nature and Extent (continued)

PFOA and PFOS are present in vadose zone soil near Building 2709 and Facilities 1, 2, and 11 at depths between 0 and 100 feet bgs (CH2M, 2021a). Soil sampling results are shown on **Figure 10-13**. The horizontal extent of PFOA and PFOS in soil at this area is not fully delineated. Further, there is inconsistency between the depths of the highest PFOA and PFOS concentrations in soil, generally observed at shallow depths well above the water table, and the elevated groundwater concentrations seen in the shallow aquifer at and immediately downgradient of these release areas. Soil concentrations would be expected to be highest in the deepest soil samples in the capillary fringe but were generally low or below DLs in samples collected at greater than 90 feet bgs. Soil samples have not been collected from the release area west of the runway near the Keystone Well; the horizontal and vertical extent of PFAS impacts, if any, to soil in this area is unknown.

Drinking Water

Off-Base Residential Wells:

PFOA has been detected at concentrations exceeding the USEPA lifetime health advisory in eight off-base residential wells since the sampling program began in November 2016 (**Figure 10-3**). Beginning in October 2017, these eight off-base residential wells have been sampled biannually for 14 PFAS. The list of PFAS was increased to 18 in October 2019. Results from the May 2020 sampling event indicate that PFOA continues to exceed the lifetime health advisory in all eight wells. PFOS was detected in five of the eight wells, and all concentrations were less than the USEPA lifetime health advisory.

While it is understood that off-base drinking water wells have been impacted by PFAS, the focus of this RI is on groundwater impacts at OLF Coupeville that may affect off-base properties, rather than off-base drinking water itself.

Keystone Well:

The Keystone Well is sampled biannually as part of the off-base drinking water sampling program. During the September 2018 sampling event, the concentration of PFOA exceeded the USEPA lifetime health advisory. These results are consistent with the independent testing results conducted by the Town of Coupeville in summer 2018, where PFOA was detected at concentrations exceeding the USEPA lifetime health advisory. With the exception of these 2018 sampling events, PFOA and PFOS concentrations have been less than the USEPA lifetime health advisory since sampling began in 2016. In May 2020, PFOS was detected in the Keystone Well at a concentration of 1.44 ng/L, below the USEPA lifetime health advisory.

Table 10-1. OLF Coupeville Conceptual Site Model

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Table 10 1. Oti coupevi	ille Conceptual Site Model					
	Because of their chemical structure, PFAS are chemically and biologically stable and resist typical degradation processes; therefore, they persist in the environment. Additionally, PFAS are watersoluble and migrate readily from soil to groundwater where they can be transported long distances (USEPA, 2014).					
	The PFAS PA (CH2M, 2018c) identified the following migration pathways:					
	Releases to surface or subsurface soil, on-base ditch surface soil, and surface water (if and where present) ³ .					
	Vertical migration or leaching of PFAS from vadose zone sources to the underlying groundwater system.					
Migration Pathways	Horizontal and vertical transport within the aquifer system via advection (with groundwater flow).					
	Preferential pathways via nongrouted well casings: The well construction records for the water supply wells at OLF Coupeville show that the steel well casings were not grouted deeper than 18 feet bgs, potentially leaving an open annulus from 18 feet bgs to the bottom of the borehole at 180 feet bgs. The absence of a grout seal could allow shallow groundwater, potentially containing PFAS, to migrate downward and reach the deeper aquifer used locally for water supply purposes.					
	Other potential PFAS migration pathways that may be evaluated further, include the following:					
	Overland flow of media containing PFAS to downgradient off-base surface water bodies ³ .					
	Soil					
	Workers, visitors, trespassers, residents, and recreators could potentially be exposed to PFAS in soil through incidental ingestion of and dermal contact with surface and/or subsurface soil. Construction or other ground disturbing activities, including landscaping, could result in potential generation of and receptor exposure to dust. There are currently no PFAS toxicity values available to evaluate inhalation exposures.					
	Potential recreational hunters could ingest game that has been exposed to PFAS in on-base soil.					
	Groundwater					
Human Health Receptors and	Current and future off-base residents: Ingestion of PFAS in groundwater used as a drinking water supply.					
Exposure Scenarios	Current and future on-base drinking water users (visitors and workers) at OLF Coupeville: Ingestion of PFAS in groundwater.					
	Potential off-base recreational hunters and anglers could ingest game or fish that has been exposed to PFAS in surface water ³ off-base and downgradient of site. Human receptors could also consume crops watered with PFAS-containing water or livestock and poultry that have been exposed to PFAS in surface water.					
	Surface Water and Ditch Surface Soil					
	Workers, visitors, trespassers, residents, and recreators could be exposed to PFAS through incidental ingestion of surface water (if present) and ditch surface soil, and dermal contact with ditch surface soil ³ .					
Ecological Receptors and Exposure Scenarios	PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for both the Human Health and Ecological Risk Assessments. Prior to conducting an ecological risk screening, the ecological screening values that will be used for comparison to site media will be reviewed based on current science and relevancy to site-specific receptors (e.g., relevant ecological receptors present). Ecological screening values selected for the screening will be presented in a technical memorandum or other documented form for regulatory review and acceptance.					

Table 10-1. OLF Coupevi	ille Conceptual Site Model					
	Soil					
	Lower trophic level terrestrial ecological receptors (such as terrestrial plants and soil invertebrates) could be exposed to PFAS released to surface soils through root uptake, direct contact, and/or direct ingestion in this area. Because there is evidence that PFAS bioaccumulate in terrestrial food items (such as plants), there is the potential that upper trophic level receptors (such as birds and mammals) could be exposed to these substances via the food web, as well as through incidental ingestion of soil.					
	Groundwater					
Ecological Receptors and Exposure Scenarios (continued)	While ecological receptors generally do not have direct exposure to groundwater, shallow groundwater at OLF may be hydraulically connected to nearby downgradient surface water bodies to the south, potentially including Crocket Lake and Admiralty Bay, presenting a potential exposure pathway for benthic ecological ⁴ receptors at the groundwater-to-surface-water interface within the biologically active zones.					
(continued)	Surface Water, Sediment, and Ditch Surface Soil					
	Lower trophic level ecological receptors (such as plants; aquatic, benthic, or terrestrial invertebrates; and reptiles or amphibians) could be exposed to PFAS released to ditch surface soil, surface water (if present), or sediment (either directly or indirectly via surface runoff and drainage from onsite terrestrial areas or through groundwater discharge), through root uptake, direct contact, or direct ingestion. Because there is evidence that PFAS bioaccumulate in food items, there is the potential that upper trophic level receptors (such as birds and mammals) could be exposed to these compounds via the food web, as well as through incidental ingestion of ditch surface soil and, if the pathway is complete, offsite sediment, and direct ingestion of fresh surface water (if present) ³ .					
	The following data needs were identified based on the previous investigations summarized in Table 10-1 :					
	 Additional groundwater and soil concentration data are needed to further delineate the horizontal and vertical extent of PFAS in the vadose zone at the confirmed PFAS release areas at Building 2709 and Facilities 1, 2, and 11 and assess the risk to human health and ecological⁴ receptors. 					
Data Needs	 Additional groundwater and soil concentration data are needed to confirm the presence of a suspected PFAS release area west of the runway, and if identified, define the nature and extent of this release area, and assess risk to human health and ecological⁴ receptors. Determination of the nature and extent of PFAS impacts in this area may require additional sampling as part of future RI phases which have yet to be scoped. 					
	 Additional soil and groundwater concentration data are needed as follows: 1) to address data discrepancies between the distribution of PFAS in on-base groundwater and elevated PFAS concentrations in off-base residential wells south of OLF Coupeville, and 2) to refine the existing information about PFAS migration directions from on-base release areas and impacted on-base groundwater monitoring wells to off-base drinking water wells with exceedances of the USEPA lifetime health advisory for PFOA and PFOS. 					
	Additional data are needed to assess temporal variability in PFAS concentrations in groundwater at OLF Coupeville.					

SLs referred to in this SAP are the current USEPA tap water and soil regional screening levels (RSLs) for PFBS (based on a hazard quotient [HQ] of 0.1) and tap water and soil SLs for PFOS and PFOA as described in the Assistant Secretary of Defense Memorandum from October 15, 2019 (ASD, 2021) based on an HQ of 0.1.

define the nature and extent of PFAS and assess risk to ecological receptors4.

Concentration data for surface water (if present) and drainage ditch surface soil³ are needed to

- PFOA and PFOS concentrations presented on Figures 10-14 through 10-19 present data from two types of groundwater samples collected from the sampling locations: depth-discrete groundwater grab samples collected from the soil borings at (multiple depths within each boring) during drilling, and low-flow groundwater samples collected from the completed monitoring wells. Refer to note 5 on Figures 10-14 through 10-19.
- ³ If the extent of PFOA, PFOS, and PFBS extends to areas that are affected by non-Navy sources, additional consideration may be warranted.
- ⁴ PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for Ecological Risk Assessment.

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Problem Statement and Objectives

The objectives, environmental questions, investigation approach, and project quality objectives are presented in Table 11-1. Figure 11-1 shows the decision logic associated with Table 11-1. The investigation approach presented in Table 11-1 applies only to Phase 1 of the RI which includes on-base work only. The scoped off-Base phase of the RI will be described in a future addendum to this SAP. Any additional phases needed to accomplish the objectives of the RI will be described under future task order work plan documents.

Table 11-1. Project Quality Objectives/Systematic Planning Process Statements

Problem Definition/Objectives Environmenta Question(s)		General Investigation Approach	PQOs	
PFOA and PFOS have been detected in vadose zone soils at the on-base potential release areas identified in the PA; however, the extent of the release areas is not fully delineated, and there are data gaps in PFAS groundwater concentration data between the release areas and downgradient on-base well locations. The objective of this step is to further define the horizontal and vertical extent of PFAS in soil and groundwater above PFAS PALs at the on-base release areas (Building 2709 and Facilities 1, 2, and 11) and between the release areas and downgradient well locations and determine whether there is unacceptable risk to human health and ecological receptors.	What is the horizontal and vertical extent of PFAS in soil and groundwater at the Building 2709 and Facilities 1, 2, and 11 release areas and between the release areas and downgradient well locations, and are PFAS present at concentrations that pose unacceptable risk to human health and ecological¹ receptors?	Soil borings will be advanced at two locations near Building 2709 and two locations near Facilities 1, 2, and 11 (total of four locations; see Figure 11-2). Soil borings will be advanced to the top of the water table (approximately 100 feet bgs). Up to five soil samples will be collected at each of the soil borings (WI-CV-SO11 through WI-CV-SO14). Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air-water and lithologic interfaces. A shallow-zone monitoring well (WI-CV-MW34S) will be drilled (with no soil samples collected) and installed east of WI-CV-MW20S to provide information about PFAS concentrations at the Base boundary east of the release areas and near the base's eastern property boundary. A monitoring well pair (WI-CV-MW35S/M) will be drilled (with no soil sampling) and installed southwest of WI-CV-MW02S/M (Figure 11-2) and the Building 2709 release areas. The two groundwater monitoring wells will target the shallow and intermediate zones. During drilling of the intermediate soil boring, which will be drilled prior to the shallow boring, up to three depth-discrete groundwater grab samples will be collected between the water table and approximately 200 feet bgs. The analytical results from the grab samples will be evaluated and used to determine screen placement settings for both the intermediate and the shallow groundwater monitoring wells. Groundwater samples will be collected from completed groundwater monitoring wells following installation. The samples and rationale are outlined in Worksheet #17. Soil and groundwater (grab and monitoring well) samples will be submitted to Battelle for analysis of the 18 PFAS listed in USEPA Method 537.1. Groundwater grab samples will be analyzed on a standard TAT.	Groundwater and soil data will be evaluated in accordance with Figure 11-1 .	

Table 11-1. Project Quality Objectives/Systematic Planning Process Statements

Problem Definition/Objectives	Environmental Question(s)	General Investigation Approach	PQOs
PFOA and PFOS have been detected in groundwater monitoring wells west and northwest of the runway near the Town of Coupeville's Keystone Well at concentrations above the PALs for PFOA and PFOS (Figures 10-14 through 10-19). PFOA exceeded the USEPA lifetime health advisory in the Keystone Well during the September 2018 biannual drinking water sampling, and concentrations generally range from 55 to 71 ng/L. PFOS was detected above the SL at WI-CV-MW23S. Based on the results of previous basewide monitoring, well sampling, and groundwater modeling, these concentrations are not believed to be related to the release areas at Building 2709 and Facilities 1, 2, and 11 (CH2M, 2021a), and the area west of the runway near the Keystone Well was not identified in the PA as a potential PFAS release area; therefore, the source of PFAS detected in groundwater in the area northwest of the runway is uncertain. The objective of this step is define the nature and extent of this release area above PFAS PALs and determine whether there is unacceptable risk to human health and ecological¹ receptors.	What is the horizontal and vertical extent of PFAS in the soil and groundwater, and are PFAS present at concentrations that pose unacceptable risk to human health or ecological¹ receptors?	Soil borings will be advanced at four locations near groundwater monitoring wells where PFAS has been detected at concentrations above groundwater PALs (Figure 11-2, Figures 10-14 through 10-19). Soil borings will be advanced to the top of the water table (approximately 100 feet bgs). Up to five soil samples will be collected at each of the soil borings (SOO7 through SO10). Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air-water and lithologic interfaces. Two monitoring well pairs (WI-CV-MW32S/M and WI-CV-MW33S/M) will be drilled and installed in the area west of the runway (Figure 11-2). These groundwater monitoring wells will target the shallow and intermediate zones at each location. Up to five soil samples will be collected at well pair WI-CV-MW33S/M during advancement of the shallow boring to evaluate this as a release area associated with the runway. Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air-water and lithologic interfaces. During the drilling of the intermediate borings at WI-CV-MW32S/M and WI-CV-MW33S/M, which will be drilled prior to the shallow borings, up to three depth-discrete groundwater grab samples will be collected. The analytical results from the grab samples will be evaluated and used to determine screen placement settings for both the intermediate and the shallow monitoring wells at each location. Groundwater samples will be collected from completed groundwater monitoring wells following installation. The samples and rationale are outlined in Worksheet #17. Soil and groundwater (grab and monitoring well) samples will be submitted to Battelle for analysis of the 18 PFAS listed in USEPA Method 537.1. Groundwater grab samples and soil samples from select locations (SO08 through SO10) will be analyzed on an expedited (72-hour) TAT.	Groundwater and soil data will be evaluated in accordance with Figure 11-1.

Table 11-1. Project Quality Objectives/Systematic Planning Process Statements

Problem Definition Objectives Environmental Question(s)		General Investigation Approach	PQOs	
Low concentrations of PFOA/PFOS near the southern Base boundary are not consistent with higher concentrations of PFOA/PFOS in off-base residential wells south of OLF Coupeville. The objective of this step is to refine the data representative of groundwater flow and potential PFAS migration from the known on-base PFAS release areas and the release area west of the runway to impacted off-base residential drinking water wells with PFAS concentrations above the USEPA lifetime health advisory for PFOA and PFOS.	How are on-base concentrations of PFAS in the known release areas (adjacent to Building 2709 and Facilities 1, 2, and 11) and the release area west of the runway related to PFAS concentrations above the USEPA lifetime health advisory at offbase residential wells?	Two monitoring well pairs (WI-CV-MW36M/D and WI-CV-MW37M/D) will be drilled (with no soil sampling) and installed near the southwestern Base boundary (Figure 11-2). The groundwater monitoring wells will target the intermediate and deep zones at each location. During the drilling of the deep borings, which will be drilled prior to the intermediate borings, up to four depth-discrete groundwater grab samples will be collected. The analytical results will be evaluated and used to determine screen placement settings for both the deep and intermediate monitoring well at each location. Groundwater samples will be collected from completed groundwater monitoring wells following installation. The samples and rationale are outlined in Worksheet #17. Groundwater grab samples and monitoring well samples will be submitted to Battelle for analysis of the 18 PFAS listed in USEPA Method 537.1. Groundwater grab samples will be analyzed on an expedited TAT so that data can be evaluated for screen placement. Monitoring well samples will be analyzed on a standard TAT.	The groundwater data will be evaluated in accordance with Figure 11-1 and will be used to refine the CSM with respect to the distribution of PFAS in groundwater on base and migration pathways on the western side of the runway relative to impacted off-base residential wells.	
Multiple groundwater and drinking water sampling events have been performed at or near OLF Coupeville beginning in 2016; however, PFAS concentrations in groundwater may change over time. The objective of this step is to refine the data demonstrating the temporal variability in PFAS concentrations in groundwater at OLF Coupeville.	Have PFAS concentrations in groundwater changed over time at OLF Coupeville?	In addition to the groundwater sampling to be conducted at all new monitoring wells, groundwater sampling will be conducted at all previously existing monitoring wells at OLF Coupeville. Samples will be submitted to Battelle for analysis of the 18 PFAS listed in USEPA Method 537.1 at standard TAT.	Groundwater sampling results from existing wells will be appended to the existing data. If PFAS concentrations are consistent with previous sampling results, this will indicate that no temporal change in concentration has occurred. If PFAS concentrations in existing wells are inconsistent with previous sampling results, the results will be evaluated in accordance with Figure 11-1 to determine whether the inconsistency is significant enough to warrant a change to the CSM.	

Table 11-1. Project Quality Objectives/Systematic Planning Process Statements

Problem Definition Objectives	Environmental Question(s)	General Investigation Approach	PQOs
It is unknown whether PFAS releases have occurred to the drainage ditch system at OLF Coupeville. The objective of this step is to determine whether PFAS are present in ditch surface soil and surface water (if present), define the nature and extent of PFAS in ditch surface soil and surface water (if present) above PFAS PALS, and determine whether there is unacceptable risk to human health and ecological ¹ receptors.	Question(s) Are PFAS present in ditch surface soil or surface water (if present) at OLF Coupeville above the PALs? If yes, what are is the nature and extent of PFAS in ditch surface soil and surface water (if present) at OLF Coupeville? Are concentrations indicative of risks to human health or ecological ¹	The approach to investigating PFAS in ditch surface soil and surface water (if present) will be to collect paired ditch surface soil and surface water samples (where possible) from four on-base locations in drainage ditches along the runway (Figure 11-2). Ditch surface soil and surface water samples will be submitted to Battelle for analysis of the 18 PFAS listed in USEPA Method 537.1 at standard TAT. The samples and rationale are outlined in Worksheet #17.	The surface water data (if it can be collected) and ditch surface soil data will be evaluated in accordance with Figure 11-1.
	receptors?		

PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for both the Human Health and Ecological Risk Assessments. Prior to conducting an ecological risk screening, the ecological screening values that will be used for comparison to site media will be reviewed based on current science and relevancy to site-specific receptors (e.g., relevant ecological receptors present). Ecological screening values selected for the screening will be presented in a technical memo or other documented form for regulatory review and acceptance.

Problem Definition, Environmental Questions, and Project Quality Objectives

The problem definition, environmental questions, general investigation approaches, and PQOs contained in this SAP are described in **Table 11-1** and are based on the USEPA *Guidance on Systematic Planning Using the Data Quality Objectives Process* (USEPA, 2006) and its seven-step process. The detailed sampling approach, including numbers of samples, is provided in **Worksheet #17**. **Figure 11-1** shows the decision logic that is associated with **Table 11-1**. Planned sample locations are shown on **Figure 11-2**.

What Are the Project Action Limits?

PALs are media-specific standards and criteria chosen for evaluation to help provide a conservative assessment of site conditions and determine if further evaluation or action is needed to address concentrations of chemicals present onsite. The PALs for data collected at OLF Coupeville are presented in **Worksheet #15** and are summarized as follows:

Groundwater and Surface Water

Groundwater and surface water (if present) data will be screened against the SLs based on a hazard quotient (HQ) of 0.1 for PFOA (40 ng/L), PFOS (40 ng/L), and perfluorobutane sulfonate (PFBS) (600 ng/L) (ASD, 2021; USEPA, 2021). There are no PALs for other PFAS analytes.

Soil

Soil data will be screened against residential soil SLs based on an HQ of 0.1 for PFOA (130 micrograms per kilogram [μ g/kg]), PFOS (130 μ g/kg), and PFBS (1,900 μ g/kg) (ASD, 2021; USEPA, 2021). There are no PALs for other PFAS analytes.

Soil data will also be evaluated for leaching potential based on a multiple lines of evidence approach including assessment of groundwater concentrations, consideration of soil type and PFAS distribution in the vadose zone, solute transport modeling, and state-of-the-science research on PFAS leaching. While preliminary screening against soil-to-groundwater SLs will be used as a part of the multiple lines of evidence approach to assess leaching potential, soil-to-groundwater SLs are not considered PALs for this project and are not considered clean up goals or intended for use in remedial action or risk assessment decision making.

For What Will the Data Be Used?

Data will be used to address the environmental questions and PQOs listed in **Table 11-1**. Ongoing sampling and other activities being done to address off-base drinking water that exceeds the USEPA lifetime health advisory will not be conducted as a part of RI activities at OLF Coupeville.

There are uncertainties to be considered with regard to data use. Scientific research and regulatory guidelines related to PFAS are rapidly evolving. As such, the information provided in this SAP presents the state of the science at the time of issuance of this SAP. The Navy will re-evaluate changing science and regulations at the time of reporting to ensure that data evaluation and risk assessments presented in this report reflect any changes to toxicology information, regulatory standards, and DoD and Navy policy and guidance. Any changes to the proposed data evaluation and risk assessment approaches will be discussed with stakeholders prior to issuance of the report and any revisions.

What Types of Data Are Needed?

The types of data needed include the following:

- Subsurface lithology of the soil borings to determine where semi-confining aquitards and aquifers are located; this information will affect monitoring well installations and provide input for updates to the CSM and the numerical groundwater flow model.
- Synoptic groundwater level surveys from on-base wells to evaluate groundwater flow and solute transport directions, and for use as numerical groundwater flow model calibration targets (to be collected following installation of off-base wells in Phase 2 of RI, not included in this SAP).
- Soil samples from release areas will be submitted for standard turnaround time (TAT) PFAS analysis via LC/MS/MS in compliance with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which the lab is accredited at the time of analysis to help inform the spatial and vertical distribution of PFAS in the vadose zone, and to assess whether soil source zones represent a potential continuing threat to underlying groundwater quality.
- Ditch surface soil and surface water samples will be submitted for standard TAT PFAS analysis via LC/MS/MS in compliance with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which the lab is accredited at the time of analysis to determine whether PFAS are present at concentrations warranting further investigation, and if present above PALs, determine the horizontal and vertical extent of PFAS in ditch surface soil and surface water.
- Field measurements of groundwater quality (pH, dissolved oxygen, temperature, conductivity, oxidation-reduction potential, and turbidity) will be completed during both vertical profile sampling and sampling of the monitoring well network.
- Depth-discrete groundwater samples will be submitted for 72-hour TAT PFAS analysis via LC/MS/MS in compliance with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which the lab is accredited at the time of analysis. Sample results will inform monitoring well construction and improve the team's understanding of the vertical distribution of PFAS in the groundwater system. Following drilling of the borehole, the drill casing will be left in place to maintain an open borehole pending the analytical results. The expedited TAT for groundwater analyses is intended to minimize the timeframe that drilling equipment is left in the ground prior to monitoring well construction.
- Contemporaneous groundwater sampling for PFAS will be used to estimate the horizontal and vertical extent
 of PFAS in groundwater at OLF Coupeville. Groundwater samples will be analyzed for PFAS via LC/MS/MS in
 compliance with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which the lab is
 accredited at the time of analysis. Analysis of groundwater monitoring well samples will be performed under
 standard laboratory TAT.
- Numerical three-dimensional groundwater flow modeling and solute transport modeling will be used to improve the understanding of past and potential future PFAS transport at OLF Coupeville.

Samples will be collected and analyzed as described in **Worksheet #17** to meet the project objectives. The field methodology is included in **Worksheet #14**. Justification for individual sample locations is provided in **Worksheet #17**. The specific target analytes and PALs are included in **Worksheet #15**.

Are There Special Data Quality Needs, Field or Laboratory, to Support Environmental Decisions?

Offsite laboratory analytical data will be of the quantity and quality necessary to provide technically sound and defensible assessments with respect to the aforementioned project objectives. Laboratory DLs will be suitable for detecting PFOA, PFOS, and PFBS at or below the PALs in accordance with **Worksheet #15**. QC sample requirements are detailed in **Worksheet #20**. For action decisions, the laboratory will follow the Measurement Performance Criteria (MPC) in **Worksheets #24** and **#28** for laboratory QC samples. These MPC are consistent with the DoD QSM (DoD, 2019a) as applicable and laboratory in-house limits where the QSM does not apply.

Where, When, and How Should the Data Be Collected and Generated?

Sample locations are shown on **Figure 11-2**. Field activities will be conducted in accordance with **Worksheets #14**, **#17**, **and #18**, and the project schedule outlined in **Worksheet #16**. The data will be collected following the standard operating procedures (SOPs) presented in **Worksheet #21**.

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SAP Worksheet #12-1—Measurement Performance Criteria Table – Field QC Samples

Matrix: Groundwater/Surface Water

Analytical Group: PFAS

Analytical Method: LC/MS/MS Compliant with QSM 5.3¹ Table B-15

QC Sample	Analytical Group	Frequency	Data Quality Indicators	Measurement Performance Criteria
Matrix Spike (MS)/Matrix Spike Duplicate (MSD)		One per 20 samples	Accuracy/Precision	See Worksheet #28.
Field Duplicate (FD)		One per 10 samples	Precision	Relative percent difference (RPD) less than (<) 25%
Equipment Rinsate Blank	PFAS	One per site per day of sampling for decontaminated equipment	Contamination	No analytes detected greater than (>) ½ LOQ or > 1/10 sample concentration, whichever is greater
Field Blank		One per site	Bias/Contamination	No analytes detected > ½ LOQ or > 1/10 sample concentration, whichever is greater
Cooler Temperature Indicator		One per cooler	Accuracy/Representativeness	Temperature less than or equal to (≤) 10°C, storage in the laboratory ≤ 6°C, not frozen

¹ QSM 5.3 or the latest version of the QSM for which the laboratory is certified at the time of sampling

Notes:

°C = degrees Celsius

% = percent

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${\sf SAP\ Worksheet\ \#12-2-Measurement\ Performance\ Criteria\ Table-Field\ QC\ Samples}$

Matrix: Surface Soil/Ditch Surface Soil, Soil

Analytical Group: PFAS

Analytical Method: LC/MS/MS Compliant with QSM 5.3¹ Table B-15

QC Sample	Analytical Group	Frequency	Data Quality Indicators	Measurement Performance Criteria
MS/MSD		One per 20 samples	Accuracy/Precision	See Worksheet #28.
FD	PFAS	One per 10 samples	Precision	RPD < 35%
Equipment Rinsate Blank		One per day of field sampling for decontaminated equipment	Bias/Contamination	No analytes detected > ½ LOQ or > 1/10 sample concentration, whichever is greater
Field Blank		One per site	Bias/Contamination	No analytes detected > ½ LOQ, or > 1/10 sample concentration, whichever is greater
Cooler Temperature Indicator		One per cooler	Accuracy/Representativeness	Temperature ≤ 10°C, storage in the laboratory ≤ 6°C, not frozen

¹ QSM 5.3 or the latest version of the QSM for which the laboratory is certified at the time of sampling.

SAP Worksheet #13—Secondary Data Criteria and Limitations Table

Secondary Data	Data Source (originating organization, report title and date)	Data Generator(s) (originating organization, data types, data generation/ collection dates)	How Data Will Be Used	Limitations on Data Use	
Groundwater elevation and analytical data and geology data from monitoring and base supply wells within the OLF Coupeville	CH2M. 2018a. Final Technical Memorandum, Evaluation of Per- and Polyfluoroalkyl Substances in Groundwater Outlying Landing Field Coupeville. Naval Air Station Whidbey Island Coupeville, Washington. May 2018.	CH2M. Groundwater and geology. February and March 2017.	Data will be used to assist the placement of groundwater monitoring wells, soil borings, and vertical profiling locations and for human health and ecological ¹ risk assessments	None	
Groundwater elevation and analytical data and	CH2M. 2018b. Aquifer Test, Groundwater Sampling, and Drinking Water Sampling Data Evaluation and Groundwater Modeling Report, Per- and Polyfluoroalkyl Substances (PFAS), Outlying Landing Field Coupeville, Naval Air Station Whidbey Island Oak Harbor and Coupeville, Washington. September 2018.	Campling, and Drinking Water Sampling Data Evaluation and Groundwater Modeling Report, Per- and Polyfluoroalkyl Substances (PFAS), Outlying Landing Field Coupeville, Naval Air Station Whidbey Sland Oak Harbor and Coupeville,			
geology data from groundwater monitoring wells within the OLF Coupeville	CH2M. 2021a. Supplemental Site Inspection Report for Per- and Polyfluoroalkyl Substances Outlying Landing Field Coupeville, Naval Air Station Whidbey Island, Oak Harbor, Washington.	CH2M. Groundwater and geology. August through December 2019,	Data will be used to assist the placement of groundwater monitoring wells, soil borings,	None	
	CH2M. Forthcoming. Draft Supplemental Site Inspection Report Addendum for Perand Polyfluoroalkyl Substances Outlying Landing Field Coupeville, Naval Air Station Whidbey Island, Oak Harbor, Washington.	February through March and October 2020.	and vertical profiling locations and for human health and ecological ¹ risk assessments.	None	
Details regarding potential release areas of PFAS on-base at OLF Coupeville	CH2M. 2018c. Preliminary Assessment for Per- and Polyfluoroalkyl Substances (PFAS), Outlying Landing Field Coupeville, Naval Air Station Whidbey Island Oak Harbor and Coupeville, Washington. November.	CH2M. Geology, historical information through 2018.	Data will be used to assist the placement of groundwater monitoring wells, soil borings, and vertical profiling locations.	None	

SAP Worksheet #13—Secondary Data Criteria and Limitations Table (continued)

Secondary Data	Data Source (originating organization, report title and date)	Data Generator(s) (originating organization, data types, data generation/ collection dates)	How Data Will Be Used	Limitations on Data Use
Well construction and historical well testing data (pumping test results and water quality testing results) from the Keystone Well	Robinson Noble, Inc. 2008. Town of Coupeville Keystone Hill well Construction and Testing Report. Tacoma, Washington. April 2008.	Robinson Noble, Inc. Well, geology, and groundwater. January through March 2008.	Data will be used as input to the aquifer modeling software and assist in the placement of groundwater monitoring wells, and selection of monitoring well screen intervals.	None
Geohydrology data	USGS. 1982. Preliminary Survey of Ground-water Resources for Island County, Washington.	USGS. Geology. 1979 and 1980.	Data will be used as input to the aquifer modeling software and assist in the placement of groundwater monitoring wells, and selection of monitoring well screen intervals.	None
Off-base Monitoring Well data	Island County. 2016. Island County Hydrogeologic Database Well Search Utility Data. Data Generated on 11/4/2016.	Island County. Well. 1963 to 2016.	Data will be used for input to the groundwater modeling software.	None
Off-base drinking water results	CH2M. 2020b. Results of Investigation of Per- and Polyfluoroalkyl Substances in Off-Base Drinking Water—Ault Field, Area 6, and Outlying Landing Field Coupeville, Naval Air Station Whidbey Island, Washington. December 2020.	CH2M. Groundwater. October 2019 through September 2020.	Data will be used to assist in the selection of vertical profiling locations.	None

PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for both the Human Health and Ecological Risk Assessments..

SAP Worksheet #14—Summary of Project Tasks

All project tasks described in this SAP will be completed under the on-base Phase 1 of the RI. Applicable SOPs for project tasks outlined in this section are listed in **Worksheet #21** and provided in **Appendix B**.

Premobilization Tasks

- Work Plan development and approval (SAP, Accident Prevention Plan Site Safety and Health Plan, and Waste Management Plan Environmental Protection Plan)
- National Historic Preservation Act Section 106 Consultation with the State Historic Preservation Officer and/or the Advisory Council on Historic Preservation to identify possible conflicts between historic preservation objectives and the proposed activities in the clearance area shown on Figure 11-2
- Subcontractor procurement
 - Analytical laboratory
 - Data Validation
 - Utility locator
 - Driller
 - Surveyor
 - Investigation-derived waste (IDW) transportation and disposal contractor
- Fieldwork scheduling
- Coordination with NAS Whidbey Island for site access and IDW staging at OLF Coupeville

Mobilization

Mobilization for the field effort includes procurement of necessary field equipment and initial transport to the site. Equipment and supplies will be brought to the site when the CH2M field team mobilizes for field activities. Before beginning any work, CH2M and its subcontractors will have field meetings to discuss the work items and worker responsibilities, and to familiarize workers with the Accident Prevention Plan and Site Safety and Health Plan.

Utility Locating

Utilities will be cleared before beginning intrusive activities. CH2M will coordinate utility clearance. In addition, a third-party utility clearance subcontractor will be procured by CH2M to clearly mark the proposed groundwater monitoring well locations. Any proposed well locations within 5 feet of utility locations will be relocated to avoid impacts on utilities. If a monitoring well location needs to be relocated, the field team will consult with the CH2M PM and NAVFAC Northwest RPM to establish a new well location.

Soil Borings and Groundwater Monitoring Well Installation and Development

Eight on-base soil borings will be advanced with soil sampling (up to five soil samples collected at each location, with no groundwater sampling and no groundwater monitoring well installations). Target depth is the water table at each location. Soil sample depths will be determined in the field and will target lithologic transitions observed in the soil cores.

Eleven on-base soil borings (WI-CV-MW32 through WI-CV-MW37) will be advanced with groundwater monitoring wells installed. Groundwater vertical profile sampling will occur in three of the intermediate depth soil borings (WI-CV-MW32M, WI-CV-MW33M, and WI-CV-MW35M) and two of the deep soil borings (WI-CV-MW36D and W-CV-MW37D). Groundwater vertical profiling depths will be determined in the field and will target first-encountered groundwater and up to two additional depths separated by silt/clay layers. At pair WI-CV-

SAP Worksheet #14—Summary of Project Tasks (continued)

WI-CV-MW33S/M, up to five soil samples will be collected during the drilling of the shallow boring. Soil sample depths will be determined in the field and will target lithologic transitions observed in the soil cores.

Soil borings will be advanced via sonic drilling techniques, and groundwater monitoring wells will be installed and developed in accordance with the SOPs listed in **Worksheet #21** and provided in **Appendix B**.

Soil Logging

All soil borings will be logged for lithology and field-screened by a photoionization detector (PID) at every interval in accordance with the SOPs listed in **Worksheet #21** and provided in **Appendix B**.

Surveying

The newly installed groundwater monitoring wells and soil borings, and off-base well WI-CV-MW18M installed during the Supplemental SI, will be surveyed by a Washington-licensed surveyor in accordance with the SOPs listed in **Worksheet #21** and provided in **Appendix B**.

Sampling Tasks

Applicable field notes and forms will be filled out completely each day.

- Soil Sampling
 - Soil sampling will be completed in accordance with the SOPs listed in Worksheet #21 and provided in Appendix B.
 - Up to five soil samples will be collected from each of soil borings SO07 through SO14.
 - Soil samples from selected locations (WI-CV-SO08 through WI-CV-SO10) will be sent to Battelle for PFAS
 analysis with a 72-hour TAT. Remaining soil samples will be sent to Battelle for PFAS analysis at standard
 TAT.
- Groundwater Vertical Profile Sampling
 - Depth-discrete groundwater vertical profile sampling will be completed, with up to four groundwater samples collected from each of the following locations: WI-CV-MW32M, WI-CV-MW33M, WI-CV-MW35M, WI-CV-MW36D, and WI-CV-MW37D. All depth-discrete groundwater vertical profile sampling during the RI will be in accordance with the SOPs listed in Worksheet #21 and provided in Appendix B.
 - Groundwater grab samples will be sent to Battelle for PFAS analysis with a 72-hour TAT; groundwater profiling results will be used to determine target well settings for the paired wells.
- Groundwater Monitoring Well Sampling
 - Groundwater sampling will be completed at all new and existing groundwater monitoring wells in accordance with the SOPs listed in Worksheet #21 and provided in Appendix B.
 - Groundwater samples will be sent to Battelle for PFAS analysis on standard TAT.
- Surface Water and Ditch Surface Soil Sampling
 - Sampling of surface water (if present) and ditch surface soil is proposed at up to four locations. Surface water and ditch surface soil sampling will be conducted in accordance with the SOPs listed in Worksheet #21 and provided in Appendix B.

SAP Worksheet #14—Summary of Project Tasks (continued)

Synoptic Water Level Survey

Manual groundwater levels will be measured at all new and existing groundwater monitoring wells, in accordance with the SOPs listed in **Worksheet #21** and provided in **Appendix B.**

Decontamination

All drilling equipment used during groundwater monitoring well installation and reusable sampling equipment will be decontaminated immediately after each use in accordance with applicable SOPs referenced in **Worksheet #21** and provided in **Appendix B**. Sensitive instrumentation such as equipment used to collect water quality parameters will be decontaminated in accordance with the equipment manufacturer's guidelines.

IDW Management

IDW is expected to consist of drill cuttings from the soil borings generated during monitoring well installations, purge water from well development and groundwater sampling, and decontamination fluids. Aqueous IDW and solid IDW will be stored in separate roll-off containers, portable tanks, or drums. IDW will be managed in accordance with the Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC RPMs, November 2020 Update (NAVFAC, 2020) and in accordance with SOPs listed in **Worksheet #21** and provided in **Appendix B**. Solid IDW is expected to be non-hazardous. Aqueous IDW with concentrations exceeding 70 ng/L (combined PFOA and PFOS) will be treated to levels less than 70 ng/L prior to disposal. Incineration will only be conducted if other treatment options are determined to be infeasible. NAVFAC Headquarters approval must be obtained prior to incineration of any PFAS-containing waste. To minimize the volume of aqueous IDW requiring treatment, all efforts will be made to segregate release area IDW from that generated in downgradient areas where concentrations are likely to be lower.

Analyses and Testing Tasks

- Battelle will process and prepare soil samples (including ditch surface soil) for analysis and analyze samples in accordance with **Worksheets #18** and **#19**.
- Soil samples will be analyzed for 18 PFAS by Battelle using LC/MS/MS in compliance with QSM 5.3 Table B-15 (or the most recent version of the DoD QSM for which the laboratory is accredited) in accordance with Worksheets #18 and #19.
- Groundwater samples will be analyzed for 18 PFAS by Battelle by LC/MS/MS in compliance with QSM 5.3
 Table B-15(or the most recent version of the DoD QSM for which the laboratory is accredited) in accordance with Worksheets #18 and #19.
- Surface water samples will be analyzed for 18 PFAS by Battelle using LC/MS/MS in compliance with QSM 5.3
 Table B-15(or the most recent version of the DoD QSM for which the laboratory is accredited) in accordance
 with Worksheets #18 and #19.

Modeling

Data collected during the RI will be used to update the CSM and refine the existing groundwater flow and solute transport models. Model refinement will include, as necessary or applicable, updating the model parameterization, re-evaluation of boundary conditions, recalibration to steady-state conditions, updating initial concentrations for PFOA, PFOS, and PFBS, and using the most up-to-date solute transport parameters available for PFAS. HYDRUS software, or similar, will be used to conceptualize PFAS transport from the soil release areas to shallow groundwater. The solute mass flux values will be used as flux boundary conditions for the solute transport model. The model will be used to evaluate up to three future scenarios (such as changes to pumping rates or distributions).

SAP Worksheet #14—Summary of Project Tasks (continued)

QC Tasks

- Implement SOPs for field and laboratory activities being performed.
- QC samples are described on Worksheet #20.

Secondary Data

See Worksheet #13.

DV, Review, and Management Tasks

See Worksheets #34 through #36 for a discussion of data management procedures.

Documentation and Reporting

 A summary of field activities as well as a data evaluation will be documented in an RI Report and submitted to the NAVFAC Northwest RPM and stakeholder agencies for review and approval.

Assessment and Audit Tasks

Worksheets #31 and #32.

Demobilization

Full demobilization will occur when the project is completed, and appropriate QA/QC checks have been performed. Personnel no longer needed during the course of field operations may be demobilized prior to the final project completion date. The following will occur prior to demobilization:

- Chain-of-custody records will be reviewed to verify that all samples were collected as planned and submitted for appropriate analyses.
- Restoration of the site to an appropriate level will be verified by the CH2M FTL.
- All equipment will be inspected, packaged, and shipped to the appropriate location.

SAP Worksheet #15-1—Reference Limits and Evaluation Tables

Matrix: Groundwater

Analytical Group: PFAS – PFAS by LC/MS/MS Compliant with DoD QSM 5.3 Table B-15

Analyte	Chemical Abstract Service (CAS)	P/	Laboratory Limits (ng/L)			LCS and MS/MSD Recovery Limits and RPD (%) ³			
,	Number	Tapwater SLs (ng/L)	PAL Reference	LOQs (ng/L)	LODs (ng/L)	DLs (ng/L)	LCL	UCL	RPD
PFOA	335-67-1	40	USEPA RSL calculator ¹	5.00	1.50	0.511	49	141	30
PFOS	1763-23-1	40	USEPA RSL calculator ¹	5.00	1.00	0.437	40	144	30
PFBS	375-73-5	600	PPR Toxicity Values for PFBS (USEPA, 2021) ²	5.00	0.500	0.144	56	134	30
Perfluorohexanoic acid (PFHxA)	307-24-4			5.00	1.50	0.527	51	137	30
Perfluoroheptanoic acid (PFHpA)	375-85-9			5.00	1.00	0.263	48	136	30
Per Perfluorohexanoic fluorohexanesulfonic acid (PFHxS)	355-46-4			5.00	0.400	0.112	52	128	30
Perfluorononanoic acid (PFNA)	375-95-1			5.00	1.00	0.309	58	122	30
Perfluorodecanoic acid (PFDA)	335-76-2			5.00	0.500	0.142	59	135	30
Perfluoroundecanoic acid (PFUnA)	2058-94-8			5.00	0.500	0.219	64	134	30
Perfluorododecanoic acid (PFDoA)	307-55-1			5.00	0.50	0.192	75	131	30
Perfluorotridecanoic acid (PFTrDA)	72629-94-8			5.00	0.500	0.154	42	148	30
Perfluorotetradecanoic acid (PFTeDA)	376-06-7			5.00	2.00	0.733	42	158	30
N-Ethyl Perfluorooctanesulfonamidoacetic Acid (EtFOSAA)	2991-50-6			5.00	1.00	0.500	51	131	30

SAP Worksheet #15-1—Reference Limits and Evaluation Tables (continued)

Analyte	Chemical Abstract Service (CAS)	P.	PALs			Laboratory Limits (ng/L)			LCS and MS/MSD Recovery Limits and RPD (%) ³		
	Number	Tapwater SLs (ng/L)	PAL Reference	LOQs (ng/L)	LODs (ng/L)	DLs (ng/L)	LCL	UCL	RPD		
N-Methyl Perfluorooctanesulfonamidoacetic Acid (MeFOSAA)	2355-31-9			5.00	1.00	0.350	50	146	30		
Perfluoro-2-methyl-3-oxahexanoic acid (HFPO-DA)	13252-13-6			5.00	0.500	0.248	60	126	30		
4,8-dioxa-3H-perfluorononanoic acid (ADONA)	919005-14-4			5.00	1.00	0.265	61	130	30		
9-chlorohexadecafluoro-3-oxanone-1- sulfonic acid (9CI-PF3ONS)	756426-58-1			5.00	1.00	0.268	60	126	30		
11-chloroeicosafluoro-3-oxaundecane-1- sulfonic acid (11Cl-PF3OUdS)	763051-92-9			5.00	0.500	0.231	56	125	30		

Notes:

- SLs for PFOA and PFOS are based on an HQ of 0.1 and were generated using the USEPA RSL calculator as described in the Assistant Secretary of Defense (ASD) September 2021 Memo (ASD, 2021).
- SLs for PFBS were generated similarly to PFOA and PFOS, but values were updated from those listed in the 2021 memorandum to reflect reference doses provided in Provisional Peer-Reviewed Toxicity Values for Perfluorobutane Sulfonic Acid (PFBS) and Related Compound Potassium Perfluorobutane Sulfonate (PPR Toxicity Values for PFBS) (USEPA, 2021).
- ³ Accuracy and precision are in accordance with DoD QSM 5.3. For analytes not found in DoD QSM 5.3, laboratory in-house limits will be used. Analytes using in-house laboratory limits are bolded

Limits are verified on a quarterly basis in accordance with DoD QSM and may be subject to change. Any changes to these limits that affect the project SAP objectives must be approved by the NAVFAC RPM and NAVFAC Atlantic QAO in advance of sample testing.

-- = not applicable

LCL = lower control limit

LCS = laboratory control sample

RPD = relative percent difference

UCL = upper control limit

SAP Worksheet #15-2—Reference Limits and Evaluation Table

Matrix: Surface Soil, Ditch Surface Soil, Subsurface Soil

Analytical Group: PFAS – PFAS by LC/MS/MS Compliant with DoD QSM 5.3 Table B-15

Aughte	CAC Number		PALs	Labora	tory Limits (μg/kg)³	LCS and MS/MSD Recovery Limits and RPD (%) ⁴		
Analyte	CAS Number	Residential Soil SLs (μg/kg)	PAL Reference	LOQs (µg/kg)	LODs (μg/kg)	DLs (μg/kg)	LCL	UCL	RPD
PFOA	335-67-1	130	USEPA RSL calculator ¹	5.00	2.00	0.607	50	130	30
PFOS	1763-23-1	130	USEPA RSL calculator ¹	5.00	2.00	0.692	56	136	30
PFBS	375-73-5	1,900	PPR Toxicity Values for PFBS (USEPA, 2021) ²	5.00	1.00	0.350	57	145	30
PFHxA	307-24-4			5.00	2.00	0.707	45	135	30
PFHpA	375-85-9			5.00	1.50	0.506	60	128	30
PFHxS	355-46-4			5.00	2.00	0.808	52	132	30
PFNA	375-95-1			5.00	1.00	0.491	54	130	30
PFDA	335-76-2			5.00	1.00	0.463	55	141	30
PFUnA	2058-94-8			5.00	1.00	0.457	57	137	30
PFDoA	307-55-1			5.00	2.00	0.612	62	134	30
PFTrDA	72629-94-8			5.00	1.00	0.280	51	127	30
PFTeDA)	376-06-7			5.00	2.50	1.08	34	162	30
EtFOSAA	2991-50-6			5.00	2.00	0.748	54	124	30
MeFOSAA	2355-31-9			5.00	2.50	1.02	52	146	30
HFPO-DA	13252-13-6			5.00	2.00	0.641	71	153	30

SAP Worksheet #15-2—Reference Limits and Evaluation Table (continued)

Analyte CAS Niverban		PALs		Laboratory Limits (µg/kg) ³			LCS and MS/MSD Recovery Limits and RPD (%) ⁴		
Analyte	CAS Number	Residential Soil SLs (μg/kg)	PAL Reference	LOQs (µg/kg)	LODs (μg/kg)	DLs (μg/kg)	LCL	UCL	RPD
ADONA	919005-14-4			5.00	2.00	0.830	61	139	30
9CI-PF3ONS	756426-58-1			5.00	1.00	0.482	60	140	30
11Cl-PF3OUdS	763051-92-9			5.00	1.50	0.524	40	160	30

¹ Residential soil SLs for PFOA and PFOS are based on an HQ of 0.1 and were generated using the USEPA RSL calculator as described in the ASD September 2021 Memo (ASD, 2021).

Limits are verified on a quarterly basis per DoD QSM and may be subject to change. Any changes to these limits which impact the SAP objectives must be approved by the NAVFAC RPM and NAVFAC QAO in advance of sample testing.

-- = not applicable

LCL = lower control limit

LCS = laboratory control sample

RPD = relative percent difference

UCL = upper control limit

RSLs for PFBS were generated similarly to PFOA and PFOS, but values were updated from those listed in the 2021 memorandum to reflect reference doses provided in PPR Toxicity Values for PFBS (USEPA, 2021).

Results for nonaqueous samples are reported on a dry-weight basis.

⁴ Accuracy and precision limits are according to DoD QSM 5.3. For analytes not found in DoD QSM 5.3, laboratory in-house limits will be used. Analytes using in house laboratory limits are bolded.

SAP Worksheet #15-3—Reference Limits and Evaluation Table

Matrix: Surface Water

Analytical Group: PFAS – PFAS by LC/MS/MS Compliant with DoD QSM 5.3 Table B-15

Analyte	CAS Number	P.	ALs	Laboratory Limits (ng/L)			LCS and MS/MSD Recovery Limits and RPD (%) ³		
Andryce	CAS Hamber	Tapwater SLs (ng/L)	PAL Reference	LOQs (ng/L)	LODs (ng/L)	DLs (ng/L)	LCL	UCL	RPD
PFOA	335-67-1	40	USEPA RSL calculator ¹	5.00	1.50	0.511	56	136	30
PFOS	1763-23-1	40	USEPA RSL calculator ¹	5.00	1.00	0.437	50	130	30
PFBS	375-73-5	600	PPR Toxicity Values for PFBS (USEPA, 2021) ²	5.00	0.500	0.144	57	145	30
PFHxA	307-24-4			5.00	1.50	0.527	45	135	30
PFHpA	375-85-9			5.00	1.00	0.263	60	128	30
PFHxS	355-46-4			5.00	0.400	0.112	52	132	30
PFNA	375-95-1			5.00	1.00	0.309	54	130	30
PFDA	335-76-2			5.00	0.500	0.142	55	141	30
PFUnA	2058-94-8			5.00	0.500	0.219	57	137	30
PFDoA	307-55-1			5.00	0.50	0.192	62	134	30
PFTrDA	72629-94-8			5.00	0.500	0.154	51	127	30
PFTeDA	376-06-7			5.00	2.00	0.733	34	162	30
EtFOSAA	2991-50-6			5.00	1.00	0.500	54	124	30
MeFOSAA	2355-31-9			5.00	1.00	0.350	52	146	30
HFPO-DA	13252-13-6			5.00	0.500	0.248	60	126	30

SAP Worksheet #15-3—Reference Limits and Evaluation Table (continued)

Analyte	Analyte CAS Number		PALs			Laboratory Limits (ng/L)			LCS and MS/MSD Recovery Limits and RPD (%) ³			
		Tapwater SLs (ng/L) PAL Reference		LOQs (ng/L)	LODs (ng/L)	DLs (ng/L)	LCL	UCL	RPD			
ADONA	919005-14-4			5.00	1.00	0.265	61	130	30			
9CI-PF3ONS	756426-58-1			5.00	1.00	0.268	60	126	30			
11Cl-PF3OUdS	763051-92-9			5.00	0.500	0.231	56	125	30			

Notes:

- 1 SLs for PFOA and PFOS are based on an HQ of 0.1 and were generated using the USEPA RSL calculator as described in the ASD September 2021 Memo (ASD, 2021).
- 2 RSLs for PFBS were generated similarly to PFOA and PFOS, but values were updated from those listed in the 2021 memorandum to reflect reference doses provided in PPR Toxicity Values for PFBS (USEPA, 2021).
- Accuracy and precision limits are according to DoD QSM 5.3. For analytes not found in DoD QSM 5.3, laboratory in-house limits will be used. Analytes using in house laboratory limits are bolded.

Limits are verified on a quarterly basis per DoD QSM and may be subject to change. Any changes to these limits which impact the project SAP objectives, must be approved by the NAVFAC RPM and NAVFAC Atlantic QAO in advance of sample testing.

-- = not applicable

LCL = lower control limit

LCS = laboratory control sample

RPD = relative percent difference

UCL = upper control limit

SAP Worksheet #16—Project Schedule/Timeline Table

		Da	ates		
Phase 1 RI Activities ¹	Organization	Anticipated Date(s) of Initiation	Anticipated Date of Completion	Deliverable	
Internal Draft SAP preparation	CH2M	November 2020	April 2021	Internal Draft SAP	
Navy SAP review NAVFAC Northwest and NAVFAC Atlantic		May 2021	June 2021	Comments	
Draft SAP preparation	CH2M	June 2021	July 2021	Draft SAP	
	USEPA Region 10				
	Island County Public Health, Washington				
Stakeholder review	Town of Coupeville, Washington	October 2021	November 2021	Comments	
	Washington Department of Ecology				
	Washington Department of Health				
Final SAP	CH2M	December 2021	January 2022	Final SAP	
Subcontractor procurement	CH2M	TBD	TBD		
Groundwater monitoring well installation and survey	CH2M, Subcontractor	January 2022	March 2022		
Groundwater and soil sampling	CH2M	January 2022	March 2022		
Surface water and ditch surface soil sampling	CH2M	January 2022	March 2022		
Laboratory analysis Subcontractor		Varied turnaround times are	e detailed on Worksheet #30.	Analytical data	
Data management	CH2M	TBD	TBD		
Data validation	Subcontractor	TBD	TBD		
Reporting CH2M		TBD	TBD	Internal Draft, Draft, and Final RI Report	

Phase 1 of the RI includes on-Base activities only. Off-Base activities will be described in a future addendum to this SAP. Additional RI phases, as necessary, will be described under future task order work plan documents.

PLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE IL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON ARY 2022 64	

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SAP Worksheet #17—Sampling Design and Rationale

The objectives of the investigation described in this worksheet are listed in **Worksheet #11**. Media to be investigated for this SAP include soil from on-base soil borings, groundwater from new well borings and new and existing completed on-base monitoring wells, and on-base ditch surface soil and surface water (if present). The sampling strategy and rationale are detailed in **Table 17-1**. Groundwater, soil, and surface water samples will be analyzed for PFAS by LC/MS/MS in compliance with QSM 5.3 Table B-15 or the latest version of the QSM for which the laboratory is certified at the time of sampling. Actual sample locations may vary from the proposed sample locations presented on **Figure 11-2** because of field conditions.

Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
						Soil samples will be collected for laboratory analysis of PFAS from an on-base soil boring location near a groundwater monitoring well where PFOS has been detected above the USEPA tap water RSL.	Samples will be used to determine the presence of PFOS in soil near groundwater monitoring well WI-CV-MW23S, where PFOS was previously detected above the USEPA tap water RSL but for which a source is not
WI-CV-SO07						Up to 5 soil samples will be collected from this location: at the surface (0 to 6 inches bgs), at the top of the capillary fringe in the unsaturated soil, and at up to three additional lithologic interfaces determined during drilling.	apparent. Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air water and lithologic interfaces. Data will be used to determine whether the sampling location is considered an additional vadose zone source and to characterize the vertical distribution of PFAS in the vadose zone.
WI-CV-SO08						Soil samples will be collected for laboratory analysis of PFAS from on-base soil borings	
WI-CV-SO09						and from the WI-CV-MW33S borehole (prior to monitoring well installation) west of the runway near the Keystone Well.	
WI-CV-SO10				LC/MS/MS in	Up to 5 at each location ¹	Up to five soil samples will be collected from each location: at the surface (0 to 6 inches bgs), at the top of the capillary fringe in the unsaturated soil, and at up to three	Samples will be used to determine the presence of PFAS in soil near groundwater monitoring wells west of the runway (WI-CV-MW15S/M, WI-CV-MW16S/M) where PFAS have been detected above the SL (on-base near the Keystone Well), but evidence suggests that PFAS is not related to release areas east of the runway. Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air-water and
NA// CN/ NANA/22C	Soil	TBD¹ (between the ground surface [0 to 6	PFAS	compliance with DoD QSM		additional lithologic interfaces determined during drilling.	lithologic interfaces. Data will be used to determine whether the sampling locations are considered an addition vadose zone source.
WI-CV-MW33S	3011	inches] and the water table)	FFAS	5.3, Table B-15/ SOP 5-369-08		Soil borings will be installed early in the investigation to help drive later decisions on location and screen placement of groundwater monitoring wells in this area.	
WI-CV-SO11						Soil samples will be collected for laboratory analysis of PFAS from the soil borings near	
WI-CV-SO12						confirmed on-base release areas.	Samples will provide greater delineation of on-base release areas, which were confirmed as PFAS vadose zone
WI-CV-SO13 WI-CV-SO14						Up to five soil samples will be collected from each location: at the surface (0 to 6 inches bgs), at the top of the capillary fringe in the unsaturated soil, and at up to three additional lithologic interfaces determined during drilling.	sources during the Supplemental SI (Building 2709 and Facility 1, 2, and 11). Depths targeted for analysis will be identified based on boring-specific conditions and will focus on air-water and lithologic interfaces. Data will be used to better define the horizontal and vertical extent of PFAS in the vadose zone at these areas.
TBD					TBD¹	Additional soil samples may be collected at other soil boring locations or depths for laboratory analysis of PFAS from on-base soil borings advanced at or near release areas.	Samples will provide additional data as necessary to delineate the horizontal and vertical extent of PFAS in soil of Base at release areas. Depths targeted for analysis will be identified based on boring-specific conditions and will
עסו					-טסו	Samples will be collected from surface or subsurface soil as deep as the top of the capillary fringe, targeting lithologic interfaces identified during drilling.	focus on air-water and lithologic interfaces.

Table 17-1. Sampling Strategy Table – OLF Coupeville RI

Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
WI-CV-MW32M WI-CV-MW33M						Groundwater will be collected as vertical profiling grab samples for in-field water	Depth-discrete groundwater profiling with PFAS sampling will be conducted at soil borings located west of the runway near the Keystone Well; data will be used to determine well screen intervals for paired shallow/intermediate permanent groundwater monitoring wells (targeting elevated PFAS concentrations and discrete aquifer zones for well installation), and to confirm the presence of an additional suspected release in this area. If no elevated PFAS concentrations are detected, or multiple discrete aquifer zones are not observed during
					Estimated	quality parameters and laboratory analysis of PFAS at depths between the surface and	drilling, well construction at this location will be determined based on observed lithology, in consultation with the project technical team and the NAVFAC Northwest RPM.
WI-CV-MW35M					3 at each location ² b	200 feet bgs as determined in the field based on lithology and targeting discrete aquifer intervals observed. Groundwater samples will be collected via a drive-point sampling device and submitted for PFAS analysis with a 72-hour TAT to allow for determination of groundwater monitoring	Depth-discrete groundwater profiling with PFAS sampling will be conducted at a soil boring east of the runway, downgradient (southwest) of Building 2709 and Facilities 1, 2, and 11; data will be used to determine well screen intervals for paired shallow/intermediate permanent groundwater monitoring wells (targeting elevated PFAS concentrations and discrete aquifer zones for well installation), and to address PFAS concentration data gaps between vadose zone sources (Building 2709 and Facilities 1, 2, and 11) and impacted downgradient monitoring wells to the west and southwest of the runway.
						well screen intervals. Up to three groundwater grab samples per location will be collected during	If no elevated PFAS concentrations are detected, or multiple discrete aquifer zones are not observed during drilling, well construction at this location will be determined based on observed lithology, in consultation with the project technical team and the NAVFAC Northwest RPM.
WI-CV-MW36D	Groundwater	TBD¹ (between the water table and approximately 200 feet bgs)	PFAS	LC/MS/MS in compliance with DoD QSM 5.3, Table B-15/ SOP 5-369-08		advancement of soil borings for wells WI-CV-MW32M, WI-CV-MW33M, and WI-CV-MW35M, targeting the first-encountered groundwater, and up to two additional discrete aquifer zones separated by a low-permeability confining unit between the	Depth-discrete groundwater profiling with PFAS sampling will be conducted at soil borings located near the southwestern base boundary; data will be used to determine well screen intervals for paired intermediate/deep permanent groundwater monitoring wells (targeting elevated PFAS concentrations and discrete aquifer zones for well installation), provide bounding data near the southwestern base boundary, and address a PFAS concentration data gap between on-base wells west of the runway and off-base wells where PFAS has been detected in exceedance of the USEPA lifetime health advisory.
				SUP 5-369-08	Estimated	water table and the total depth of boring. Up to four groundwater grab samples per location will be collected during	If no elevated PFAS concentrations are detected, or multiple discrete aquifer zones are not observed during drilling, well construction at this location will be determined based on observed lithology, in consultation with the project technical team and the NAVFAC Northwest RPM.
WI-CV-MW37D					4 at each location ²	advancement of soil borings for wells WI-CV-MW36D and WI-CV-MW37D, with up to three samples collected from between the discrete aquifer zones between the water table and the total depth of boring.	Depth-discrete groundwater profiling with PFAS sampling will be conducted at soil borings located near the tree line in the southwestern part of the Base; data will be used to determine well screen intervals for paired intermediate/deep permanent groundwater monitoring wells (targeting elevated PFAS concentrations and discrete aquifer zones for well installation), and to identify potential PFAS migration pathways between northern on-base wells west of the runway and off-base drinking water wells where PFAS has been detected in exceedance of the USEPA lifetime health advisory.
							If no elevated PFAS concentrations are detected, or multiple discrete aquifer zones are not observed during drilling, well construction at this location will be determined based on observed lithology, in consultation with the project technical team and the NAVFAC Northwest RPM.
TBD					TBD ¹	Additional groundwater may be collected as vertical profiling grab samples for in-field water quality parameters and laboratory analysis of PFAS at depths between the surface and 200 feet bgs as determined in the field based on lithology and aquifer intervals observed.	Samples will provide additional data as necessary to delineate the vertical extent of PFAS in groundwater at proposed monitoring well locations on-base and to determine well screen intervals for monitoring wells. Depths targeted for analysis will be identified based on boring-specific conditions and will focus collection of samples within discrete water-bearing aquifer zones.

Table 17-1. Sampling Strategy Table – OLF Coupeville RI

Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
WI-CV-MW32S	Mi Mi We TB Groundwater Mi Mi We TB Mi Mi We TB	Middle of well screen; well screen settings	PFAS	LC/MS/MS in compliance with DoD QSM 5.3, Table B-15/ SOP 5-369-08	1 at each location	Groundwater will be collected for in-field water quality parameters and laboratory analysis of PFAS using low-flow sampling techniques.	Groundwater analytical data from a newly installed on-base shallow/intermediate groundwater monitoring well pair south of the WI-CV-MW16 pair and west of WI-CV-MW25M-R will be used to evaluate the connection between the release area near the WI-CV-MW15 and WI-CV-MW16 well pairs and PFOA/PFOS concentrations further south. The data will also be used to provide bounding on PFOA and PFOS exceedances of the SL (40 ng/L) near the western Base boundary.
WI-CV-MW32M							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.
WI-CV-MW33S		TBD ¹					Groundwater analytical data from a newly installed on-base shallow/intermediate groundwater monitoring well pair east of the WI-CV-MW15 well pair will be used to confirm the presence of a release area near the WI-CV-MW15 and WI-CV-MW16 well pairs.
WI-CV-MW33M							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.
NA/I CV/ NAVA/2 AC		Middle of well screen; well screen settings TBD ¹			1 at each location	Groundwater will be collected for in-field water quality parameters and laboratory analysis of PFAS using low-flow sampling techniques.	Groundwater analytical data from a newly installed on-base shallow groundwater monitoring well east of WI-CV-MW20S will be used to establish bounding on PFOA and PFOS exceedances of the SL (40 ng/L) near the eastern Base boundary.
WI-CV-MW34S							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.
WI-CV-MW35S							Groundwater analytical data from a newly installed on-base shallow/intermediate groundwater monitoring well pair between WI-CV-MW02S and WI-CV-MW05S/M will be used to evaluate the southward/southwesterly migration of PFAS from the vadose zone sources near Building 2709 and Facilities 1, 2, and 11.
WI-CV-MW35M							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.
WI-CV-MW36M							Groundwater analytical data from a newly installed on-base intermediate/deep groundwater monitoring well pair near the southwestern Base boundary will be used to provide bounding data near the southwestern base boundary, and address a PFAS concentration data gap between on-base wells west of the runway and off-base wells where PFAS has been detected in exceedance of the USEPA lifetime health advisory (70 ng/L).
WI-CV-MW36D							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.
WI-CV-MW37M							Groundwater analytical data from a newly installed on-base intermediate/deep groundwater monitoring well pair along the tree line in the southwestern part of the Base will be used to identify potential PFAS migration pathways between on-base wells west of the runway and off-base drinking water wells where PFAS has been detected in exceedance of the USEPA lifetime health advisory (70 ng/L).
WI-CV-MW37D							Future groundwater analytical data from these wells may be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville. However, only one sample collection event is scoped for this investigation.

Table 17-1. Sampling Strategy Table – OLF Coupeville RI

Table 17-1. Sampling Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
WI-CV-MW01M		Well screened from 148 to 158 feet bgs	PFAS 5.	LC/MS/MS in compliance with DoD QSM 1 a		Groundwater will be collected for in-field water quality parameters and laboratory analysis of PFAS using low-flow sampling techniques.	Groundwater analytical data from existing on-base wells at OLF Coupeville will be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville.
WI-CV-MW01D		Well screened from 202 to 212 feet bgs					
WI-CV-MW02S		Well screened from 91.5 to 101.5 feet bgs					
WI-CV-MW02M		Well screened from 153 to 163 feet bgs					
WI-CV-MW03S		Well screened from 120 to 130 feet bgs					
WI-CV-MW03M		Well screened from 145 to 155 feet bgs					
WI-CV-MW03D		Well screened from 222 to 232 feet bgs					
WI-CV-MW04S		Well screened from 111.6 to 121.6 feet bgs			1 at each location		
WI-CV-MW04M		Well screened from 148.7 to 158.7 feet bgs					
WI-CV-MW05S		Well screened from 114 to 124 feet bgs					
WI-CV-MW05M	Groundwater	Well screened from 160 to 170 feet bgs					
WI-CV-MW06S		Well screened from 130 to 140 feet bgs					
WI-CV-MW06M		Well screened from 174 to 184 feet bgs					
WI-CV-MW07S		Well screened from 129.5 to 139.5 feet bgs					
WI-CV-MW07M		Well screened from 183 to 193 feet bgs					
WI-CV-MW08S		Well screened from 121 to 131 feet bgs					
WI-CV-MW08M		Well screened from 150 to 160 feet bgs					
WI-CV-MW09S		Well screened from 96 to 106 feet bgs					
WI-CV-MW09M		Well screened from 182 to 192 feet bgs					
WI-CV-MW10M		Well screened from 144 to 154 feet bgs					
WI-CV-MW10D		Well screened from 191 to 201 feet bgs			!		

SAP Worksheet #17—Sampling Design and Rationale (continued)

Table 17-1. Sampling	Strategy Table – C	DLF Coupeville RI	T	T	1		
Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
WI-CV-MW11S		Well screened from 130 to 140 feet bgs	PFAS	LC/MS/MS in compliance with DoD QSM 5.3, Table B-15/SOP 5-369-08	1 at each location	Groundwater will be collected for in-field water quality parameters and laboratory analysis of PFAS using low-flow sampling techniques.	Groundwater analytical data from existing on-base wells at OLF Coupeville will be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville.
WI-CV-MW11M		Well screened from 155 to 165 feet bgs					
WI-CV-MW12S		Well screened from 97 to 107 feet bgs					
WI-CV-MW12D		Well screened from 183 to 193 feet bgs					
WI-CV-MW13S		Well screened from 105 to 115 feet bgs					
WI-CV-MW13M		Well screened from 173 to 183 feet bgs					
WI-CV-MW14M		Well screened from 161 to 171 feet bgs					
WI-CV-MW15S		Well screened from 132 to 142 feet bgs					
WI-CV-MW15M	Groundwater	Well screened from 164 to 174 feet b					
WI-CV-MW16S		Well screened from 130 to 140 feet bgs					
WI-CV-MW16M		Well screened from 165 to 183 feet bgs					
WI-CV-MW17M		Well screened from 150 to 160 feet bgs					
WI-CV-MW18M		Well screened from 120 to 135 feet bgs					
WI-CV-MW20S		Well screened from 100 to 110 feet bgs					
WI-CV-MW21S		Well screened from 107 to 117 feet bgs					
WI-CV-MW22S		Well screened from 112 to 122 feet bgs					
WI-CV-MW23S		Well screened from 130 to 140 feet bgs					
WI-CV-MW25M-R		Well screened from 150 to 160 feet bgs					
WI-CV-MW26D		Well screened from 189.5 to 199.5 feet bgs					
WI-CV-MW28M		Well screened from 160 to 170 feet bgs					
WI-CV-MW29M		Well screened from 160 to 170 feet bgs					

Table 17-1. Sampling Strategy Table – OLF Coupeville RI

Location	Matrix	Depth of Samples (feet bgs)	Analysis	Laboratory Method	Number of Samples	Sampling Strategy	Rationale
WI-CV-MW30M	Groundwater	Well screened from 160 to 170 feet bgs	PFAS	LC/MS/MS in compliance with DoD QSM 5.3, Table B-15/ SOP 5-369-08	1 at each location	Groundwater will be collected for in-field water quality parameters and laboratory analysis of PFAS using low-flow sampling techniques.	Groundwater analytical data from existing on-base wells at OLF Coupeville will be used to monitor temporal variability in PFAS concentrations in groundwater at OLF Coupeville.
WI-CV-MW31S		Well screened from 107 to 117 feet bgs					
WI-CV-MW31M		Well screened from 140 to 150 feet bgs					
WI-CV-SW01					1 at each	Collect surface water samples (if sufficient water) from drainage ditches along runway ³ (Figure 11-2).	
WI-CV-SW02		N/A					
WI-CV-SW03					location		
WI-CV-SW04	Surface Water		co	!			
TBD		TBDª		LC/MS/MS in compliance with DoD QSM	TBDª	Additional surface water samples may be collected at other drainage ditch locations for laboratory analysis of PFAS based on extent of observed surface water at the time of sampling.	Impacts on surface water (if present) and ditch surface soil have not been evaluated at OLF Coupeville. Sample data for ditch surface soil and surface water (if present) will be evaluated to determine whether runoff to on-base ditches from PFAS release
WI-CV-SS01			5.3, Table B-15/				areas occurred, PFAS may have accumulated in surface water (if present) and/or ditch surface soil and
WI-CV-SS02	Ditch Surface Soil	O to O E foot has		1 at each	Collect ditch surface soil samples from	provide a receptor pathway for maintenance and landscape workers as well as an additional transport pathway to groundwater.	
WI-CV-SS03		0 to 0.5 foot bgs		l le		drainage ditches along runway ³ (Figure 11-2).	
WI-CV-SS04							
TBD		TBD ¹			TBD ¹	Additional ditch surface soil samples may be collected at other drainage ditch locations for laboratory analysis of PFAS based on extent of present or potential surface water pathways observed.	

The final number and placement of samples may be modified in the field based on the field team's professional opinion in consultation with CH2M management and technical teams and the NAVFAC Northwest RPM. Sampling will target surface soil (0 to 0.5 foot bgs), the top of the capillary fringe in the unsaturated soil, and at up to three additional lithologic interfaces encountered during drilling for laboratory analysis of PFAS from each soil boring.

The final number and placement of samples may be modified in the field based on the field team's professional opinion in consultation with CH2M management and technical teams and the NAVFAC Northwest RPM. Sampling will target the first-encountered groundwater (water table) and up to two additional discrete aquifer zones separated by a confining unit.

³ If surface water is not present in runway drainage ditches or wetlands, only ditch surface soil samples will be collected from the specified locations.

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
VAIL CV COO7	WI-CV-SO07-TDBD		Up to 5			
WI-CV-SO07	WI-CV-SO07P-TDBD		1 (FD)			
WI-CV-SO08	WI-CV-SO08-TDBD		Up to 5			
VVI-CV-3008	WI-CV-SO08P-TDBD		1 (FD)			
	WI-CV-SO09-TDBD		Up to 5			
WI-CV-SO09	WI-CV-SO09-TDBD-MS		1 (MS)			
	WI-CV-SO09-TDBD-MSD		1 (MSD)			
WI-CV-SO10	WI-CV-SO10-TDBD		Up to 5	Surface (0 to 0.5 foot bgs),		
WI-CV-3010	WI-CV-SO10P-TDBD		1 (FD)	subsurface (at the top of the		
WI-CV-SO11	WI-CV-SO11-TDBD	Soil	Up to 5	capillary fringe), and up to three samples at other lithologic interfaces encountered during drilling	PFAS (LC/MS/MS compliant in accordance with DoD QSM 5.3, Table B-15 ¹)	Worksheet #21
	WI-CV-SO11P-TDBD		1 (FD)			
WI-CV-SO12	WI-CV-SO12-TDBD		Up to 5			
WI-CV-3012	WI-CV-SO12P-TDBD		1 (FD)			
	WI-CV-SO13-TDBD		Up to 5			
WI-CV-SO13	WI-CV-SO13-TDBD-MS		1 (MS)			
	WI-CV-SO13-TDBD-MSD		1 (MSD)			
	WI-CV-SO14-TDBD		Up to 5			
WI-CV-SO14	WI-CV-SO14-TDBD-MS		1 (MS)			
	WI-CV-SO14-TDBD-MSD		1 (MSD)			
WI-CV-MW01M	WI-CV-GW01M-MMYY		1	148 to 158		
WI-CV-MW01D	WI-CV-GW01D-MMYY	Groundwater	1	202 to 212		
VVI-CV-IVIVVOID	WI-CV-GW01DP-MMYY	Jordanawater	1 (FD)	202 10 212		
WI-CV-MW02S	WI-CV-GW02S-MMYY		1	91.5 to 101.5		

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
WI-CV-MW02M	WI-CV-GW02M-MMYY		1	153 to 163		
WI-CV-MW03S	WI-CV-GW03S-MMYY		1	120 to 130		
	WI-CV-GW03M-MMYY		1			
WI-CV-MW03M	WI-CV-GW03M-MMYY-MS		1 (MS)	145 to 155		
	WI-CV-GW03M-MMYY-MSD		1 (MSD)			
WI-CV-MW03D	WI-CV-GW03D-MMYY		1	222 to 232]	
WI-CV-MW04S	WI-CV-GW04S-MMYY		1	111.6 to 121.6]	Worksheet #21
WI-CV-MW04M	WI-CV-GW04M-MMYY		1	148.7 to 158.7		
WI-CV-MW05S	WI-CV-GW05S-MMYY	7	1	114 to 124	PFAS (LC/MS/MS compliant in accordance with DoD QSM 5.3, Table B-15 ¹)	
VVI-CV-IVIVVU33	WI-CV-GW05SP-MMYY		1 (FD)	114 (0 124		
WI-CV-MW05M	WI-CV-GW05M-MMYY	1	1	160 to 170		
WI-CV-MW06S	WI-CV-GW06S-MMYY Gr	roundwater	1	130 to 140		
WI-CV-MW06M	WI-CV-GW06M-MMYY		1	174 to 184		
WI-CV-MW07S	WI-CV-GW07S-MMYY		1	129.5 to 139.5		
	WI-CV-GW07M-MMYY		1			
WI-CV-MW07M	WI-CV-GW07M-MMYY-MS		1 (MS)	183 to 193		
	WI-CV-GW07M-MMYY-MSD		1 (MSD)			
WI-CV-MW08S	WI-CV-GW08S-MMYY		1	121 to 131		
WI-CV-MW08M	WI-CV-GW08M-MMYY		1	150 to 160]	
WI-CV-MW09S	WI-CV-GW09S-MMYY		1	96 to 106		
WI-CV-MW09M	WI-CV-GW09M-MMYY		1	182 to 192]	
WI-CV-MW10M	WI-CV-GW10M-MMYY		1	144 to 154		
VVI-CV-IVIVV I UIVI	WI-CV-GW10MP-MMYY		1 (FD)	7 144 (0 104		

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
WI-CV-MW10D	WI-CV-GW10D-MMYY		1	191 to 201		
WI-CV-MW11S	WI-CV-GW11S-MMYY		1	130 to 140		
WI-CV-MW11M	WI-CV-GW11M-MMYY		1	155 to 165		
WI-CV-MW12S	WI-CV-GW12S-MMYY		1	97 to 107		
WI-CV-MW12D	WI-CV-GW12D-MMYY		1	183 to 193		
WI-CV-MW13S	WI-CV-GW13S-MMYY		1	105 to 115		Worksheet #21
\\\\\ C\\\ \\\\\\\\\\\\\\\\\\\\\\\\\\\	WI-CV-GW13M-MMYY		1	172 to 102		
WI-CV-MW13M	WI-CV-GW13MP-MMYY		1 (FD)	- 173 to 183	PFAS (LC/MS/MS compliant in accordance with DoD QSM 5.3, Table B-15 ¹)	
WI-CV-MW14M	WI-CV-GW14M-MMYY		1	161 to 171		
WI-CV-MW15S	WI-CV-GW15S-MMYY		1	132 to 142		
WI-CV-MW15M	WI-CV-GW15M-MMYY		1	164 to 174		
WI-CV-MW16S	WI-CV-GW16S-MMYY Gr	roundwater	1	130 to 140		
WI-CV-MW16M	WI-CV-GW16M-MMYY		1	164 to 174		
NA/I ON ANA/A 78 A	WI-CV-GW17M-MMYY		1	150 to 170		
WI-CV-MW17M	WI-CV-GW17MP-MMYY		1 (FD)	- 150 to 160		
WI-CV-MW18M	WI-CV-GW18M-MMYY		1	120 to 135		
WI-CV-MW20S	WI-CV-GW20S-MMYY		1	100 to 110		
WI-CV-MW21S	WI-CV-GW21S-MMYY		1	107 to 117		
WI-CV-MW22S	WI-CV-GW22S-MMYY		1	112 to 122		
WI-CV-MW23S	WI-CV-GW23S-MMYY		1	130 to 140		
WI-CV-MW25M-R	WI-CV-GW25M-MMYY		1	150 to 160		
WI-CV-MW26D	WI-CV-GW26D-MMYY		1	189.5 to 199.5		
WI-CV-MW28M	WI-CV-GW28M-MMYY		1	160 to 170		

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
WI-CV-MW29M	WI-CV-GW29M-MMYY		1	160 to 170		
WI-CV-MW30M	WI-CV-GW30M-MMYY		1	160 to 170		
WI-CV-MW31S	WI-CV-GW31S-MMYY		1	107 to 117		
WI-CV-MW31M	WI-CV-GW31M-MMYY		1	140 to 150		
WI-CV-MW32S	WI-CV-GW32S-MMYY	Groundwater	1	TBD ²		
	WI-CV-GW32M-XX-MMYY		An estimated 3 (grab samples)	_		
WI-CV-MW32M	WI-CV-GW32MP-XX-MMYY		1 (FD)	TBD ²		
	WI-CV-GW32M-MMYY		1			
WI-CV-MW33S	WI-CV-SO33-TDBD-MMYY	Soil	Up to 5	Surface (0 to 0.5 foot bgs), subsurface (at the top of the capillary fringe), and up to three samples at other lithologic interfaces encountered during drilling	PFAS (LC/MS/MS compliant in accordance with DoD QSM 5.3, Table B-151)	Worksheet #21
	WI-CV-GW33S-MMYY	Carriandonatas	1			
	WI-CV-GW33SP-MMYY	Groundwater	1 (FD)			
WI-CV-MW33M	WI-CV-GW33M-XX-MMYY		An estimated 3 (grab samples)			
	WI-CV-GW33M-MMYY		1	TBD ²		
	WI-CV-GW34S-MMYY	Groundwater	1			
WI-CV-MW34S	WI-CV-GW34S-MMYY-MS		1 (MS)			
	WI-CV-GW34S-MMYY-MSD		1 (MSD)			
WI-CV-MW35S	WI-CV-GW35S-MMYY		1			

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
	WI-CV-GW35M-XX-MMYY		An estimated 3 (grab samples)			
WI-CV-MW35M	WI-CV-GW35MP-XX-MMYY]	1 (FD)			
	WI-CV-GW35M-MMYY]	1			
WI-CV-MW36M	WI-CV-GW36M-MMYY]	1	TDD2		
	WI-CV-GW36D-XX-MMYY	1	An estimated 4 (grab samples)	- TBD²		Worksheet #21
WI-CV-MW36D	WI-CV-GW36D-XX-MMYY-MS	Groundwater			PFAS (LC/MS/MS compliant in accordance with DoD QSM 5.3, Table B-15 ¹)	
	WI-CV-GW36D-XX-MMYY-MSD	1				
	WI-CV-GW36D-MMYY					
WI-CV-MW37M	WI-CV-GW37M-MMYY	1	1			
WI-CV-MW37D	WI-CV-GW37D-XX-MMYY	1	An estimated 4 (grab samples)	TBD ²		
	WI-CV-GW37D-MMYY]	1			
	WI-CV-SW01-MMYY	Curtoso vuotor	1	NI/A		
WI-CV-SW01/WI-	WI-CV-SW01P-MMYY	- Surface water	1 (FD)	N/A		
CV-SS01	WI-CV-SS01-MMYY	Ditch surface soil	1	0 to 0.5		
	WI-CV-SW02-MMYY		1			
	WI-CV-SW02-MMYY-MS	Surface water	1 (MS)	N/A		
WI-CV-SW02/WI- CV-SS02	WI-CV-SW02-MMYY-MSD		1 (MSD)			
	WI-CV-SS02-MMYY	Ditch surface soil	1	0 to 0.5		

Station ID	Sample ID	Matrix	Number of Samples (Identify FDs)	Depth (feet bgs)	Analytical Group	Sampling SOP Reference
	WI-CV-SW03-MMYY	Surface water	1	N/A		
WI-CV-SW03/WI- CV-SS03	WI-CV-SS03-MMYY	Ditch surface	1	0 to 0.5		
	WI-CV-SS03P-MMYY	soil	1 (FD)	0 10 0.5	PFAS (LC/MS/MS	
WI-CV-SW04/WI- CV-SS04	WI-CV-SW04-MMYY	Surface water	1	N/A	compliant in accordance with	Worksheet
	WI-CV-SS04-MMYY		1		DoD QSM 5.3, Table B-15 ¹)	#21
	WI-CV-SS04-MMYY-MS	Ditch surface soil	1 (MS)	0 to 0.5	Tuble B 10)	
	WI-CV-SS04-MMYY-MSD		1 (MSD)			
	1	1	Field QC Samples		1	
	WI-CV-EB01-MMDDYY		1		PFAS (LC/MS/MS	
	WI-CV-EB02-MMDDYY	7	1			
WI-CV-QC ³	WI-CV-EBXX-MMDDYY ³	QC	TBD	N / A	compliant in	Worksheet
VVI-UV-QU	WI-CV-FB01-MMDDYY		1	- N/A	accordance with DoD QSM 5.3, Table B-15 ¹)	#21
	WI-CV-FB02-MMDDYY		1			
	WI-CV-FBXX-MMDDYY ³		TBD			

Analytical method is compliant with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which Battelle has DoD ELAP certification.

Notes:

ID = identification

MMDDYY = Month/Day/Year

MMYY = Month/Year

TDBD = top depth/bottom depth (soil samples)

XX = depth (groundwater grab samples)

² Depth intervals will be determined during drilling.

One field blank should be collected weekly and one equipment blank should be collected daily with samples from decontaminated equipment.

SAP Worksheet #19—Analytical SOP Requirement Table

Matrix	Analytical Group	Analytical and Preparation Method/SOP Reference	Containers	Sample Volume	Preservation Requirements	Maximum Holding Time ¹ (Preparation/Analysis)
Groundwater/surface water	PFAS	PFAS by LC/MS/MS in compliance with DoD QSM 5.3 Table B-15 ² /SOP 5-370-12/SOP 5-369-08	2 x 250-milliliter HDPE bottles	2 x 250 milliliters	Temperature must be above freezing and less than or equal to 10°C when received at the laboratory. Samples stored in the lab must be held at or below 6°C until extraction but should not be frozen.	14 days to extraction/28 days to analysis
Soil (subsurface soil and ditch surface soil)	PFAS	PFAS by LC/MS/MS in compliance with DoD QSM 5.3 Table B-15 ² /SOP 5-370-12/SOP 5-369-08	One 8-ounce HDPE jar	20 grams	Temperature must be above freezing and less than or equal to 10°C when received at the laboratory. Samples stored in the lab must be held at or below 6°C until extraction but should not be frozen.	14 days to extraction/28 days to analysis

¹ Maximum holding time is calculated from the time the sample is collected to the time the sample is prepared/extracted.

Note:

HDPE = high-density polyethylene

² Analytical method is compliant with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which Battelle has DoD ELAP certification.

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SAP Worksheet #20—Field Quality Control Sample Summary Table

Matrix	Analytical Group	No. of Field Samples ¹	No. of FDs ^a	No. of MS/MSDs ¹	No. of Field Reagent Blanks ¹	No. of Equipment Blanks ¹	Total No. of Samples to Laboratory ¹
Groundwater	PFAS	73	8	4/4	1	23	113
Soil	PFAS	45	5	3/3	1	10	67
Surface Water	PFAS	4	1	1/1	1	1	9
Ditch Surface Soil	PFAS	4	1	1/1	1	1	9

Samples will be collected as detailed in Worksheets #14, #17, and #18 of this SAP. Field QA/QC samples will be collected as detailed in Worksheet #12.

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Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP CH2M-1	Logging of Soil Borings Rev. Sept.2021	CH2M	None	N	Guides staff in accurately and consistently recording the field data necessary to characterize soil borings and recovered soil samples, to guide CH2M staff in accurately and consistently recording the field data necessary to characterize soil borings and recovered soil samples.
SOP CH2M-2	Water-Level Measurements Rev. Feb. 2021	CH2M	Transducer and datalogger	N	Describes procedure for collecting continuous water level measurements. Only PFAS-free equipment will be used.
SOP CH2M-3	Multi RAE Photoionization Detector (PID) Rev. Feb. 2021	CH2M	Multi RAE PID	N	Describes procedure for operation and general maintenance of the Multi RAE PID.
SOP CH2M-4	Groundwater Sampling for Per- and Polyfluoroalkyl Substances (PFAS) Rev. Jan. 2021	CH2M	Teflon-free tubing, Teflon-free bailer (if using bailer), PFAS-free pump, sample bottles (HDPE bottle with HDPE screw cap), laboratory pre-filled HDPE bottles containing field blank water, loose leaf paper without waterproof coating, clip board, pen (not Sharpie), nitrile or latex gloves	N	Provides guidance for groundwater sample collection for samples that will be analyzed for PFAS via LC/MS/MS in compliance with QSM 5.3 (or the most recent version of the QSM for which laboratory has been certified at the time of sampling) for Navy CLEAN projects under Contract N62470-16-D-9000.
SOP CH2M-5	Rotosonic Groundwater Sample Collection for Per- and Polyfluoroalkyl Substances Rev. Mar. 2019	CH2M	Teflon-free tubing, Teflon-free bailer (if using bailer), PFAS-free pump, sample bottles (HDPE bottle with HDPE screw cap), laboratory pre-filled HDPE bottles containing field blank water, loose leaf paper without waterproof coating, clip board, pen (not Sharpie), nitrile or latex gloves	N	Provides guidance for groundwater sample collection for samples that will be analyzed for PFAS via LC/MS/MS in compliance with QSM 5.3 (or the most recent version of the QSM for which laboratory has been certified at the time of sampling) for Navy CLEAN projects under Contract N62470-16-D-9000.

Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP CH2M-6	Soil Sampling for Per- and Polyfluoroalkyl Substances Rev. Jan 2021	CH2M	Sample jars (HDPE bottle with HDPE screw cap), laboratory pre-filled HDPE bottles containing field blank water, loose leaf paper without waterproof coating, clip board, pen (not Sharpie), nitrile or latex gloves	N	Provides guidance for soil sample collection for samples that will be analyzed for PFAS via LC/MS/MS in compliance with QSM 5.3 (or the most recent version of the QSM for which laboratory has been certified at the time of sampling) for Navy CLEAN projects under Contract N62470-16-D-9000.
SOP CH2M-7	Management of Liquid Waste Containing Per- and Polyfluoroalkyl Substances (PFAS) Rev. Jan. 2021	CH2M	Drum, tank, HDPE sample bottles	N	Provides guidelines for managing liquid waste containing PFAS or Navy CLEAN projects under Contract N62470-16-D-9000.
SOP CH2M-8	Surface Water Sampling for Per- and Polyfluoroalkyl Substances Rev. Jan 2021	CH2M	Sample jars (HDPE bottle with HDPE screw cap), laboratory pre-filled HDPE bottles containing field blank water, loose leaf paper without waterproof coating, clip board, pen (not Sharpie), nitrile or latex gloves	N	Provides guidelines for surface water sample collection for samples that will be analyzed for PFAS via LC/MS/MS in compliance with QSM 5.3 (or the most recent version of the QSM for which the lab is certified at the time of analysis). For Navy CLEAN projects under Contract N62470-16-D-9000.
SOP CH2M-9	Sediment Sampling for Per- and Polyfluoroalkyl Substances Rev. Jan 2021	CH2M	Sample jars (HDPE bottle with HDPE screw cap), laboratory pre-filled HDPE bottles containing field blank water, loose leaf paper without waterproof coating, clip board, pen (not Sharpie), nitrile or latex gloves	N	Provides guidelines for sediment sample collection for samples that will be analyzed for PFAS via LC/MS/MS in compliance with QSM 5.3 (or the most recent version of the QSM for which the lab is certified at the time of analysis). For Navy CLEAN projects under Contract N62470-16-D-9000.
SOP I-A-1	Planning Field Sampling Activities, Rev. Feb. 2015	NAVFAC Northwest	None	N	Establishes SOPs for planning and scheduling field sampling activities.

Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP I-A-7	IDW Management, Rev. Feb. 2015	NAVFAC Northwest	None	N	Describes activities and responsibilities of NAVFAC Northwest and its subcontractors regarding management of IDW. Field activities will deviate slightly from the SOP to eliminate use of PFAS-containing materials.
SOP I-A-9	General Field Operation, Rev. Feb. 2015	NAVFAC Northwest	All field equipment	N	Defines organization and structure of sample collection, identification, record keeping, field measurements, and data collection.
SOP I-A-10	Monitoring/Sampling Location Recording, Rev. Feb. 2015	NAVFAC Northwest	Loose leaf paper and clipboard	N	Establishes guidelines for generating information to be recorded for each physical location where sampling is conducted.
SOP I-A-11	Sample Naming, Rev. Feb. 2015	NAVFAC Northwest	None	N	Describes the naming convention to be used for samples collected, analyzed, and reported for NAVFAC Northwest projects.
SOP 1-C-1	Monitoring Well/Piezometer Installation, Rev. Mar. 2015	NAVFAC Northwest	Drilling equipment	N	Describes the methods by which NAVFAC Northwest field personnel and their contractors will conduct groundwater monitoring well installation.
SOP 1-C-2	Monitoring Well Development, Rev. Mar. 2015	NAVFAC Northwest	Drilling equipment	N	Describes the methods by which NAVFAC Northwest field personnel and their contractors will conduct groundwater monitoring well development.

Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP 1-C-5	Low-Flow Groundwater Purging and Sampling, Rev. Mar. 2015	NAVFAC Northwest	Bladder or peristaltic pump, water level and water quality meters, and buckets. Compressor and controller required for bladder pump.	N	Describes the conventional groundwater monitoring well sampling procedures to be used by all NAVFAC Northwest personnel and contractors.
SOP 1-D-5	Water Level Measurements, Rev. Mar. 2015	NAVFAC Northwest	Water level meters	N	Establishes standard protocols for all NAVFAC Northwest field personnel for use in making water level measurements.
SOP I-D-7	Field Parameter Measurements, Rev. Mar. 2015	NAVFAC Northwest	Water quality meters	N	Provides instructions for the calibration, use, and checking of instruments and equipment for field measurements. Field activities will deviate slightly from the SOP to eliminate use of PFAS-containing materials.
SOP I-G-1	Land Surveying, Rev. Aug. 2014	NAVFAC Northwest	Surveying equipment	N	Describes the methods by which NAVFAC Northwest field personnel and their contractors will conduct land surveying.
SOP III-B	Field QC Samples (Water, Soil, Sediment, Tissue), Rev. Apr. 2015	NAVFAC Northwest	Sample containers	N	Describes the number and types of field QC samples that will be collected during NAVFAC Northwest site fieldwork.
SOP III-D	Logbooks, Rev. Apr. 2015	NAVFAC Northwest	Loose-leaf paper and clipboard	N	Describes the activities and responsibilities of NAVFAC Northwest personnel and/or their contractors pertaining to the identification, use, and control of logbooks and associated field data records.

Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP III-E	Record Keeping, Sample Labeling, and Chain-of-Custody Procedures, Rev. Apr. 2015	NAVFAC Northwest	Samples, sample labels, COC	N	Establishes standard protocols for all NAVFAC Northwest field personnel and their contractors for use in maintaining field and sampling activity records, writing sample logs, labeling samples, ensuring that proper sample custody procedures are used, and completing chain-of-custody/analytical request forms.
SOP III-F	Sample Containers and Preservation, Rev. Apr. 2015	NAVFAC Northwest	Sample containers	N	Sets forth the methods for use by NAVFAC Northwest field personnel and their contractors engaged in handling, storing, and transporting water, soil, or sediment samples. Field activities will deviate slightly from the SOP to eliminate use of PFAScontaining materials.
SOP III-G	Sample Handling, Storage, and Shipping, Rev. Apr. 2015	NAVFAC Northwest	Samples	N	Sets forth the methods for use by NAVFAC Northwest field personnel and their contractors engaged in handling, storing, and transporting water, soil, or sediment samples. Field activities will deviate slightly from the SOP to eliminate use of PFAScontaining materials.
SOP III-I	Equipment Decontamination, Rev. Apr. 2015	NAVFAC Northwest	Non-disposable sampling equipment	Υ	Describes general methods of equipment decontamination for use by NAVFAC Northwest field personnel and their contractors during field sampling activities. Field activities will deviate slightly from the SOP to eliminate use of PFAS-containing materials. Rinse water will be certified PFAS-free.

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Reference Number	Title, Revision Date and/or Number	Originating Organization of Sampling SOP	Equipment Type	Modified for Project Work? (Y/N)	Comments
SOP III-J	Equipment Calibration, Operation, and Maintenance, Rev. Apr. 2015	NAVFAC Northwest	Field meters	N	Describes the activities and responsibilities of the NAVFAC Northwest personnel pertaining to the operation, calibration, and maintenance of equipment used to collect environmental data. Field activities will deviate slightly from the SOP to eliminate use of PFAScontaining materials.
N/A	Final Environmental Restoration Program Recordkeeping Manual, Feb. 2017	NAVFAC	None.	N	Provides instructions on how to submit documents for inclusion in the program's Environmental Document Management System and information about compiling, documenting, managing, and maintaining CERCLA Environmental Restoration Administrative Record Files; Post-Decision Files; Site Files; and Petroleum, Oil, and Lubricant Files.

SAP Worksheet #22—Field Equipment Calibration, Maintenance, Testing, and Inspection Table

Field Equipment	Activity ¹	Frequency	Acceptance Criteria	CA	Resp. Person	SOP Reference ²	Comments
Horiba U-22	Calibration	Daily before		Clean probe with deionized water and calibrate again.	FTL	SOP-007	Appendix B
pH probe	Calibration	use	(±) 3%	Do not use instrument if not able to calibrate properly.	TIL	301-007	Аррениіх в
Horiba U-22 Specific	Calibration	Daily before	Conductivity reads 4.49 ±	Clean probe with deionized water and calibrate again.	FTL	SOP-007	Appendix B
conductance probe	Calibration	use	not able properly	Do not use instrument if not able to calibrate properly.	FIL	30F-007	Аррениіх в
Horiba U-22	6.111	Daily before		Clean probe with deionized water and calibrate again.	FTL	SOP-007 Appendi	Annondiy P
Turbidity probe	Calibration	use	Turbidity reads 0 ± 3%	Do not use instrument if not able to calibrate properly.	FIL		Аррепаіх в
Horiba U-22 DO		esting Daily before use	Consistent with the current atmospheric	Clean probe with deionized water and calibrate again.	F.T.	COD 007	
Temperature Probes	Testing		pressure and ambient temperature	Do not use instrument if not able to calibrate properly.	FTL	SOP-007	Appendix B
	Maintenance - Check mechanical and electronic parts,	Daily before	Stable readings after 3 minutes	Clean probe with deionized water and		SOP-007	
Horiba U-22	verify system continuity, check battery, and clean probes	tem use, at the end of the day, and	pH reads 4.0 ± 3% Conductivity reads 4.49 ± 3%	calibrate again. Do not use instrument if not able to calibrate	FTL		Appendix B
	Calibration check		Turbidity reads 0 ± 3%	properly.			

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SAP Worksheet #22—Field Equipment Calibration, Maintenance, Testing, and Inspection Table (continued)

Field Equipment	Activity ¹	Frequency	Acceptance Criteria	CA	Resp. Person	SOP Reference ²	Comments
			Isobutylene reads 100 ppm				Appendix B
	Calibrate using ambient air and isobutylene 100 parts per million (ppm) calibration gas	Daily and as needed	Methane reads 50 percent lower explosive level	Manufacturer technical	FTL	SOP CH2M-3, SOP-III-J	
Multi RAE PID			Oxygen reads 20.9 percent Hydrogen sulfide reads 25 ppm	support for calibration errors			
			Carbon monoxide reads 50 ppm				
Groundwater sampling pumps and tubing	Inspect pumps, tubing, and air/sample line quick-connects	Regularly	Maintained in good working order according to manufacturer's recommendations	Replace items	FTL	SOP-III-J	Appendix B

¹ Activities may include calibration, verification, testing, and maintenance.

² Specify the appropriate reference letter or number from the Project Sampling SOP References table (**Worksheet #21**).

SAP Worksheet #23—Analytical SOP References Table

Lab SOP Number	Title, Revision Date, and/or Number	Definitive or Screening Data	Matrix and Analytical Group	Instrument	Organization Performing Analysis	Modified for Project Work (Y/N)
5-370-12	Extraction of Poly and Perfluoroalkyl Substances from Environmental Matrices, 04/06/2021, Rev. 12	Definitive	Soil/Groundwater/Ditch Surface Soil/Surface Water/PFAS	N/A	Battelle	N
5-369-08	Analysis of Poly and Perfluoroalkyl Substances in Environmental Samples by Liquid Chromatography and Tandem Mass Spectrometry (LC-MS/MS), 07/28/2020, Rev. 8	Definitive	Soil/Groundwater/Ditch Surface Soil/Surface Water/PFAS	LC/MS/MS	Battelle	N
6-010-19	Sample Receipt, Custody, and Handling, 12/15/2020, Rev. 19	N/A	Soil/Groundwater/Ditch Surface Soil/Surface Water/PFAS	N/A	Battelle	N
5-291-17	Determination of Method Detection Limits in the Analytical Laboratory, 09/13/2019, Rev. 17	N/A	Soil/Groundwater/Ditch Surface Soil/Surface Water/PFAS	N/A	Battelle	N

Note:

Laboratory SOPs meet DoD QSM 5.3 (ASD, 2021, Appendix D) requirements for Battelle.

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Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	CA	Person Responsible for CA	SOP Reference
			Solid Phase Extraction (SPE) must be used unless samples are known to contain high PFAS concentrations (e.g., AFFF formulations). Inline SPE is acceptable.			
	Aqueous Sample	Each sample and associated batch QC	Entire sample plus bottle rinsate must be extracted using SPE.	N/A		
	Preparation	samples	Known high PFAS concentration samples require serial dilution be performed in duplicate.			
			Documented project approval is needed for samples prepared by serial dilution as opposed to SPE.			
LC/MS/MS	Solid Sample Preparation	Each sample and associated batch QC samples	Entire sample received by the laboratory must be homogenized prior to subsampling.	N/A	- Analyst/Laboratory	5-369-08 DoD QSM 5.3
(PFAS)	Sample	Each sample and associated batch QC samples	ENVI-CarbTM or equivalent must be used	NI/A	PM	
	Cleanup Procedure	Not applicable to AFFF and AFFF Mixture Samples	on each sample and batch QC sample.	N/A		
		Instrument must have a valid mass	Calibrate the mass scale of the MS with calibration compounds and procedures described by the manufacturer.			
	Mass	calibration prior to any sample analysis	Mass calibration range must bracket the ion masses of interest. The most recent mass calibration must be used for every	If the mass calibration fails, then recalibrate. If it fails again, consult manufacturer		
	Calibration	Mass calibration is verified after each mass calibration, prior	acquisition in an analytical run.	instructions on corrective maintenance.		
		to initial calibration (ICAL)	Mass calibration must be verified to be ± 0.5 atomic mass unit of the true value, by acquiring a full scan continuum mass spectrum of a PFAS stock standard.			

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	CA	Person Responsible for CA	SOP Reference
	Mass Spectral Acquisition Rate	Each analyte, Extracted Internal Standard Analyte	A minimum of 10 spectra scans are acquired across each chromatographic peak.	N/A		
			Standards containing both branched and linear isomers must be used when commercially available.			
LC/MS/MS (PFAS) Calibration.		PFAS method analytes may consist of both branched and linear isomers, but quantitative standards that contain the linear and branched isomers do not exist for all method analytes.		Analyst/	5-369-08	
(cont.)	Calibration, Calibration Verification, and Spiking Standards	All analytes	For PFAS that do not have a quantitative branched and linear standard, identify the branched isomers by analyzing a qualitative standard that includes both linear and branched isomers and determine retention times, transitions and transition ion ratios. Quantitate samples by integrating the total response (i.e., accounting for peaks that are identified as linear and branched isomers) and relying on the ICAL that uses the linear isomer quantitative standard.	N/A	Analyst/ Laboratory PM	DoD QSM 5.3

The chemical derivation of the ion transitions must be documented. A minimum of two ion transitions (Precursor → quant ion and precursor →	Responsible for CA	Reference
LC/MS/MS (PFAS) (cont.) Sample PFAS Identification All analytes detected in a sample N/A N/A N/A N/A	Analyst/ Laboratory PM	5-369-08 DoD QSM 5.3

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	CA	Person Responsible for CA	SOP Reference
			In order to avoid biasing results high due to known interferences for some transitions, the following transitions must be used for the quantification of the following analytes:			
LC/MS/MS (PFAS)	lon Transitions (Precursor-> Product)	Every field sample, standard, blank, and QC sample.	PFOA: $413 \rightarrow 369$ PFOS: $499 \rightarrow 80$ PFHxS: $399 \rightarrow 80$ PFBS: $299 \rightarrow 80$ 4:2 FTS: $327 \rightarrow 307$ 6:2 FTS: $427 \rightarrow 407$ 8:2 FTS: $527 \rightarrow 507$ NEtFOSAA: $584 \rightarrow 419$ NMeFOSAA: $570 \rightarrow 419$	N/A	Analyst/ Laboratory PM	5-369-08 DoD QSM 5.3
(cont.)			If these transitions are not used, the reason must be technically justified and documented (e.g., alternate transition was used due to observed interferences).			3.0
	Peak Asymmetry	With each calibration	First two eluting peaks in a mid-level calibration standard must have an asymmetry factor between 0.8 and 1.5.	When the asymmetry factor does not pass, perform CA to address the issue. Modification of the standard or extract composition to more aqueous content is not permitted.		

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	CA	Person Responsible for CA	SOP Reference
	Calibration, Calibration Verification, and Spiking Standards	All analytes	Standards containing both branched and linear isomers must be used when commercially available. If not available, the total response of the analyte must be integrated (i.e., accounting for peaks that are identified as linear and branched isomers) and quantitated using a calibration curve which includes the linear isomer only for that analyte (e.g., PFOA).	N/A		
LC/MS/MS (PFAS) (cont.)	Ion Transitions (Parent-> Product)		The chemical derivation of the ion transitions, both those used for quantitation and those used for confirmation, must be documented. Two transitions and the ion transition ratio per analyte shall be monitored and documented with the exception of PFBA and PFPeA. In order to avoid biasing results high due to known interferences for some transitions, the following transitions must be used for the quantification of the following analytes:		Analyst/ Laboratory PM	5-369-08 DoD QSM 5.3
			PFOA: 413 —> 369 PFOS: 499 —> 80 PFHxS: 399 —> 80 PFBS: 299 —> 80 4:2 FTS: 327 —> 307 6:2 FTS: 427 —> 407 8:2 FTS: 527 —> 507 NEtFOSAA: 584 —> 419 NMeFOSAA: 570 —> 419	N/A		
			If these transitions are not used, the reason must be technically justified and documented (e.g., alternate transition was used due to observed interferences).			

SAP Worksheet #24—Analytical Instrument Calibration Table

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	CA	Person Responsible for CA	SOP Reference
	Instrument Blank	Following highest calibration point		If acceptance criteria are not met after the highest calibration standard, calibration must be performed using a lower concentration for the highest standard until acceptance criteria is met.		
LC/MS/MS (PFAS) (cont.)			≤ 1/2 the LOQ	If acceptance criteria are not met after the highest standard which is not included in the calibration, the standard cannot be used to determine the highest concentration in samples at which carryover does not occur.	Analyst/ Laboratory PM	
				If acceptance criteria are not met after sample, additional instrument blanks must be analyzed until acceptance criteria are met. Additional samples shall not be analyzed until acceptance criteria are met.		5-369-08 DoD QSM 5.3
				Correct problem, rerun ISC. If problem persists, repeat		
		Prior to analysis and at least once every 12 hours.	Analyte concentrations must be at LOQ; concentrations must be within ± 30% of their true values.	ICAL. No samples shall be analyzed until ISC has met acceptance criteria.		
				ISC can serve as the initial daily continuing calibration verification (CCV).		

Notes:

The specifications in this table meet the requirements of DoD QSM 5.3.

NEtFOSAA = n-ethylperfluoro-1-octancesulfonamidoacetic acid

NMeFOSAA = n-methylperfluoro-1-octanesulfonamidoacetic acid

SAP Worksheet #25—Analytical Instrument and Equipment Maintenance, Testing, and Inspection Table

Instrument/ Equipment	Maintenance Activity	Testing Activity	Inspection Activity	Frequency	Acceptance Criteria	Corrective Action	Responsible Person	SOP Reference
LC/MS/MS	AM	PFAS	N/A	6 Months	N/A	N/A	Analyst/supervisor	5-369
Balance	Verification	Weight	N/A	Daily	± 0.02 gram or ± 0.1% of calibration weight used (whichever is greater)	Refer to manufacturer's instruction manual	Analyst/supervisor	3-160
Balance	Calibration	Weight	N/A	Annually	Per manufacturer	Remove from service, repair, replace	Analyst/supervisor	3-160
Pipette	Verification	Volume	N/A	Daily	± 2% difference from true value, <1% relative standard deviation (n=3)	Remove from service, repair, replace	Analyst/supervisor	3-181
Pipette	Calibration	Volume	N/A	Quarterly	Per manufacturer	Remove from service, repair, replace	Analyst/supervisor	3-181

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SAP Worksheet #26—Sample Handling System

SAMPLE COLLECTION, PACKAGING, AND SHIPMENT

Sample Collection (Personnel/Organization): Project Field Team, FTL/CH2M. Field SOPs are in Appendix B.

Sample Packaging (Personnel/Organization): Project Field Team, FTL/CH2M. Field SOPs are in Appendix B.

Coordination of Shipment (Personnel/Organization): FTL/CH2M

Type of Shipment/Carrier: FedEx Priority Overnight: Samples will be shipped directly to Battelle

SAMPLE RECEIPT AND ANALYSIS

Sample Receipt (Personnel/Organization): Sample Receiving/Battelle

Sample Custody and Storage (Personnel/Organization): Sample Receiving/Battelle

Sample Preparation (Personnel/Organization): Sample Preparation Staff/Battelle

Sample Determinative Analysis (Personnel/Organization): Battelle

SAMPLE ARCHIVING

Field Sample Storage (No. of days from sample collection): 60 days from receipt

Sample Extract/Digestate Storage (No. of days from extraction/digestion): 28 days after extraction/digestion

Biological Sample Storage (No. of days from sample collection): N/A

SAMPLE DISPOSAL

Personnel/Organization): Sample Disposal/Battelle

Number of Days from Analysis: 60 days after final sample results are reported, unless there is a hold on a particular sample or previous arrangements have been made

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SAP Worksheet #27—Sample Custody Requirements

Field Sample Custody Procedures (sample collection, packaging, shipment, and delivery to laboratory):

Field sample collection procedures will follow the PFAS SOPs (listed in **Worksheet #21**), and team members will use PFAS free materials while sampling. Samples will be collected by field team members under the supervision of the FTL. As samples are collected, they will be placed into containers and labeled. Labels will be taped to the jar to ensure labels do not separate from the jars. Samples will be cushioned with packaging material and placed into coolers containing enough ice to keep the samples 0 to 6° C (but not frozen; requirements for USEPA 537.1 are less than 10°C for the first 48 hours) until they are received by the laboratory.

The chain-of-custody record will be placed into the cooler in a resealable zip-top plastic bag. Coolers will be taped up and shipped to the laboratories via FedEx overnight, with the air bill number indicated on the chain of custody (to relinquish custody). Upon delivery, the laboratory will log each cooler and report the status of the samples to CH2M.

See Worksheet #21 for SOPs containing sample custody guidance.

The CH2M field team will ship all environmental samples directly to the laboratory performing the analysis. This will require shipment to Battelle.

Laboratory Sample Custody Procedures (receipt of samples, archiving, disposal):

Laboratory custody procedures can be found in the laboratory SOPs, which will be provided upon request.

Sample ID Procedures:

Sample labels will include, at a minimum, client name, site, sample ID, date/time collected, analysis group or method, preservation, and sampler's initials. The field notes will identify the sample ID with the location and time collected and the parameters requested. The laboratory will assign each field sample a laboratory sample ID based on information in the chain of custody. The laboratory will send sample log-in forms to the CH2M PC to check that sample IDs and parameters are correct.

Chain-of-Custody Procedures:

Chain-of-custody records will include, at a minimum, laboratory contact information, client contact information, sample information, and relinquished by/received by information. Sample information will include sample ID. Date/time collected, number and type of containers, preservative information, analysis method, and comments. The chain-of-custody record will link location of the sample from the field notes to the laboratory receipt of the sample. The laboratory will use the sample information to populate the Laboratory Information Management Systems database for each sample.

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SAP Worksheet #28—Laboratory QC Sample Table

Matrix: Soil/Groundwater
Analytical Group: PFAS

Analytical Method/SOP Reference: PFAS by LC/MS/MS Compliant with QSM 5.3 Table B-15/SOP 5-369-08

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for CA	Data Quality Indicator	Measurement Performance Criteria
Extracted Internal Standard Analytes	Every field sample, standard, blank, and QC sample.	Added to solid sample prior to extraction. Added to aqueous samples, into the original container, prior to extraction. For aqueous samples prepared by serial dilution instead of SPE, added to final dilution of samples prior to analysis. Extracted Internal Standard Analyte recoveries must be within 50% to 150% of ICAL midpoint standard area or area measured in the initial CCV on days when an ICAL is not performed.	Correct problem. If required, reextract and reanalyze associated field and QC samples. If recoveries are acceptable for QC samples, but not field samples, the field samples must be re-extracted and analyzed (greater dilution may be needed). Samples may be reextracted and analyzed outside of hold times, as necessary for CA associated with QC failure.		Precision/ Accuracy/Bias	
Method Blank	One per preparatory batch.	No analytes detected > 1/2 LOQ or > 1/10 the amount measured in any sample or > 1/10 regulatory limit, whichever is greater.	Correct problem. If required, reextract and reanalyze method blank and all QC samples and field samples processed with the contaminated blank. Samples may be reextracted and analyzed outside of hold times, as necessary for CA associated with QC failure. Examine the project specific requirements. Contact the client as to additional measures to be taken.		Bias/ Contamination	

SAP Worksheet #28—Laboratory QC Sample Table (continued)

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for CA	Data Quality Indicator	Measurement Performance Criteria
		Blank spiked with all analytes at a concentration ≥LOQ and ≤ the mid-level calibration concentration.	Correct problem, then re-extract and reanalyze the LCS and all samples in the associated preparatory batch for failed analytes if sufficient sample material is available.			
Laboratory Control Sample	One per preparatory batch.	DoD QSM 5.3 limits (Worksheet #15) If the analyte(s) are not listed, use in-house LCS limits if project limits are not specified.	Samples may be reextracted and analyzed outside of hold times, as necessary for CA associated with QC failure. Examine the project specific requirements. Contact the client as to additional measures to be taken.		Precision/ Accuracy/Bias	
Matrix Spike	One per preparatory batch. Not required for aqueous samples prepared by serial dilution instead of SPE.	Sample spiked with all analytes at a concentration ≥LOQ and ≤ the mid-level calibration concentration. DoD QSM 5.3 limits (See Worksheet #15 for control limits) If the analyte(s) are not listed, use in-house LCS limits if project limits are not specified.	Examine the project specific requirements. Contact the client as to additional measures to be taken. For the specific analyte(s) in the parent sample, apply J-flag if acceptance criteria are not met and explain in the Case Narrative.		Precision/ Accuracy/Bias	
Matrix Spike Duplicate or Matrix Duplicate (MD)	For MSD: One per preparatory batch. For MD: Each aqueous sample prepared by serial dilution instead of SPE.	For MSD: Sample spiked with all analytes at a concentration ≥ LOQ and ≤ the mid-level calibration concentration. DoD QSM 5.3 limits (See Worksheet #15 for control limits). If the analyte(s) are not listed, use in-house LCS limits if project limits are not specified. RPD ≤ 30% (between MS and MSD or sample and MD).	Examine the project specific requirements. Contact the client as to additional measures to be taken. For the specific analyte(s) in the parent sample, apply J-flag if acceptance criteria are not met and explain in the Case Narrative.		Precision/ Accuracy/Bias	

SAP Worksheet #28—Laboratory QC Sample Table (continued)

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for CA	Data Quality Indicator	Measurement Performance Criteria
Post Spike Sample	Only applies to aqueous samples prepared by serial dilution instead of SPE that have reported value of " <loq" analyte(s).<="" for="" td=""><td>Spike all analytes reported as < LOQ into the dilution that the result for that analyte is reported from. The spike must be at the LOQ concentration to be reported for this sample as < LOQ. When analyte concentrations are calculated as < LOQ, the post spike for that analyte must recover within 70 to 130% of its true value.</td><td>When analyte concentrations are calculated as < LOQ, and the spike recovery does not meet the acceptance criteria, the sample, sample duplicate, and post spike sample must be reanalyzed at consecutively higher dilutions until the criteria is met.</td><td></td><td>Bias/ Contamination</td><td></td></loq">	Spike all analytes reported as < LOQ into the dilution that the result for that analyte is reported from. The spike must be at the LOQ concentration to be reported for this sample as < LOQ. When analyte concentrations are calculated as < LOQ, the post spike for that analyte must recover within 70 to 130% of its true value.	When analyte concentrations are calculated as < LOQ, and the spike recovery does not meet the acceptance criteria, the sample, sample duplicate, and post spike sample must be reanalyzed at consecutively higher dilutions until the criteria is met.		Bias/ Contamination	
LOD verification	Quarterly for every analyte.	Spike a quality system matrix at concentration 2 to 4x the DL. Must meet 3:1 signal-to-noise ratio, or for data systems that do not measure noise, results must be at least 3 standard deviations greater than the mean method blank concentration.	If verification fails, the DL determination must be repeated and a LOD verification. Alternatively pass two consecutive LOD verification at a higher spike and at the LOD at the higher concentration.		Accuracy	
LOQ verification	Quarterly for every analyte.	Spike a quality system matrix at a concentration equal to or greater than the low point of the calibration curve.	Must meet laboratory-specified precision and bias limits. If LOQ fails, repeat at a higher level until limits are met.		Precision/Bias	

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SAP Worksheet #29—Project Documents and Records Table

Document		Where Maintained
Field Notebooks	•	Field data deliverables (e.g., field notes entries, chains
Chain-of-Custody Records		of custody, air bills, and EDDs) will be kept on CH2M's network server.
Air Bills	•	Field parameter data will be loaded with the analytical
Telephone Logs		data into the Navy database.
Custody Seals	•	Analytical laboratory hardcopy deliverables and DV reports will be saved on the network server and
CA Forms		archived per the Navy CLEAN contract.
Electronic data deliverables (EDDs)	•	Electronic data from the laboratory will be loaded into
ID of QC Samples		Navy database.
Meteorological Data from Field	•	Following project completion, hardcopy deliverables (e.g., field notes, chains of custody) will be archived at
Sampling Instrument Calibration Logs		Iron Mountain:
Sampling Locations and Sampling Plan		Iron Mountain Headquarters 745 Atlantic Avenue
Sampling Notes and Drilling Logs		Boston, MA 02111
Water Quality Parameter		(800) 899-IRON
Sample Receipt, Chain of Custody, and Tracking Records	•	Following project completion, hardcopy deliverables, including chains of custody and raw data, will be archived at the Washington National Records Center:
Standard Traceability Logs		Washington National Records Center
Equipment Calibration Logs		4205 Suitland Road
Sample Preparation Logs		Suitland, Maryland 20746-8001 301-778-1550
• Run Logs		
• Equipment Maintenance, Testing, and Inspection Logs		
• CA Forms		
Reported Field Sample Results		
 Reported Result for Standards, QC Checks, and QC Samples 		
 Instrument printouts (raw data) for Field Samples, Standards, QC Checks, and QC Samples 		
Data Package Completeness Checklists		
Sample disposal records		
Extraction/Clean-up Records		
Raw Data (archived per Navy CLEAN contract)		
• DV Reports		
• CA Forms		
Laboratory QA Plan		
Field Performance Audit Checklists		

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SAP Worksheet #30—Analytical Services Table

Matrix	Analytical Group	Sample Locations/ID	Analytical Method	Data Package Turnaround Time	Laboratory/ Organization	Backup Laboratory/ Organization ³
Groundwater				3 Days/28 Days ²	Battelle Analytical Services	
Soil			LC/MS/MS in	3 Days/28 Days ²	141 Longwater Drive Suite 202	
Surface Water (if present)	PFAS	Refer to Worksheet #18	compliance with DoD QSM 5.3, Table B-15 ¹	3 Days/28 Days ²	Norwell, MA 02061 POC: Jonathan Thorn	Vista Analytical
Ditch Surface Soil				3 Days/28 Days ²	(781) 681-5565	

Analytical method is compliant with DoD QSM 5.3 Table B-15 or the most recent version of the DoD QSM for which Battelle has DoD ELAP certification.

A 3-day expedited turn-around time will be required for all groundwater grab samples and select soil samples as detailed in **Worksheet #11**; all other samples will be analyzed at the standard 28-day turn-around time.

³ If the backup laboratory is used for analysis, applicable SAP worksheets will be submitted to the Navy QAO for review and approval.

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SAP Worksheet #31—Planned Project Assessments Table

Assessment Type	Frequency	Internal or External	Organization Performing Assessment	Person(s) Responsible for Performing Assessment (title and organizational affiliation)	Person(s) Responsible for Responding to Assessment Findings (title and organizational affiliation)	Person(s) Responsible for Identifying and Implementing CA (title and organizational affiliation)	Person(s) Responsible for Monitoring Effectiveness of CA (title and organizational affiliation)
Field Performance	One during	Internal	CH2M	AM	FTL	PM	PM
Audit	sampling event	iliterilai	CHZIVI	CH2M	CH2M	CH2M	CH2M
Field Document	Daily during	Internal	CH2M	PM or TM	FTL	PM	PM
Review	sampling event	internal	CITZIVI	CH2M	CH2M	CH2M	CH2M

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SAP Worksheet #32—Assessment Findings and Corrective Action Responses

Assessment Type	Nature of Deficiencies Documentation	Individual(s) Notified of Findings (name, title, organization)	Timeframe of Notification	Nature of CA Response Documentation	Individual(s) Receiving CA Response (name, title, organization)	Timeframe for Response
Field Performance Audit	Checklist and Written Audit Report	TBD, FTL, CH2M	Within 1 day of audit	Verbal and Memorandum	FTL CH2M	Within 1 day of receipt of CA Form
Field Document Review	Markup copy of field documentation	TBD, FTL, CH2M	Within 1 day of review	Verbal and Memorandum	FTL CH2M	Within 1 day of receipt of markup
Offsite Laboratory Technical Systems Audit	TBD by Perry Johnson Laboratory Accreditation, Inc.	TBD, Battelle	Within 2 months of audit	Memorandum	TBD by Perry Johnson Laboratory Accreditation, Inc.	Within 2 months of receipt of initial notification

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SAP Worksheet #32-1—Laboratory Corrective A	ction Form
Person initiating Corrective Action (CA):	Date:
Description of problem and when identified:	
Course of marklane if he same an arrange start.	
Cause of problem, if known or suspected:	
Sequence of CA (including date implemented, action	planned, and personnel/data affected):
CA implemented by:	Date:
CA initially approved by:	
Follow-up date:	
Final CA approved by:	Date:
Information copies to:	
Anita Dodson, CH2M Navy CLEAN Program Chemist	

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SAP Worksheet #32-2—Field Performance Audit Checklist

Project	: Responsibilit	ies	
Project	No.:		Date:
Project	Location:		Signature:
Team I	Members		
Yes	No	1)	Is the approved work plan being followed? Comments
Yes	No	2)	Was a briefing held for project participants? Comments
Yes	No	3)	Were additional instructions given to project participants? Comments
Sample Yes	e Collection	1)	Is there a written list of sampling locations and descriptions?
			Comments
Yes	No	2)	Are samples collected as stated in the Master SOPs? Comments
Yes	No	3)	Are samples collected in the type of containers specified in the work plan? Comments
Yes	No	4)	Are samples preserved as specified in the work plan? Comments
Yes	No	5)	Are the number, frequency, and type of samples collected as specified in the work plan? Comments

SAP vvorksneet #3/-/—Fleid Performance Augit Checklist (continued	e Audit Checklist (continued)	SAP Worksheet #32-2—Field Performan
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Yes	No	6)	Are QA checks performed as specified in the work plan? Comments
Yes	No	7)	Are photographs taken and documented? Comments
Documen	t Control		
Yes	No	1)	Have any accountable documents been lost? Comments
Yes	No	2)	Have any accountable documents been voided? Comments
Yes	No	3)	Have any accountable documents been disposed of? Comments
Yes	No	4)	Are the samples identified with sample tags? Comments
Yes	No	5)	Are blank and duplicate samples properly identified? Comments
Yes	No	6)	Are samples listed on a chain-of-custody record? Comments
Yes	No	7)	Is chain of custody documented and maintained? Comments

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SAP Worksheet #33—QA Management Reports Table

Type of Report	Frequency (daily, weekly monthly, quarterly, annually, and so forth)	Projected Delivery Date(s)	Person(s) Responsible for Report Preparation (title and organizational affiliation)	Report Recipient(s) (title and organizational affiliation)
Field Audit Report	One during sampling event	TBD	PM CH2M	Included in project files
QA Management Report/Technical Memorandum	Once results have been assessed for data usability	To be submitted with Final RI Report	СН2М РС	NAVFAC Northwest RPM and will be posted in project file

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SAP Worksheet #34-36—Data Verification and Validation (Steps I and IIa/IIb) Process Table

Data Review Input	Description ¹	Responsible for Verification or Validation	Step I/IIa/ IIb ²	Internal/ External ³
Field Notebooks	Field notebooks will be reviewed internally and placed into the project file for archival at project closeout.	FTL/CH2M	Step I	Internal
Chains of Custody and Shipping Forms	Chain-of-custody forms and shipping documentation will be reviewed internally upon their completion and verified against the packed sample coolers they represent. The shipper's signature on the chain-of-custody forms will be initialed by the reviewer, a copy of the chain-of-custody forms retained in the site file, and the original and remaining copies taped inside the cooler for shipment. Chain-of-custody forms will also be reviewed for adherence to the SAP by the PC.	FTL/CH2M PC/CH2M	Step I	Internal and External
Sample Condition upon Receipt	Any discrepancies and missing or broken containers will be communicated to the PC in the form of laboratory logins.	PC/CH2M	Step I	External
Documentation of Laboratory Method Deviations	Laboratory method deviations not included in the lab SOP and therefore not included in the DoD ELAP accreditation letter are not permitted for this project work. Any laboratory method deviation not covered under the current DoD ELAP accreditation letter will be resubmitted to the accrediting body for review and approval prior to sample collection and resubmitted to Navy QAO for review.	PC/CH2M	Step I	External
EDDs	EDDs will be compared against hardcopy laboratory results (10 percent check). If errors are found during the 10% check, an additional 25% of the EDDs will be checked against hardcopy laboratory results.	PC/CH2M	Step I	External
Case Narrative	Case narratives will be reviewed by the DV during the DV process. This is verification that case narratives were generated and applicable to the data packages.	DV/EDS	Step I	External
Laboratory Data	All laboratory data packages will be verified internally by the laboratory performing the work for completeness and technical accuracy prior to submittal.	Laboratory QAO	Step I	Internal
Laboratory Data	The data will be verified for completeness by the PC. To ensure completeness, EDDs will be compared to the SAP. This is a verification that all samples were included in the laboratory data and that correct analyte lists were reported.	PC/CH2M	Step I	External
Audit Reports	Upon report completion, a copy of all audit reports will be placed in the site file. If CAs are required, a copy of the documented CA taken will be attached to the appropriate audit report in the QA site file. Periodically, and at the completion of site work, site file audit reports and CA forms will be reviewed internally to ensure that all appropriate CAs have been taken and that CA reports are attached. If CAs have not been taken, the site manager will be notified to ensure action is taken.	PM/CH2M PC/CH2M	Step I	Internal

SAP Worksheet #34-36—Data Verification and Validation (Steps I and IIa/IIb) Process Table (continued)

Data Review Input	Description ¹	Responsible for Verification or Validation	Step I/IIa/	Internal/ External ³
CA Reports	CA reports will be reviewed by the PC or PM and placed into the project file for archival at project closeout.	PM/CH2M PC/CH2M	Step I	External
Laboratory Methods	During the pre-validation check, ensure that the laboratory analyzed samples using the correct methods specified in the SAP. If methods other than those specified in the SAP were used, the reason will be determined and documented.	PC/CH2M	Step IIa	External
Target Compound List and Target Analyte list	During the pre-validation check, ensure that the laboratory reported all analytes from each analysis group in accordance with Worksheet #15 . If the target compound list is not correct, then it must be corrected prior to sending the data for validation. Once the checks are complete, the PM is notified via email	PC/CH2M	Step IIa	External
Laboratory Limits (DL/LOD/LOQ)	During the pre-validation check, the laboratory limits (DL/LOD/LOQ) will be compared to those listed in the project SAP. If limits were not met, the laboratory will be contacted and asked to provide an explanation, which will then be discussed in the associated project report. Oftentimes, the cause for minor laboratory limit deviation from those presented in the SAP is a result of the quarterly update of the laboratory LOD.	PC/CH2M	Step IIb	External
Laboratory SOPs	Ensure that approved analytical laboratory SOPs were followed. Any discrepancies will be discussed first in the data validation narrative and will be included in the associated project report.	Laboratory QAO	Step I	Internal
Sample Chronology	Holding times from collection to extraction or analysis and from extraction to analysis will be considered during the data validation process.	DV/EDS	Step IIa and IIb	External
Raw Data	Ten percent review of raw data to confirm laboratory calculations. For a recalculated result, the DV attempts to recreate the reported numerical value. The laboratory is asked for clarification if a discrepancy is identified that cannot reasonably be attributed to rounding. In general, this is outside of a 5 percent difference. Conduct a 10 percent review of laboratory calculations. For a recalculated result, the DV attempts to recreate the reported numerical value. The laboratory is asked for clarification if a discrepancy is found, which cannot be reasonably attributed to rounding. If errors are found during the 10 percent check, an additional 20 percent of the raw data will be checked to confirm calculations. Any discrepancies will be addressed in the data validation narrative.	DV/EDS	Step IIa	External
Onsite Screening	All nonanalytical field data will be reviewed against SAP requirements for completeness and accuracy based on the field calibration records. Screening data will be included in the project report.	FTL/CH2M	Step IIb	Internal

SAP Worksheet #34-36—Data Verification and Validation (Steps I and IIa/IIb) Process Table (continued)

Data Review Input	Description ¹	Responsible for Verification or Validation	Step I/IIa/ IIb ²	Internal/ External ³
Documentation of Method QC Results	Establish that all required QC samples were run and met limits. Any deviations will be reported in the data validation narrative.	DV/EDS	Step IIa	External
Documentation of Field QC Sample Results	Establish that all required QC samples were run and met limits and discuss QC sampling in the associated project report.	PC/CH2M	Step IIa	Internal
DoD ELAP Evaluation	Ensure that each laboratory is DoD ELAP-certified for the analyses they are to perform. Ensure evaluation timeframe does not expire.	PC/CH2M	Step I	External
Analytical data for PFAS analyzed for soil, groundwater and surface water ⁴	Analytical methods and laboratory SOPs as presented in this SAP will be used to evaluate compliance against QA/QC criteria. Should adherence to QA/QC criteria yield deficiencies, data may be qualified. Data may be qualified if QA/QC exceedances have occurred. Guidance and qualifiers from <i>United States Department of Defense General Data Validation Guidelines</i> (DoD, 2019b), <i>Data Validation Guidelines Module 3: Data Validation Procedure for Per- and Polyfluoroalkyl substances Analysis by QSM Table B-15</i> (DoD, 2020), and <i>Per- and Polyfluoroalkyl Substances (PFAS): Reviewing Analytical Methods for Environmental Samples</i> Technical brief (USEPA, 2019) will be applied as appropriate. If specific guidance is not given for these methods in the General Data Validation Guidelines, the DV may adapt the guidance from <i>USEPA National Functional Guidelines for Superfund Organic Methods Data Review</i> (USEPA, 2017).	DV/EDS	Step IIa and IIb	External

Notes:

- Should CH2M find discrepancies during the verification or validation procedures above, an email documenting the issue will be circulated to the internal project team, and a Corrections to File Memo will be prepared identifying the issues and the CA needed. This memo will be sent to the laboratory, or applicable party, and maintained in the project file.
- ² Verification (Step I) is a completeness check that is performed before the data review process continues to determine whether the required information (complete data package) is available for further review. Validation (Step IIa) is a review that the data generated is in compliance with analytical methods, procedures, and contracts. Validation (Step IIb) is a comparison of generated data against MPC in the SAP (both sampling and analytical).
- ³ Internal or external is in relation to the data generator.
- Stage 4 data validation will be performed on 10 percent of all definitive analyses, which will include recalculated results from the raw data to verify calculations. The remaining (90 percent) of the definitive data will have Stage 2B data validation performed.

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SAP Worksheet #37—Usability Assessment

Summarize the usability assessment process and all procedures, including interim steps and any statistics, equations, and computer algorithms that will be used:

- Nondetected site chemicals will be evaluated to ensure that project-required PQLs in Worksheet #15 were
 achieved. If PQLs were achieved and the verification and validation steps yielded acceptable data, then the
 data are considered usable.
- During verification and validation steps, data may be qualified as estimated with the following qualifiers: J or UJ. The qualifiers represent minor QC deficiencies, which will not affect the usability of the data. When major QC deficiencies are encountered, the DV will qualify data with an X as recommended for rejection, and in most cases, data qualified with an X are not considered usable for project decisions. The data validation narrative will be reviewed along with the data quality objectives, and the X qualifier may be replaced with an R qualifier as rejected data.
 - J = Analyte present. Reported value may or may not be accurate or precise.
 - J+ = Analyte present. Reported value is estimated and may be biased high.
 - J- = Analyte present. Reported value is estimated and may be biased low.
 - UJ = Analyte not detected. Associated nondetect value may be inaccurate or imprecise.
 - R = Rejected result, team discussion. Result not reliable.
 - X = Result recommended for rejection by the validator. Result not reliable.
- The following additional qualifiers may be given by the DV:
 - N = Tentative ID. Consider Present. Special methods may be needed to confirm the chemical's presence or absence in future sampling efforts.
 - NJ = Qualitative ID questionable because of poor resolution. Chemical presumptively present at approximate quantity.
 - U = Not Detected.
- Analytical data will be checked to ensure the values and any qualifiers are appropriately transferred to the
 electronic database. The checks include comparison of hardcopy data and qualifiers to the EDD. Once the data
 have been uploaded into the electronic database, another check will be performed to ensure that all results
 were loaded accurately.
- Field and laboratory precision will be compared as RPD between the two results.
- Deviations from the SAP will be reviewed to assess whether CA is warranted and to assess impacts on achievement of project objectives.

Describe the evaluative procedures used to assess overall measurement error associated with the project:

- To assess whether enough acceptable data are available for decision making, the data will be compared to the 95 percent completeness goal and reconciled with MPC following validation and review of data quality indicators.
- If significant biases are detected with laboratory QA/QC samples, they will be evaluated to assess impacts on decision making. Low biases will be described in greater detail because they represent a possible inability to detect compounds that may be present at the site.
- If significant differences are noted between laboratory and field precision, the cause will be further evaluated to assess impacts on decision making.

SAP Worksheet #37—Usability Assessment (continued)

Describe the documentation that will be generated during the usability assessment and how usability assessment results will be presented so that they identify trends, relationships (correlations), and anomalies:

The following will be prepared by CH2M and submitted to NAVFAC Northwest for review for use in making decisions on the path forward for the site:

• Data tables will be produced to reflect detected and nondetected site analytes. Data qualifiers will be presented in the tables and discussed in the data quality evaluation, and overall results will be provided in the RI Report.

Identify the personnel responsible for performing the usability assessment:

The CH2M team, including the PM and PC, will review the data and present a usability assessment to NAVFAC Northwest for review and approval.

References

Assistant Secretary of Defense (ASD). 2021. Investigating Per- and Polyfluoroalkyl Substances within the Department of Defense Cleanup Program. September.

CH2M HILL, Inc (CH2M). 2017a. Sampling and Analysis Plan, Investigation of Perfluorinated Compounds in Drinking Water, Ault Field and Outlying Landing Field Coupeville, Naval Air Station Whidbey Island, Oak Harbor and Coupeville, Washington. January.

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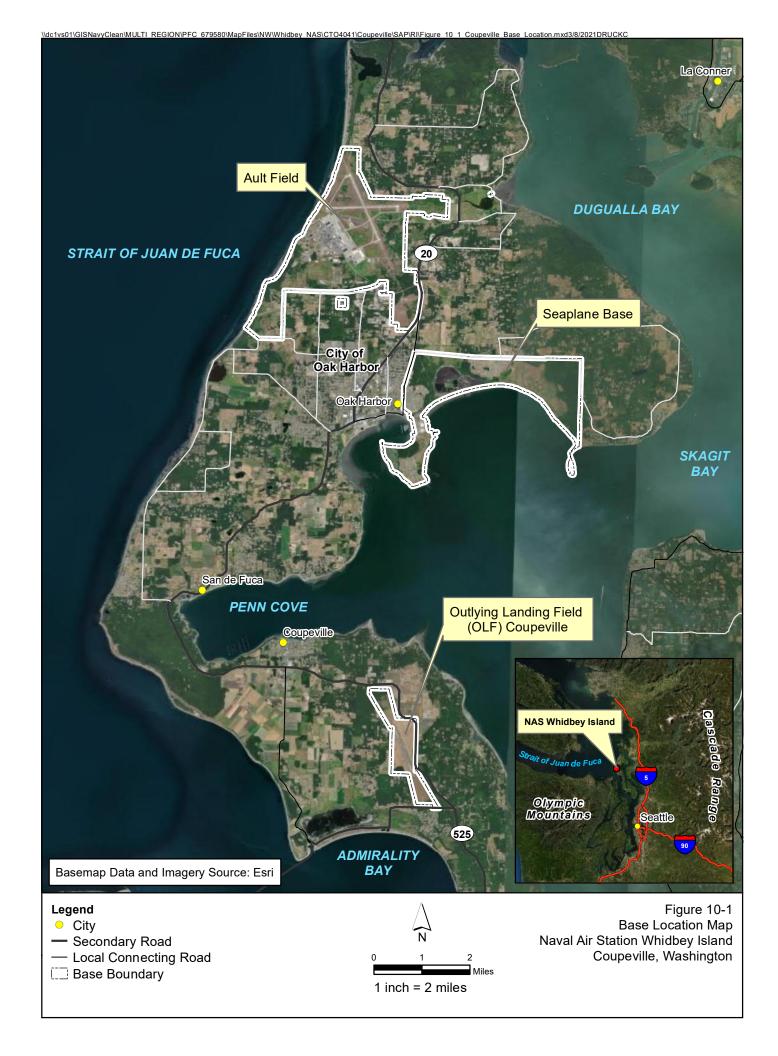
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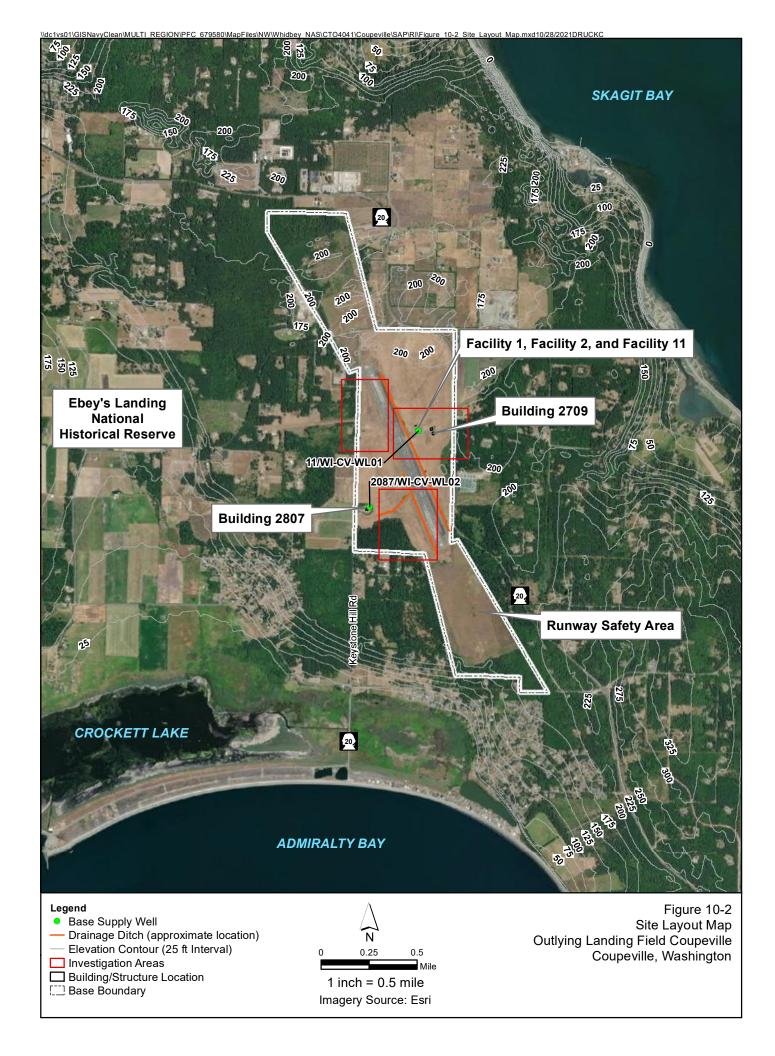
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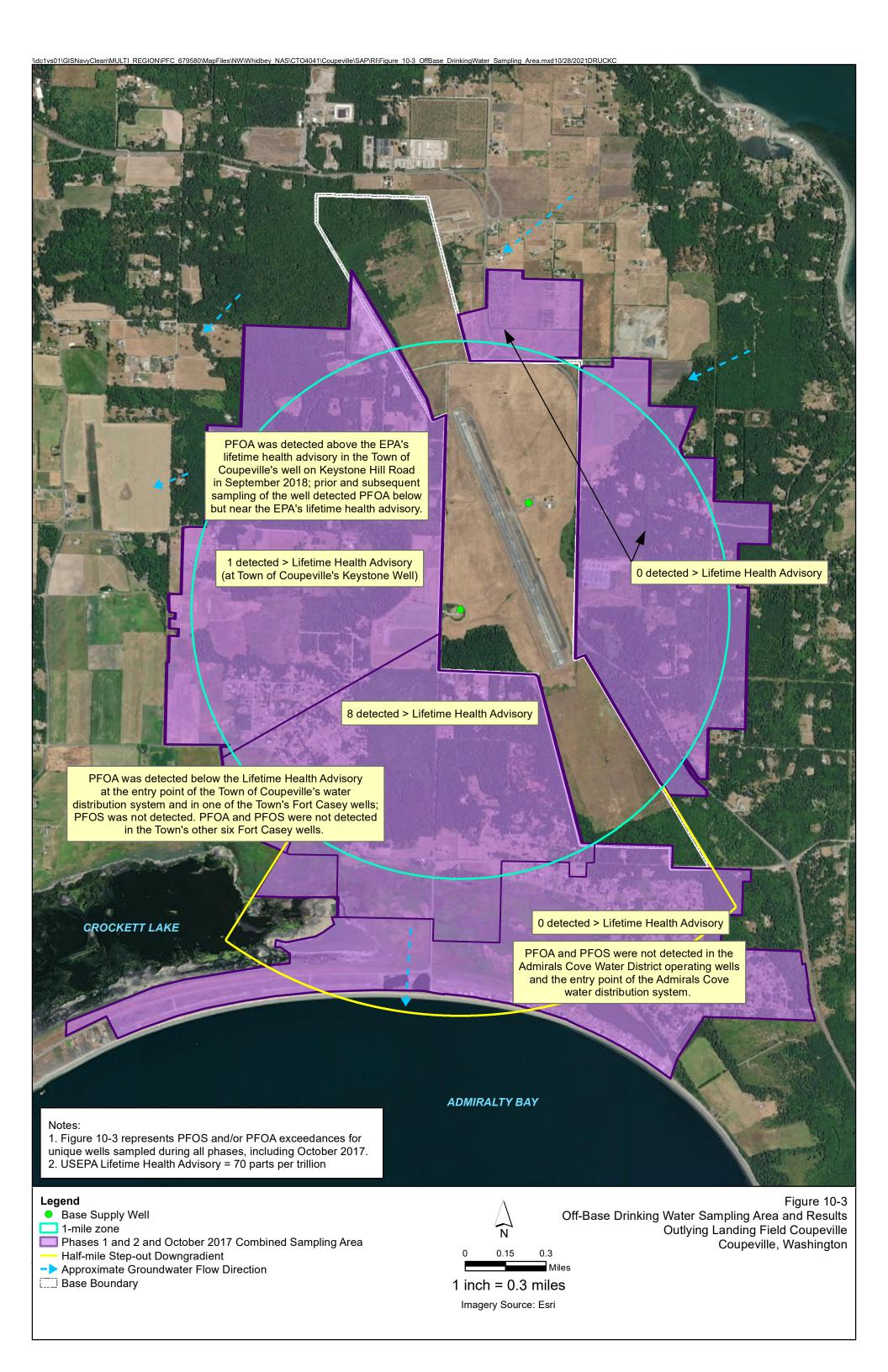
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SAMPLING AND ANALYSIS PLAN PER- AND POLYFLUOROALKYL SUBSTANCES REMEDIAL INVESTIGATION OUTLYING LANDING FIELD COUPEVILLE
NAVAL AIR STATION WHIDBEY ISLAND, OAK HARBOR, WASHINGTON
JANUARY 2022
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Figures









Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

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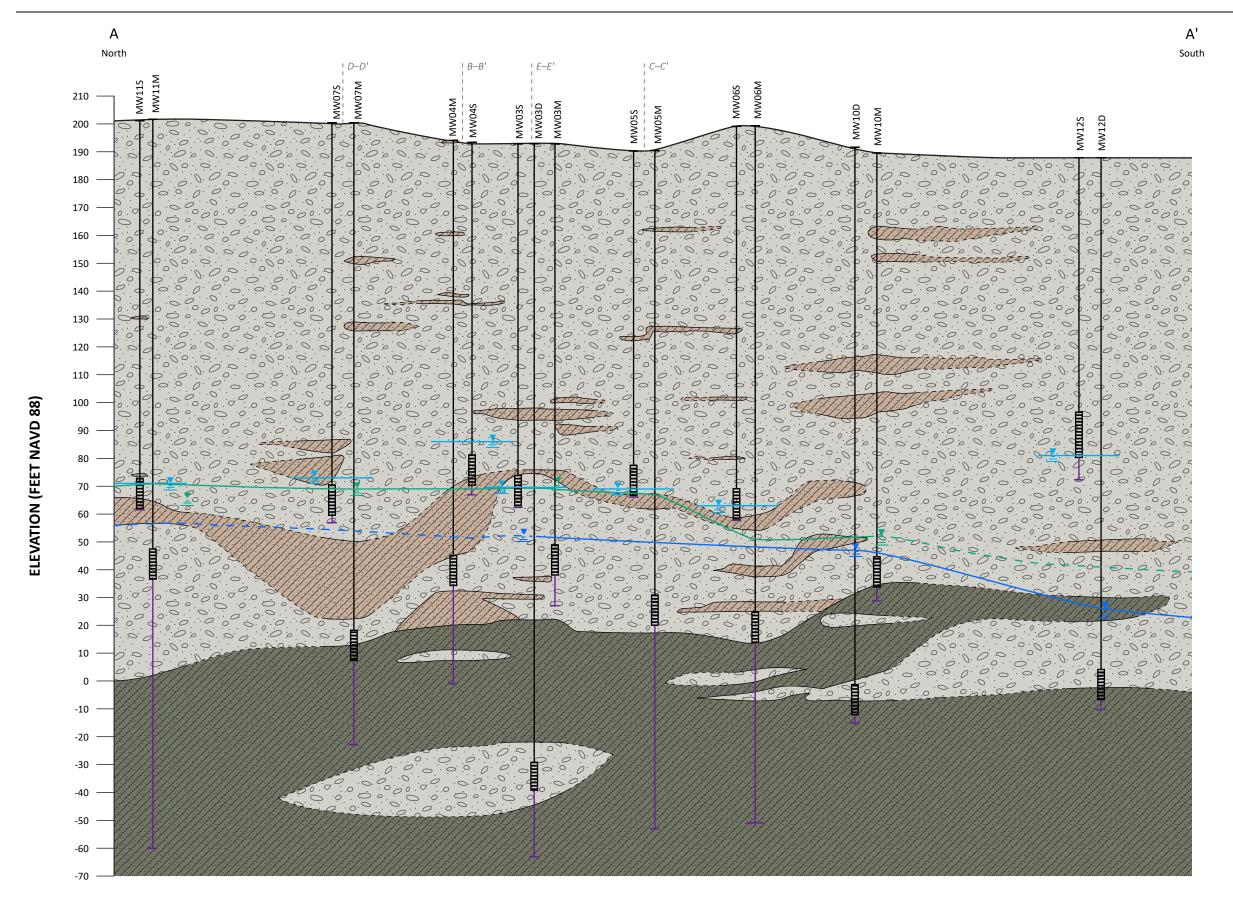
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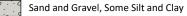
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Clay and Silt



Clay/Silt, Claystone, Some Organics and Peat



Water Table Shallow-Screened Interval (April 2020)

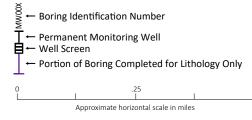


Potentiometric Surface Intermediate-Screened Interval (April 2020)



Potentiometric Surface Deep-Screened Interval

Wells



- 1. Cross section is vertically exaggerated for visualization purposes.
- 2. Cross section is conceptual in nature; predominant soil type was generally used for stratigraphic interpretation, however, in some cases similar units were combined for the purpose of simplification.
- 3. Geologic contacts are dashed where inferred.
- 4. Shallow water elevations are presumed to be noncontinuous, perched water tables.
- 5. Some wells are associated with other cross-sections. Intersections of this cross-section with other cross-sections are noted above the well labels as applicable.
- 6. NAVD 88 = North American Vertical Datum of 1988

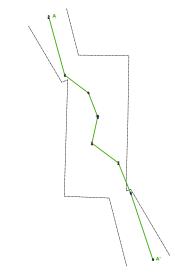
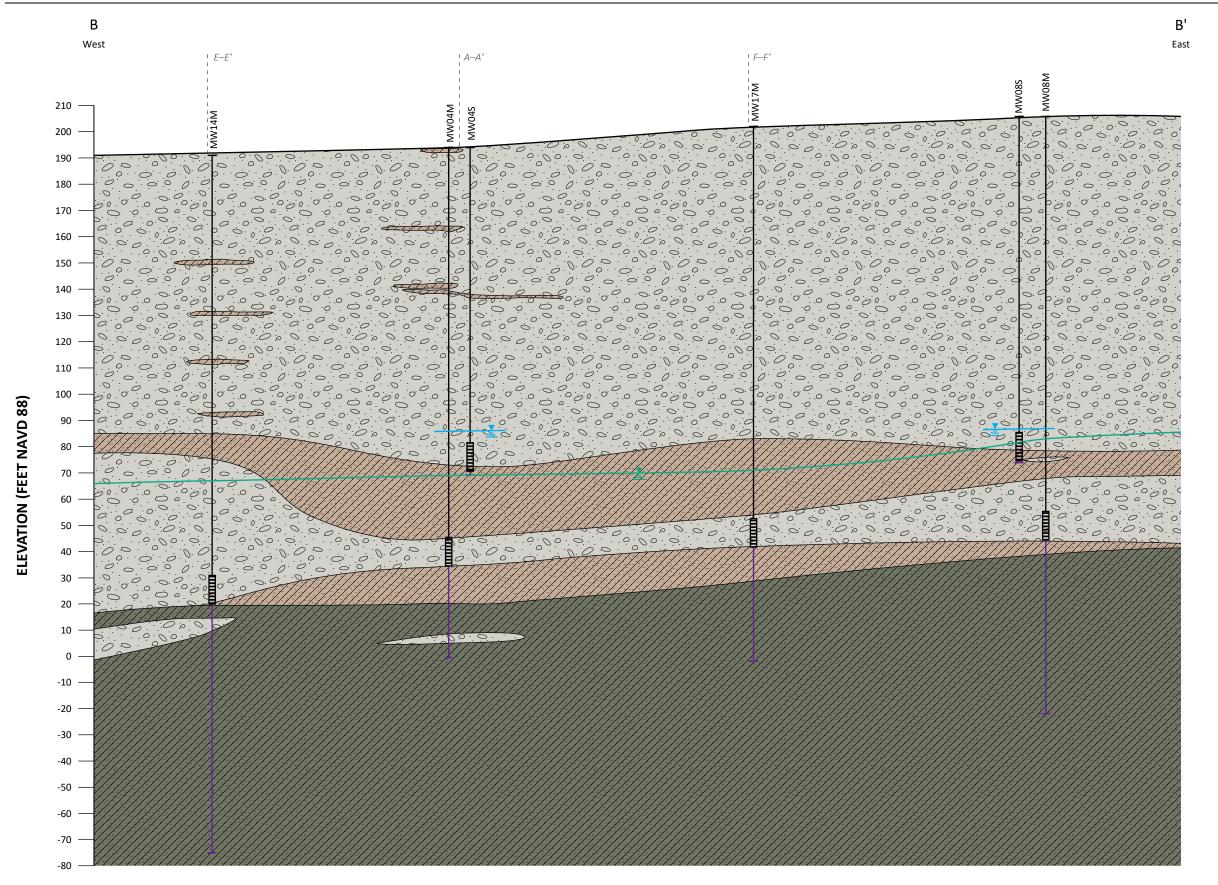
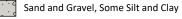


Figure 10-5 Geologic Cross Section A-A' Outlying Landing Field Coupeville Coupeville, Washington



LEGEND





Clay and Silt



Clay/Silt, Claystone, Some Organics and Peat



Water Table Shallow-Screened Interval (April 2020)



Potentiometric Surface Intermediate-Screened Interval (April 2020)

Wells





- 1. Cross section is vertically exaggerated for visualization purposes.
- 2. Cross section is conceptual in nature; predominant soil type was generally used for stratigraphic interpretation, however, in some cases similar units were combined for the purpose of simplification.
- 3. Geologic contacts are dashed where inferred.
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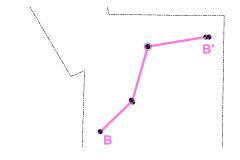
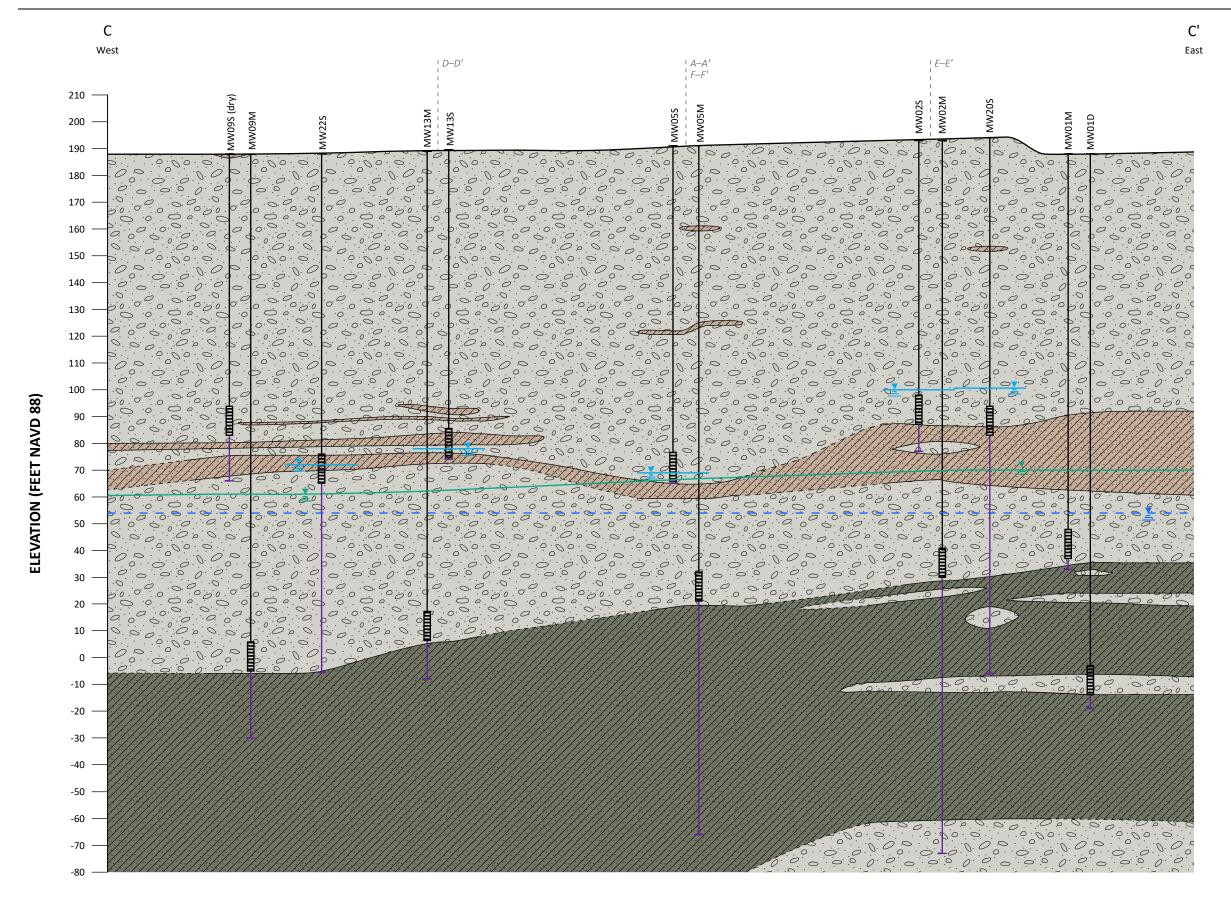
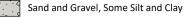
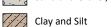


Figure 10-6 Geologic Cross Section B-B' Outlying Landing Field Coupeville Coupeville, Washington



LEGEND







Clay/Silt, Claystone, Some Organics and Peat

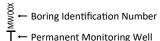


Water Table Shallow-Screened Interval (April 2020) Potentiometric Surface Intermediate-Screened



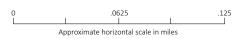
Wells

Interval (April 2020) Potentiometric Surface Deep-Screened Interval



← Well Screen

← Portion of Boring Completed for Lithology Only



Notes:

- 1. Cross section is vertically exaggerated for visualization purposes.
- 2. Cross section is conceptual in nature; predominant soil type was generally used for stratigraphic interpretation, however, in some cases similar units were combined for the purpose of simplification.
- 3. Geologic contacts are dashed where inferred.
- 4. Shallow water elevations are presumed to be noncontinuous, perched water tables.
- 5. Some wells are associated with other cross-sections. Intersections of this cross-section with other cross-sections are noted above the well labels as applicable.
- 6. NAVD 88 = North American Vertical Datum of 1988



Figure 10-7 **Geologic Cross Section C-C'** Outlying Landing Field Coupeville Coupeville, Washington



Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

NOTIFICATION: FIGURE 10-8 CONTAINS SENSITIVE BUT UNCLASSIFIED INFORMATION WHICH IS PROTECTED BY THE FREEDOM OF INFORMATION ACT

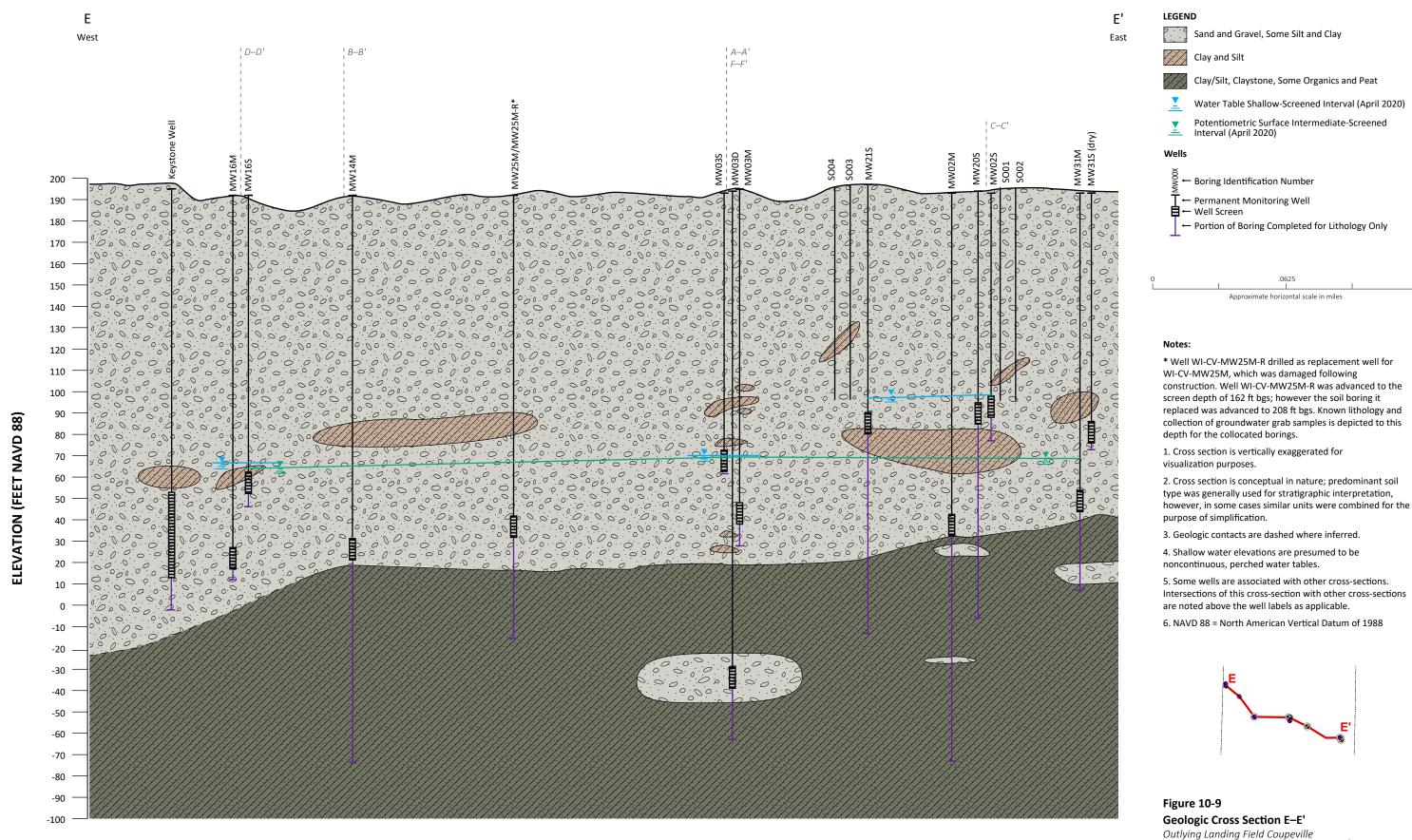
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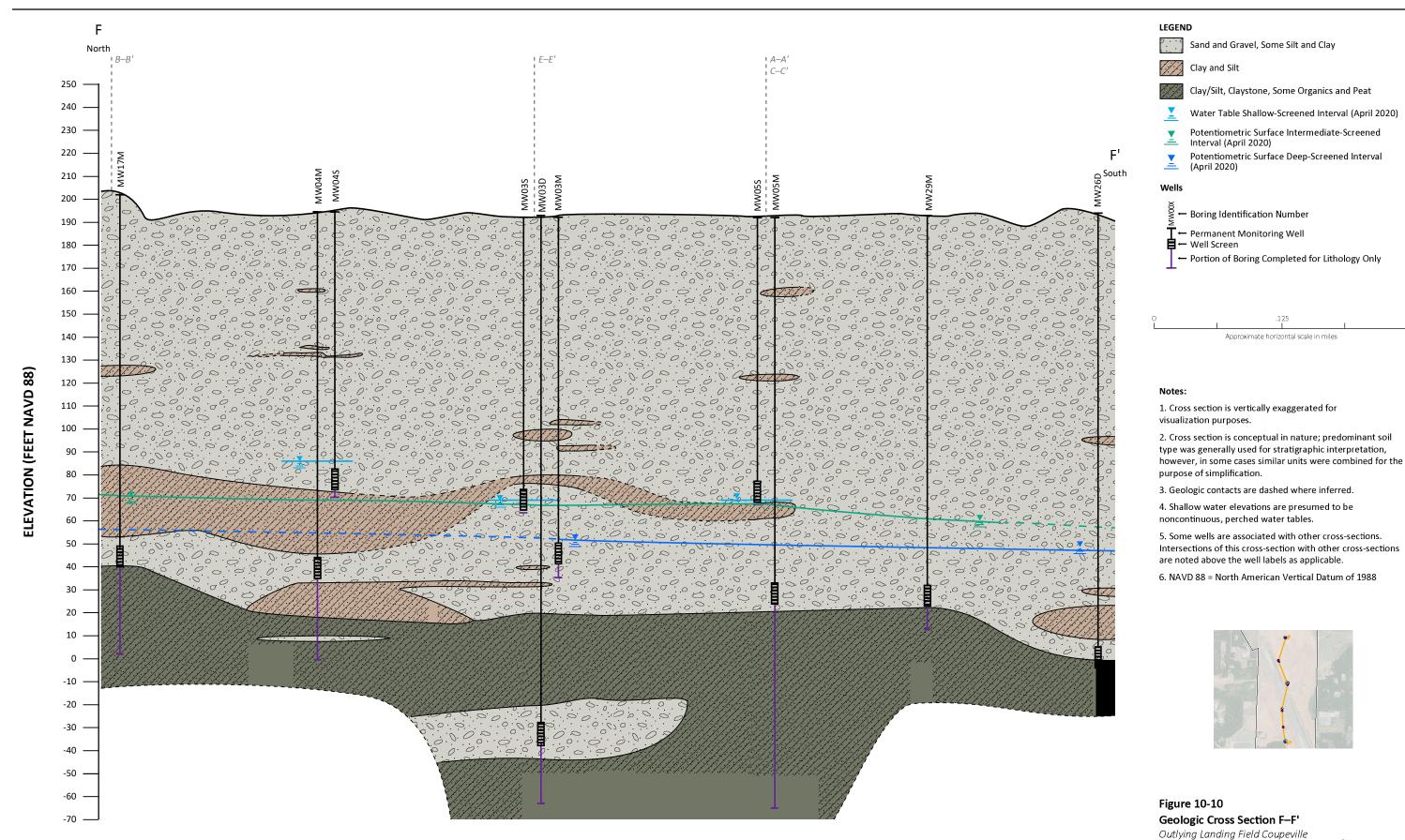
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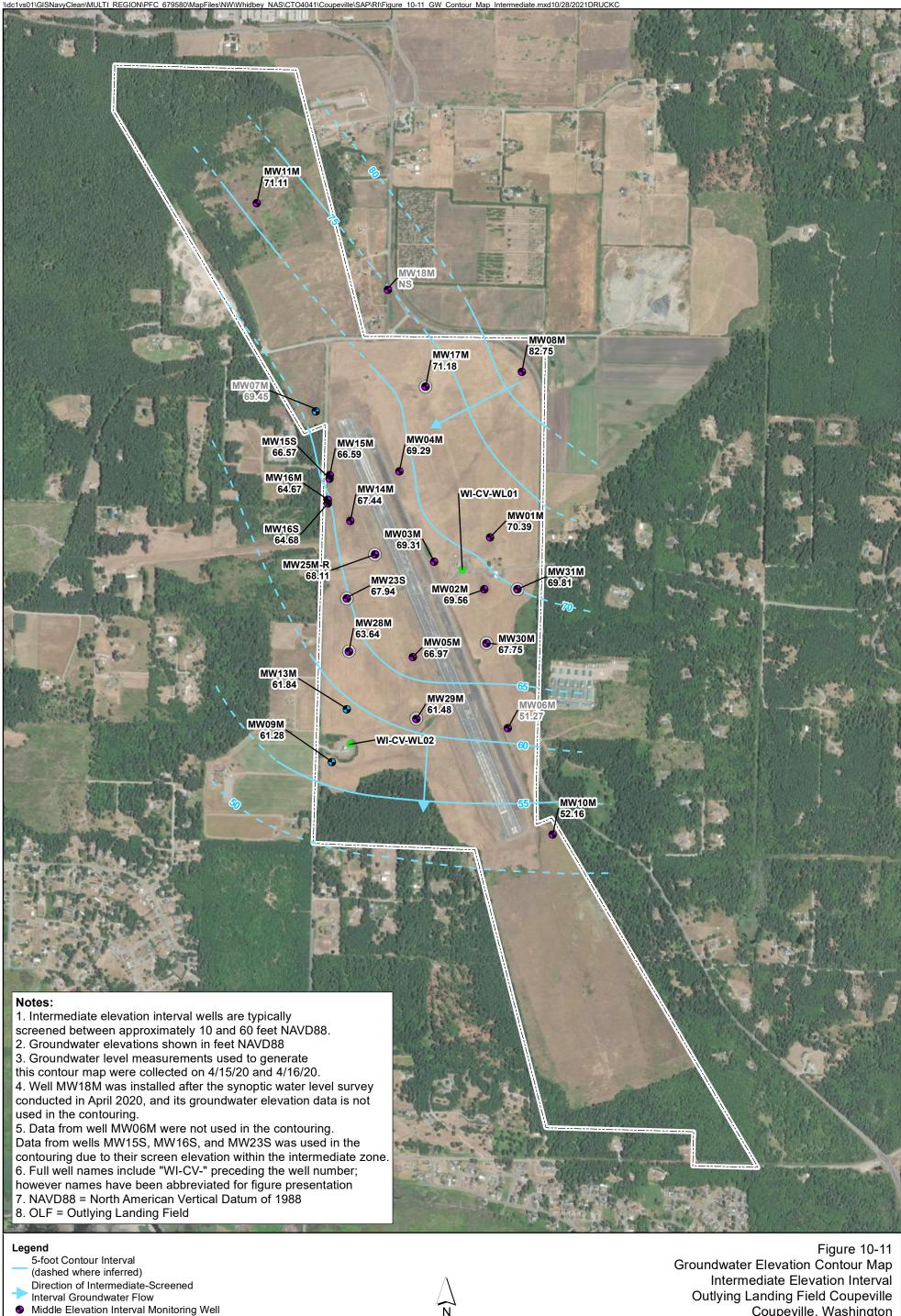
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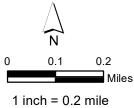
Coupeville, Washington



Coupeville, Washington

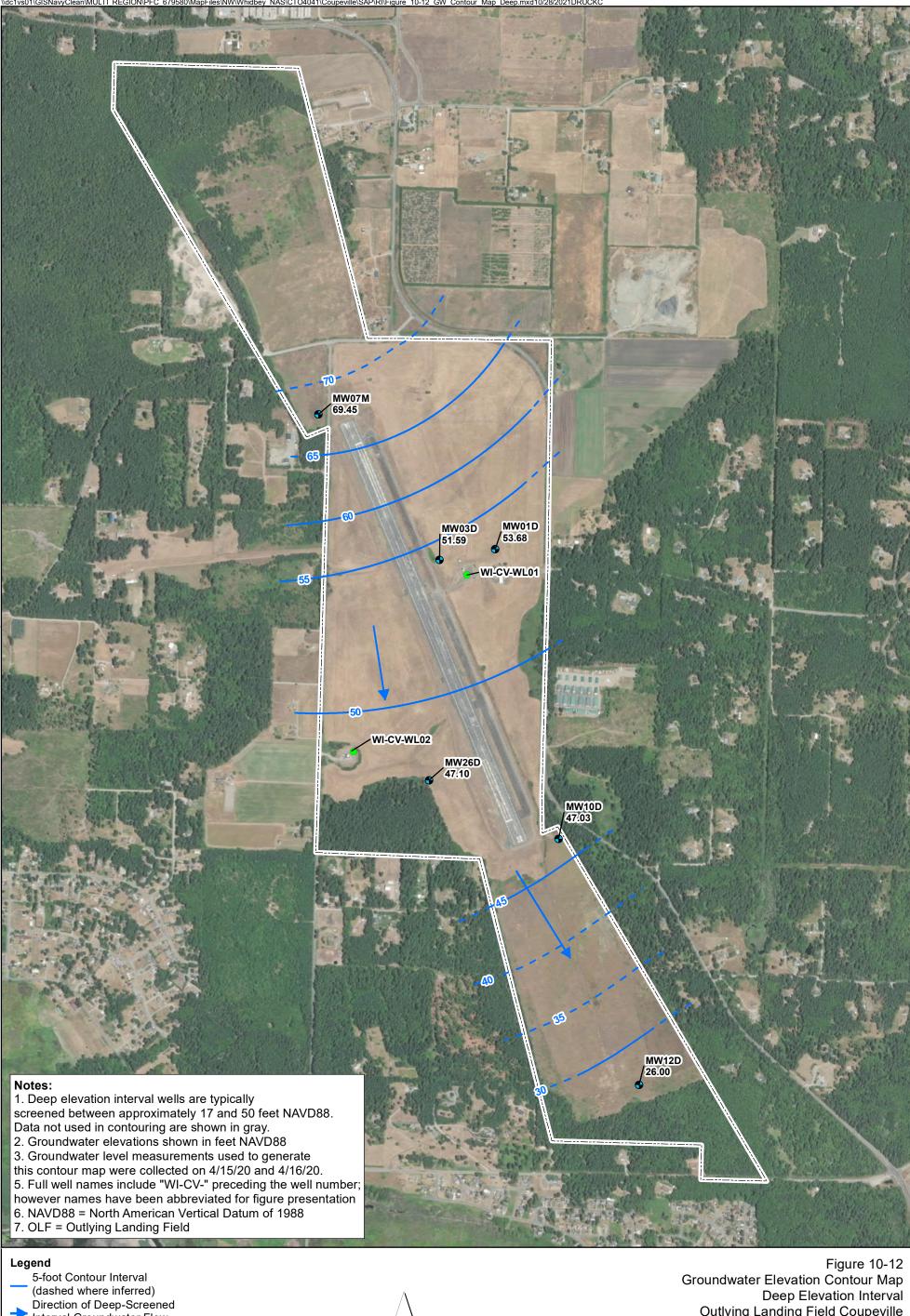


- Deep Elevation Interval Monitoring Well
 Base Supply Well
 Supplemental Site Investigation
 Monitoring Well Installation Base Boundary

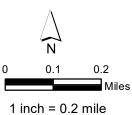


Imagery Source: Esri

Coupeville, Washington

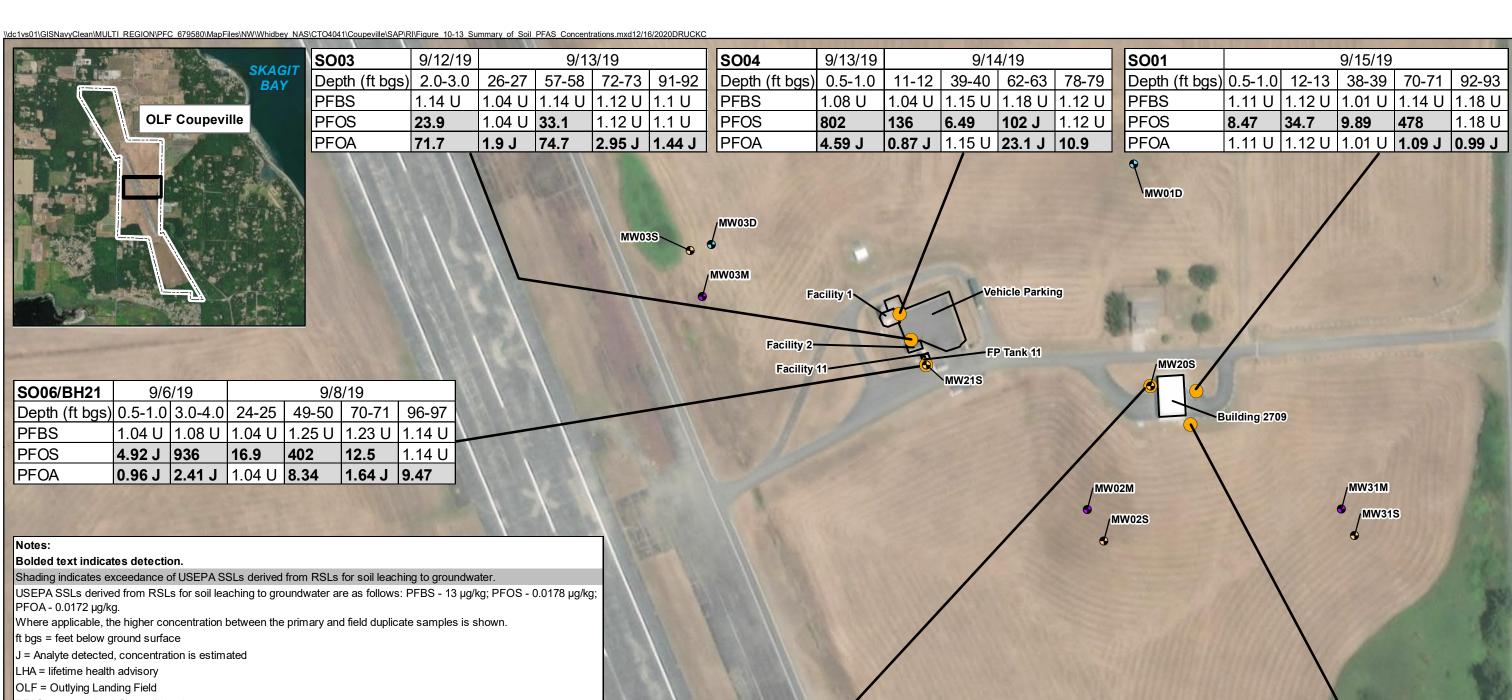


- Interval Groundwater Flow
- Deep Elevation Interval Monitoring Well
- Base Supply Well
- Supplemental Site Investigation Monitoring Well Installation
- Base Boundary



Imagery Source: Esri

Outlying Landing Field Coupeville Coupeville, Washington



PFAS = per- and polyfluoroalkyl substances

PFBS = perfluorobutanesulfonic acid

PFOA = perfluorooctanoic acid

PFOS = perfluorooctane sulfonate

RSL = regional screening level

SSL = soil screening level
U = not detected

USEPA = United States Environmental Protection Agency

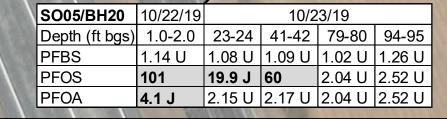
units = micrograms per kilogram (µg/kg)

Legend

- Shallow Elevation Interval Monitoring Well
- Middle Elevation Interval Monitoring Well
- Deep Elevation Interval Monitoring Well

Soil Boring

☐ Building/Structure Location



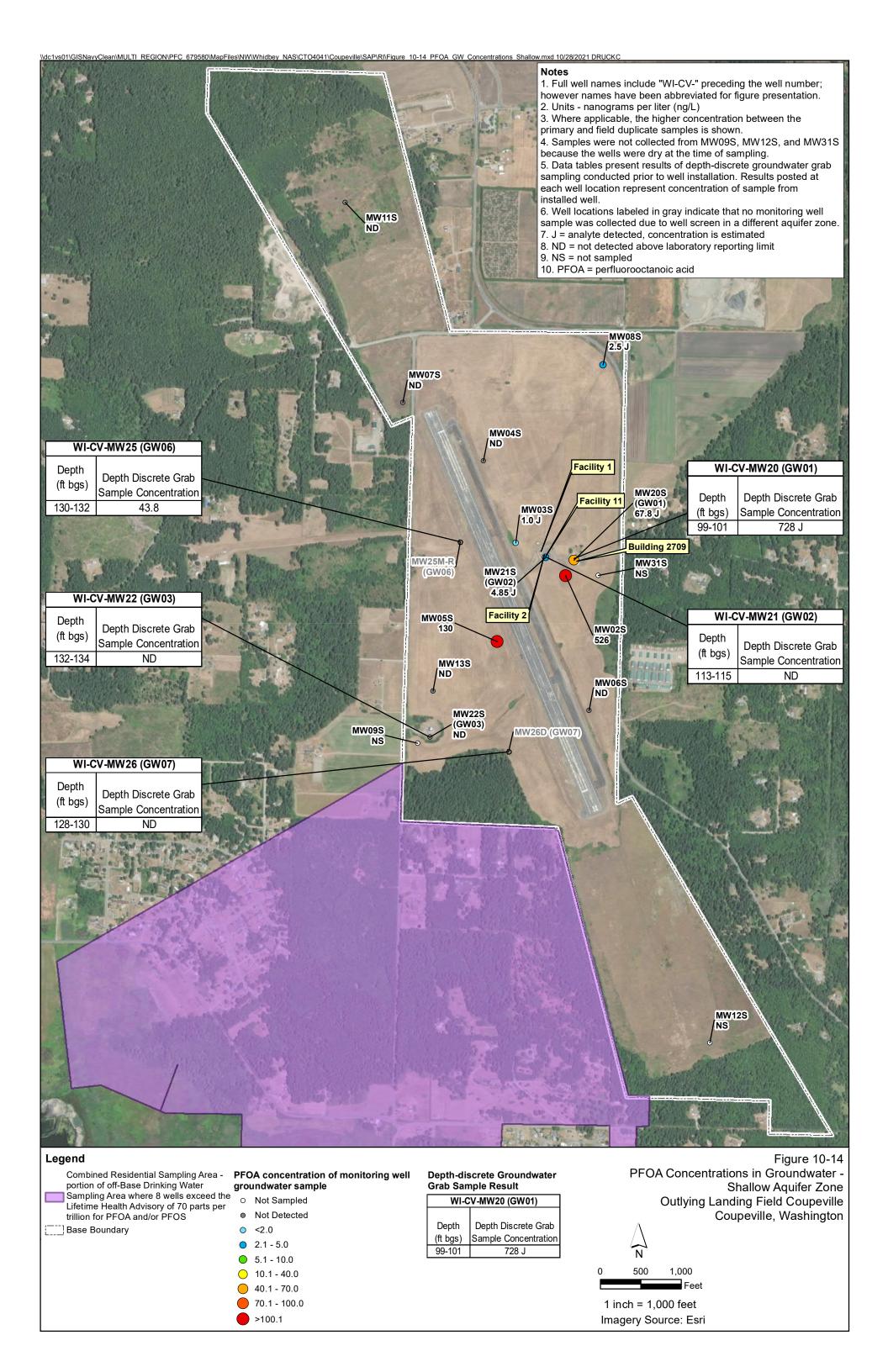
150

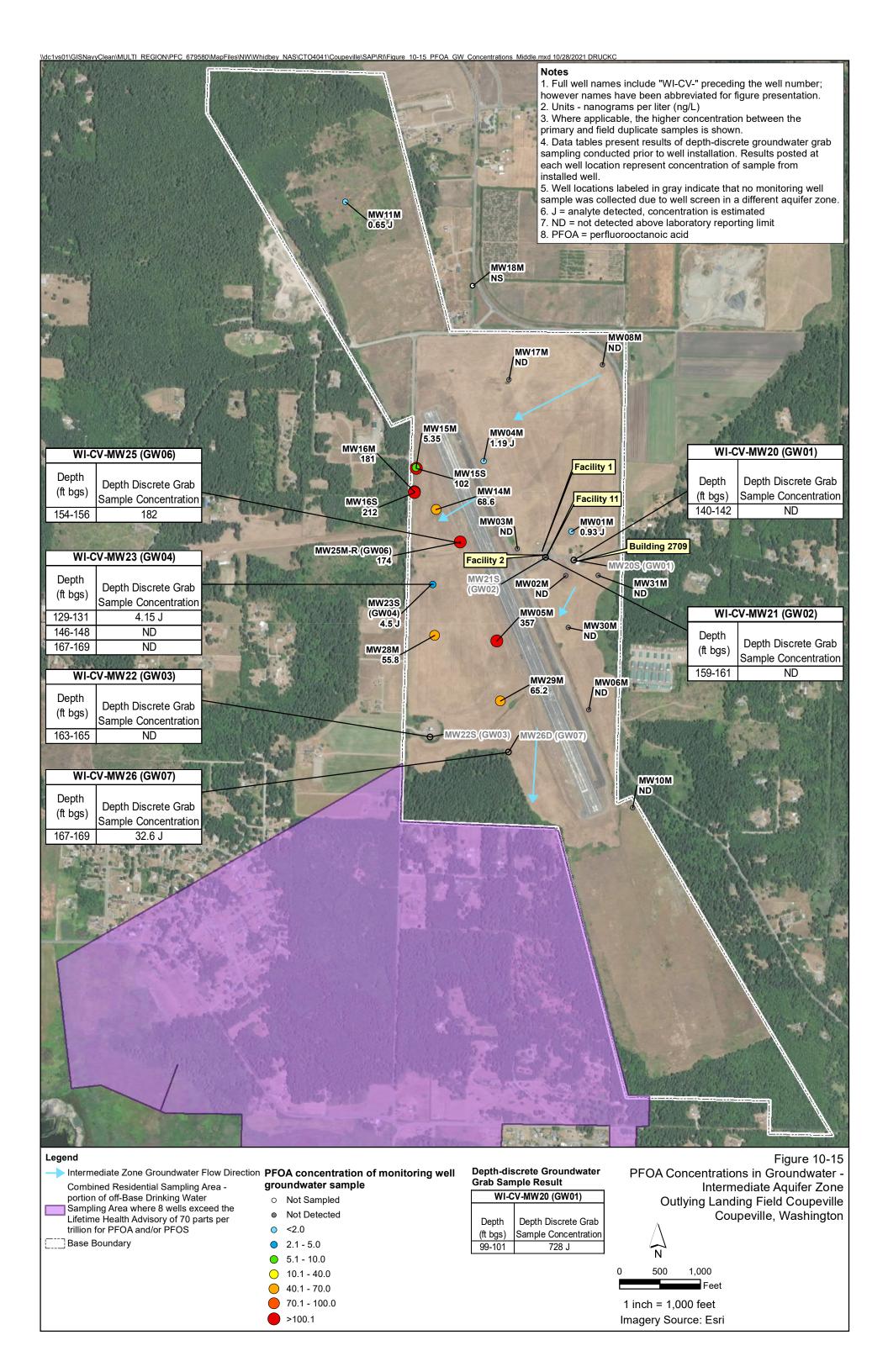
SO02	8/15/19		8/16/19		
Depth (ft bgs)	0.5-1.0	24-25	48-49	73-75	92-93
PFBS	1.09 U	1.07 U	1.06 U	1.12 U	1.18 U
PFOS	51.3	1.07 U	44.8	20.1	5.37 J
PFOA	1.09 U	1.07 U	3.85 J	1.62 U	1.18 U

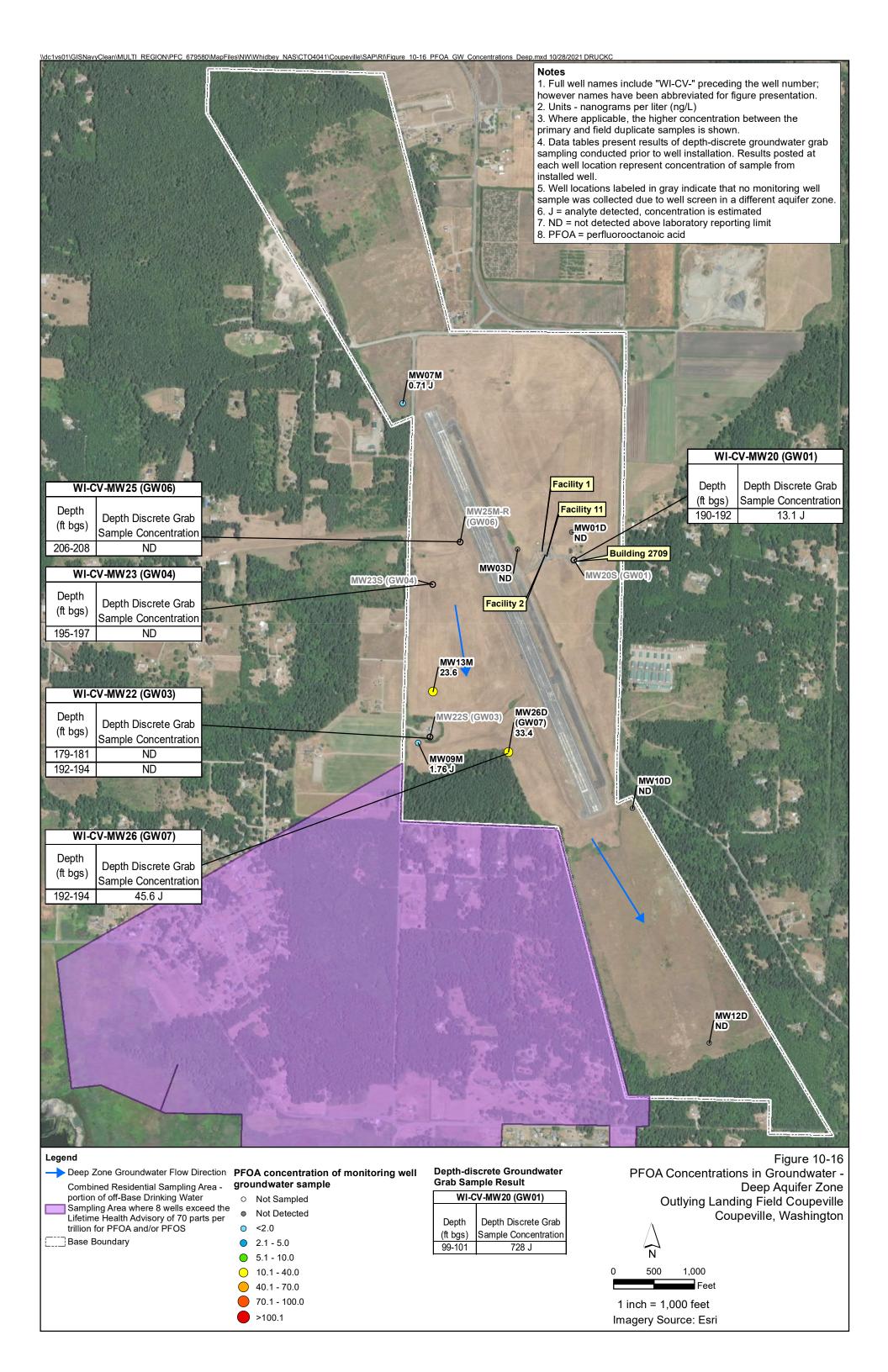
Figure 10-13

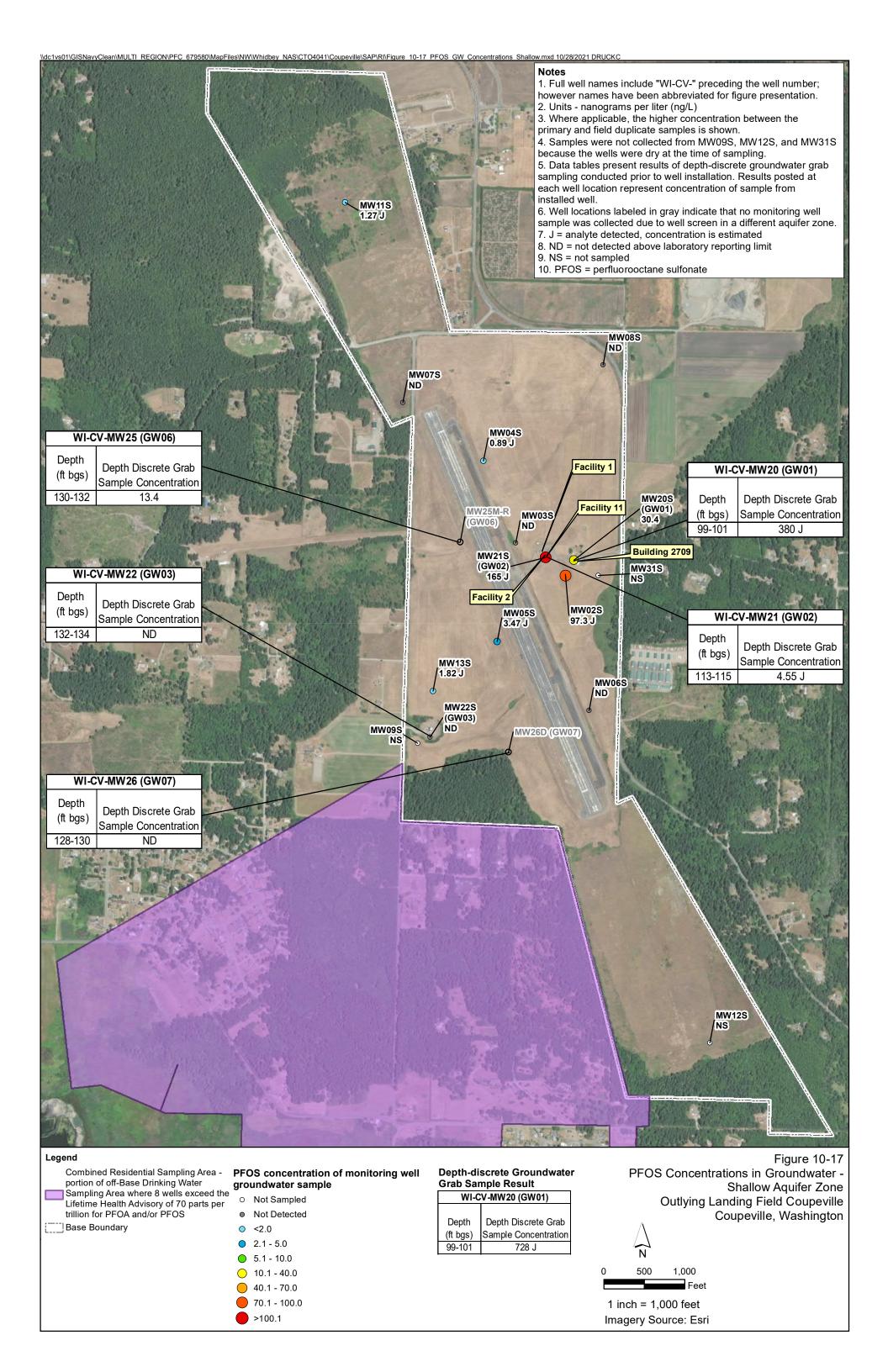
Summary of Soil PFAS Concentrations
Outlying Landing Field Coupeville
Coupeville, Washington

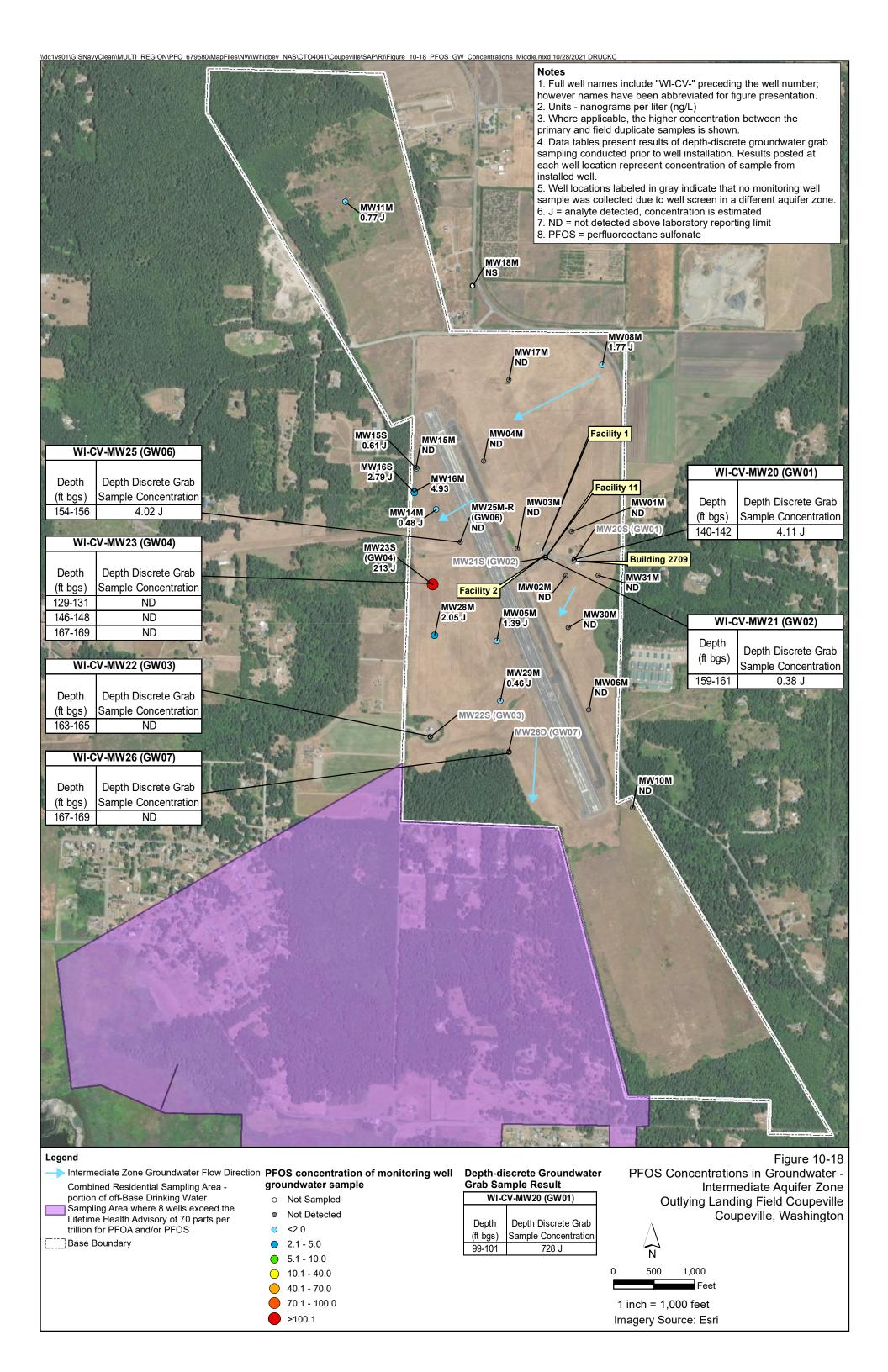
1 inch = 150 feet Imagery Source: Esri

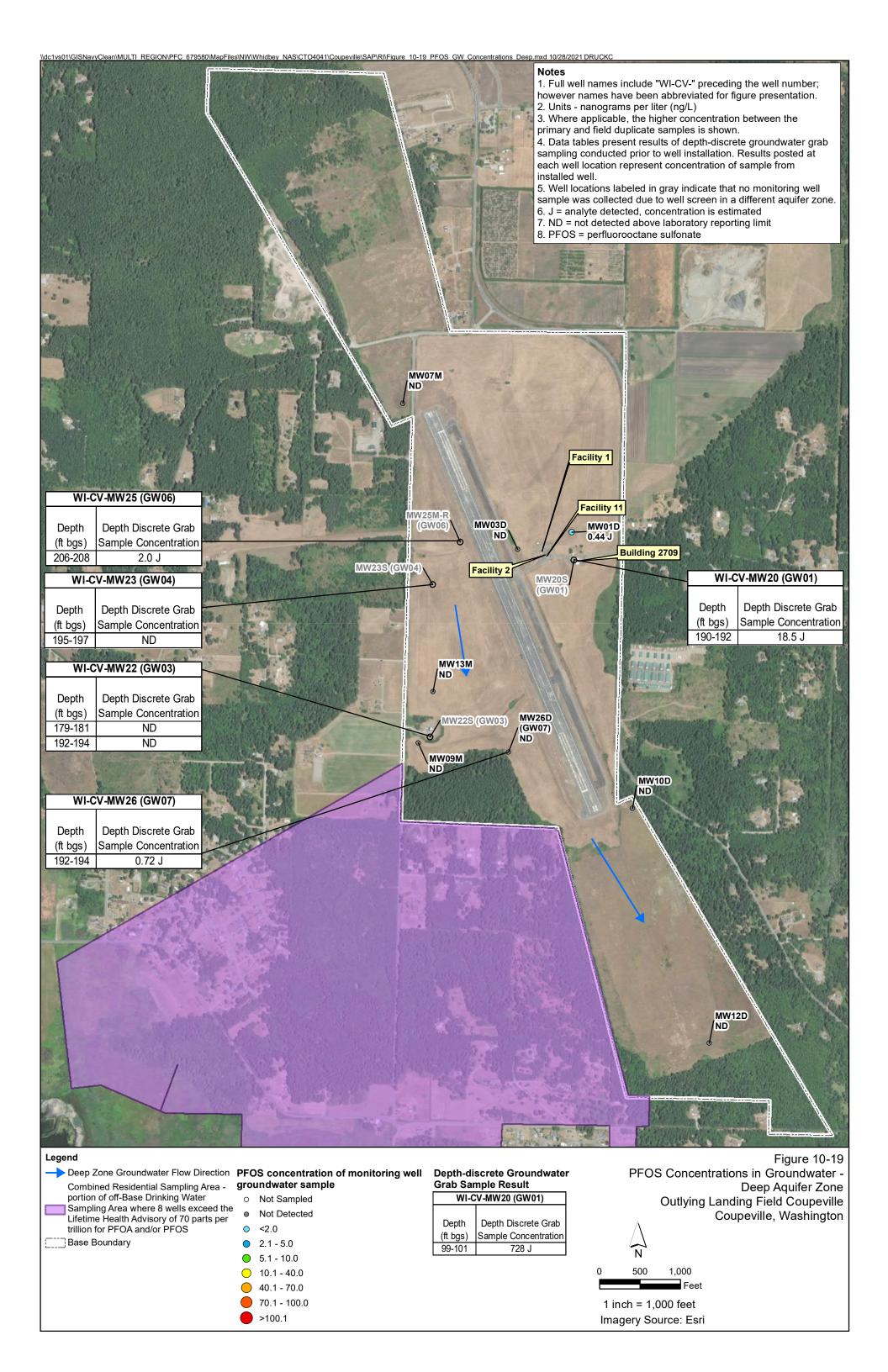


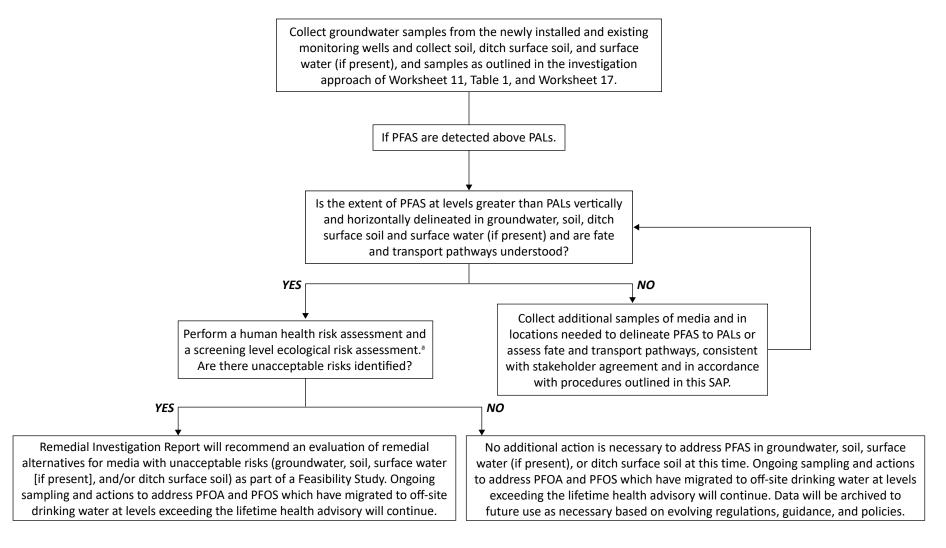












a PFAS will be evaluated within the applicable DoD, Navy, and/or EPA policy, guidance, or directives using the state-of-the-science toxicological information available and current at the time the RI is prepared for both the Human Health and Ecological Risk Assessments. Prior to conducting an ecological risk screening, the ecological screening values that will be used for comparison to site media will be reviewed based on current science and relevancy to site-specific receptors (e.g., relevant ecological receptors present). Ecological screening values selected for the screening will be presented in a technical memo or other documented form for regulatory review and acceptance.

Note:

Ongoing sampling and other activities being done to address off-base drinking water that exceeds the USEPA lifetime health advisory will not be conducted as a part of RI activities at OLF Coupeville.

Figure 11-1.

Decision Logic for Project Quality Objectives

Naval Air Station Whidbey Island – Outlying Landing Field Coupeville Coupeville, Washington

Appendix A RI Scoping Session Presentation

OLF Coupeville PFAS Remedial Investigation – Stakeholder Scoping Meeting Part 1 & 2

NAS Whidbey Island

December 10, 2020





Safety Minute – Holiday Light Safety



- 1. Never leave lights unattended (use timers, remotes, apps)
- 2. LED instead of incandescent more efficient, safer
- 3. Candle placement (or don't use!)
- 4. Use the right type of lights indoor and outdoor lights are not interchangeable
- 5. Only three light strands per outlet at most
- 6. Get rid of old lights
- 7. Securely fasten outdoor light strings, keep indoor lights out or reach from pets/ small children
- 8. Avoid electric lights on metallic trees
- 9. Be smart with extension cords keep cords dry/ above ground, use surge protectors
- 10. Keep Christmas tree from becoming fire hazard non-flammable decorations, keep watered

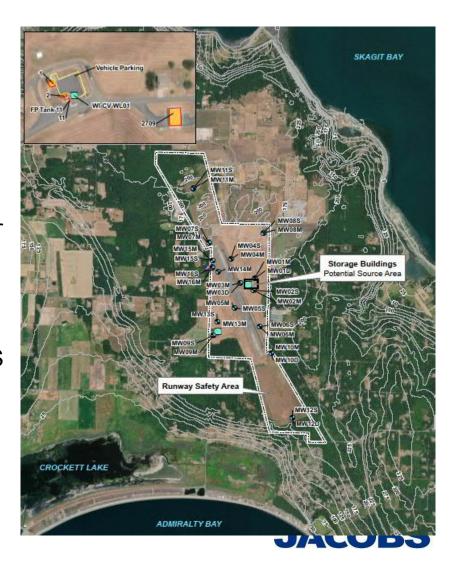


PART 1 – BACKGROUND AND SUPPLEMENTAL SITE INSPECTION RESULTS

JACOBS

Site Background

- Located on broad plateau, approx. 195 ft elevation
- Runway bordered by grass, runway safety area
- Admiralty Bay to south, Skagit Bay to NE
- Town of Coupeville community drinking water well (Keystone Well) just west of OLF; also obtain water from Ft. Casey wellfield (0.4 mi. SW of OLF)
- Other private community or individual drinking water wells near OLF
- No evidence of AFFF use at OLF Coupeville; PFAS detected in 2016 in on-Base sample, below EPA's lifetime health advisory
- GW/ DW investigations conducted following PFAS detection on-Base (detailed in next slide)

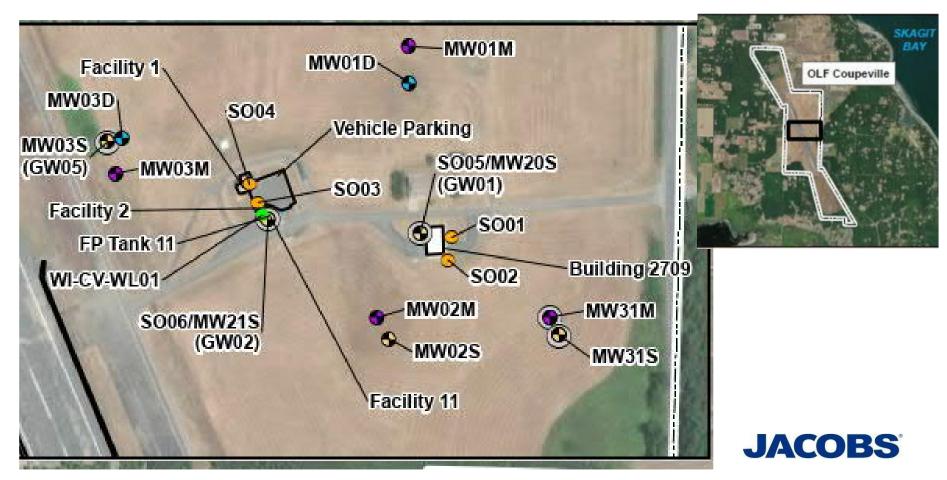


Investigation History

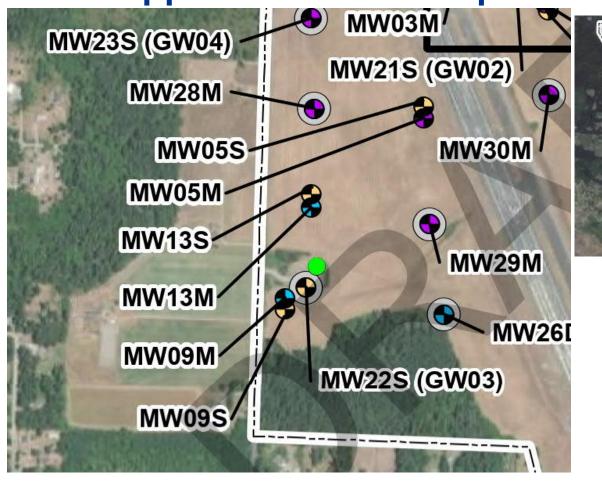
- Drinking Water Investigation (2016-2017):
 - PFOA detected above lifetime health advisory in 8 residential wells south of OLF
- Expedited Site Inspection (SI) (2016-2017):
 - 27 MWs installed, across 3 elevation intervals
 - Groundwater mound identified in intermediate aquifer interval
 - PFOA detected in 5 samples (3 above lifetime health advisory), PFOS in 5 samples
- Preliminary Assessment (2017-2018):
 - Evaluation of potential source areas, migration pathways, potential receptors
 - Identified three on-Base potential source areas:
 - Building 2709 (Crash Truck Shelter)
 - Facilities 1, 2, and 11 (Control Tower, Airfield Ops Building, and Potable Water Well Pump House)
 - Building 2807 (OLF Electronic Warfare Signal Emitter Building) not recommended for additional site Inspection



PFAS Source Areas Identified in Preliminary Assessment



Potential 'New' PFAS Source Area Identified in Supplemental Site Inspection





OLF Coupeville

Investigation History

Focused Site Inspection (2017-2018)

- 2 new well pairs (MW15M/S and MW16M/S), aquifer testing, develop GW Model
- Sampled select on-Base wells and Keystone Well; 5 exceed LHA for PFOA
- Confirmed presence of GW mound north of runway, with radial flow outward
- Keystone Well capture zone includes PFAS exceedances in intermediate interval

Supplemental SI (2019-present):

- Phase 1 on-Base soil borings (6); soil, GW, geotechnical sampling, depth-discrete
 GW sampling; MW installs (8)
- Phase 2 5 additional wells, aquifer testing (May 2020)
- Phase 3 1 off-Base well NE of OLF (for hydraulic data) Oct. 2020
- Update to GW model and conduct solute transport modeling



Supplemental SI Investigation Results

Soil data

- Overall, heterogeneous depth distribution
- · PFOS more frequent, at relatively higher concentrations
- Detections above PALs indicates likely release, confirms source areas (Bldg. 2709 and Facilities 1, 2, and 11)
- PFOA detected in 18 samples, concentrations ranged from 0.87 74.7 μg/kg
- PFOS detected in 22 samples, concentrations ranged from 4.92 936 μg/kg

Groundwater data

- Depth-discrete samples used to determine well screen placement
- PFBS detected in 26 MWs; none above PAL
- PFOA detected in 23 MWs; 11 exceeded PAL (40 ng/L) most frequently detected
 - Shallow zone: highest near Bldg. 2709, near Keystone Well, and west of runway
 - Intermediate zone: highest near Keystone Well and west of runway
 - Deep zone: only exceeded PAL at MW26D, south of runway
- PFOS detected in 16 MWs; 3 exceeded PAL (40 ng/L)
 - Shallow zone: highest near Bldg. 2709, Facilities 1, 2, 11, and due west (MW23S)
 - No PAL exceedances intermediate and deep wells



Supplemental SI Investigation Results

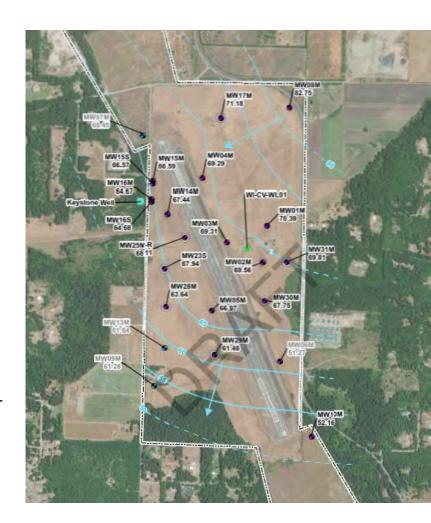
Conceptual Site Model (CSM) Update

– Groundwater flow:

- Discontinuous perched shallow GW 90-130 ft bgs
- Discontinuous clay/silt layer below, pinches out in south
- Semiconfined intermediate aguifer unit, 124 to 145 ft bgs
- Intermediate aquifer underlain by confining layer
- Flow in intermediate zone to S-SW, Deep GW flow to S-SE
- GW mound further to NE than previously thought
- Heads near west Base boundary are influenced by Keystone Well

PFAS Pathways

- PFOA most prominent; above PALs in shallow, intermediate aquifer zones downgradient of source areas
- Likely downward migration along flow direction
- High PFOA west of runway suggest secondary source
- Elevated PFOS confined to shallow zone near source areas





Supplemental SI Investigation Results – Groundwater Model

GW flow model

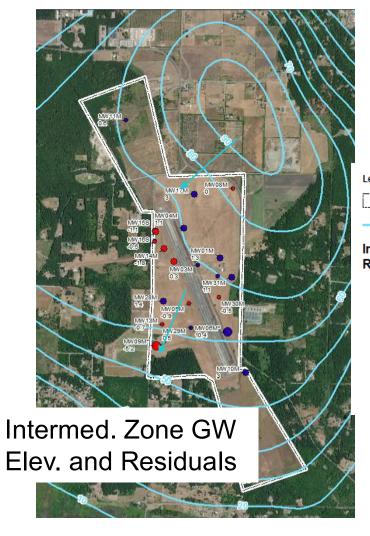
- Recalibration / update to 2018 3-D GW flow model using Supplemental SI data
- Retains most design and construction elements of 2018 model
- Increased number of model layers (from 5 to 7 layers)
- Updated subsurface hydraulic parameters, boundary conditions, evapotranspiration and recharge rates
- Evaluation of pre-Keystone Well flow field

Solute transport modeling for PFOA and PFOS

- Simulates loading of PFOA and PFOS to groundwater from vadose zone soils at on-Base source areas
- Selection of boundary conditions, subsurface hydraulic parameters, and solute transport parameters
- Solute transport parameters for PFOA, PFOS based on literature review
- Current PFOA and PFOS soil concentrations with depth from Supplemental SI soil sampling data
- Vadose zone model output is simulated PFOA, PFOS porewater concentrations reaching water table over time (initial conditions through 100 years of transport)



Groundwater Flow Modeling Results



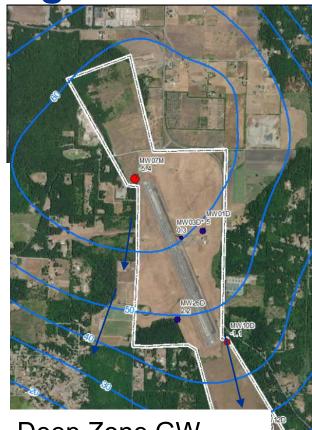
Legend

OLF Boundary

Internediate Zone Groundwater Elevation Contour

Intermediate Zone Residuals

- -10 -5
- -5 -1
- -1 0
- 0 1
- 1-5 5-10.5
- 5 10.



Deep Zone GW Elev. and Residuals





Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

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Solute Transport Modeling - Conclusions

- Simulated PFOA plume migration is consistent with current CSM and observations of elevated PFAS in drinking water wells south of OLF
- Simulated future migration of PFOS is dependent on assumed solute transport parameters
- Continued mass flux from vadose zone PSAs has a strong influence on plume migration distance and persistence
- High degree of uncertainty in the scientific understanding of PFAS solute transport parameters



Data Gaps and Remaining Uncertainties

- Source Areas
 - On-Base near Bldg. 2709, Facility 1, 2, 11 additional delineation
 - Lag in loading: PFOS vs. PFOA
 - Need for additional soil borings, delineation and vadose zone work
 - Current PFAS plume extents inconsistent with low soil concentrations at depth
- Potential New Source Area
 - On- and off-Base near Keystone Well
- On-Base data gaps at south end of Base
- Connectivity between on-Base sources / PAL exceedances and off-Base exceedances south of Base
- Groundwater flow and quality off-Base
 - Uncertainty to east and west of OLF



PART 2 - REMEDIAL INVESTIGATION APPROACH



Objectives of OLF Coupeville RI

- Install, sample, and survey up to 15 new groundwater wells on-Base to refine the vertical and horizontal extent of PFOS, PFOA, and PFBS and confirming source areas;
- Collect soil data to refine source area information;
- Install, sample, and survey up to 5 new groundwater wells off-Base to refine the vertical and horizontal extent of PFOS, PFOA, and PFBS off-Base, originating from OLF Coupeville;
- Sample all previously installed groundwater monitoring wells on-Base;
- Conduct aquifer testing after all new wells installed, as appropriate, to refine CSM;
- Update fate and transport groundwater model with new and existing data;
- Collect and analyze up to 10 sediment and surface water samples, as necessary, to support the human health risk assessments;
- Conduct human health risk assessments for PFAS, as appropriate;
- Determine if Feasibility Study is necessary or if more data needed to refine RI.



RI Approach

- Work planning and field work in two phases:
 - RI SAP (on-Base)
 - RI Field Work (on-Base)
 - RI SAP Addendum (off-Base)
 - Real Estate documents for off-Base access
 - RI Field Work (off-Base)
 - RI Report (on- and off-Base)



Data Gaps and Remaining Uncertainties

- Source Areas
 - On-Base near Bldg. 2709, Facility 1, 2, 11 additional delineation
 - Lag in loading: PFOS vs. PFOA
 - Need for additional soil borings, delineation and vadose zone work
 - Current PFAS plume extents inconsistent with low soil concentrations at depth
- Potential New Source Area
 - On- and off-Base near Keystone Well
- On-Base data gaps at south end of Base
- Connectivity between on-Base sources / PAL exceedances and off-Base exceedances south of Base
- Groundwater flow and quality off-Base
 - Uncertainty to east and west of OLF

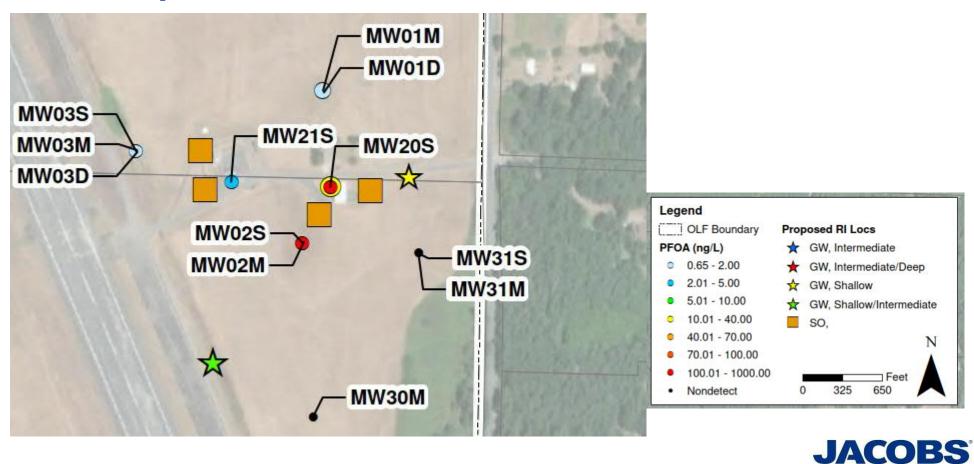


Proposed RI Locations – on-Base Source Areas and Data Gaps

Item/ Task	Location	Rationale	Comments
Monitoring Well Pair	Downgradient of MW02S, between MW02S, MW05 wells	Data gap between source areas at 02S, 20S, 21S and runway/ downgradient locations (MW05 PFOA exceedances)	Paired shallow, intermediate wells GW vertical profile sampling on intermediate location
Monitoring Well (single)	East of MW20S	Bounding to shallow PFOA/ PFOS exceedances at east Base boundary	Targeting shallow aquifer only No GW vertical profiling
Soil Boring	East of MW20S	Provide greater delineation	Assumes soil boring to water table, collection of 4-5 soil samples at each location
Soil Boring	Southwest of MW20S	of source areas near Building 2709 and Facilities	
Soil Boring	Northwest of MW21S	1, 2, and 11	
Soil Boring	Southwest of MW21S		



Proposed RI Locations – on-Base Source Areas and Data Gaps





Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

NOTIFICATION: CONTAINS SENSITIVE BUT UNCLASSIFIED INFORMATION WHICH IS PROTECTED BY THE FREEDOM OF INFORMATION ACT

FOIA Exemption 6 (5 USC 552(b)(6))
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Proposed RI Locations – Sediment/ Surface Water

(on-Base)

Approximate sediment/ surface water sample location



JACOBS



Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

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Schedule (approximate)

- Generate RI SAP (on-Base) now to May 2021
- RI SAP Stakeholder Review May/June 2021
- Field work (on-Base) Fall 2021 (Sept Nov)
- Generate RI SAP Addendum (off-Base) Jan Apr 2021
- RI SAP Addendum Stakeholder Review May 2021
- Field work (off-Base) Fall 2021 (Oct Nov)
- RI Report Dec 2021 June 2022
- RI Report Stakeholder Review June/July 2022



Thank you for your time



Appendix B Field Standard Operating Procedures

Logging of Soil Borings

I. Purpose and Scope

This SOP provides guidance to obtain accurate and consistent descriptions of soil characteristics during soil-sampling operations. The characterization is based on visual examination and manual tests, not on laboratory determinations.

II. Equipment and Materials

- Indelible pens
- Tape measure or ruler
- Field logbook
- Spatula
- HCL, 10 percent solution
- Squirt bottle with water
- Rock- or soil-color chart (e.g., Munsell)
- Grain-size chart
- Hand lens
- Unified Soil Classification System (USCS) index charts and tables to help with soil classification (attached)

III. Procedures and Guidelines

This section covers several aspects of soil characterization: instructions for completing the soil boring log form (attached), field classification of soil, and standard penetration test procedures.

A. Instructions for Completing Soil Boring Logs

Soil boring logs will be completed in the field log books or on separate soil boring log sheets. Information collected will be consistent with that required for ASTM D1586, a standard soil boring log form (attached), or an equivalent form that supplies the same information.

The information collected in the field to perform the soil characterization is described below.

Field personnel should review completed logs for accuracy, clarity, and thoroughness of detail. Samples also should be checked to see that information is correctly recorded on both sample jar labels and on the log sheets.

B. Heading Information

Boring/Well Number. Enter the boring/well number. A numbering system should be chosen that does not conflict with information recorded for previous exploratory work done at the site. Number the sheets consecutively for each boring.

Location. If station, coordinates, mileposts, or similar project layout information is available, indicate the position of the boring to that system using modifiers such as "approximate" or "estimated" as appropriate.

Elevation. Elevation will be determined at the conclusion of field activities through a survey.

Drilling Contractor. Enter the name of the drilling company and the city and state where the company is based.

Drilling Method and Equipment. Identify the bit size and type, drilling fluid (if used), and method of drilling (e.g., rotary, hollow-stem auger, sonic). Information on the drilling equipment (e.g., CME 55, Mobile B61) also is noted.

Water Level and Date. Enter the depth below ground surface to the apparent water level in the borehole. The information should be recorded as a comment. If free water is not encountered during drilling or cannot be detected because of the drilling method, this information should be noted. Record date and time of day (for tides, river stage) of each water level measurement.

Date of Start and Finish. Enter the dates the boring was begun and completed. Time of day should be added if several borings are performed on the same day.

Logger. Enter the first and last name.

C. Technical Data

Depth Below Surface. Use a depth scale that is appropriate for the sample spacing and for the complexity of subsurface conditions.

Sample Interval. Note the depth at the top and bottom of the sample interval.

Sample Type and Number. Enter the sample type and number. SS-1 = split spoon, first sample. Number samples consecutively regardless of type. Enter a sample number even if no material was recovered in the sampler.

Sample Recovery. Enter the length to the nearest 0.1-foot of soil sample recovered from the sampler. Often, there will be some wash or caved material above the sample; do not include the wash material in the measurement. Record soil recovery in feet.

Standard Penetration Test Results. In this column, enter the number of blows required for each 6 inches of sampler penetration and the "N" value, which is the sum of the blows in the middle two 6-inch penetration intervals. A typical standard penetration test involving successive blow counts of 2, 3, 4, and 5 is recorded as 2-3-4-5 and (7). The standard penetration test is terminated if the sampler encounters

refusal. Refusal is a penetration of less than 6 inches with a blow count of 50. A partial penetration of 50 blows for 4 inches is recorded as 50/4 inches. Penetration by the weight of the slide hammer only is recorded as "WOH."

Samples should be collected using a 140-pound hammer and 2-inch diameter split spoons. Samples may be collected using direct push sampling equipment. However, blow counts will not be available. A pocket penetrometer may be used instead to determine relative soil consistency of fine grained materials (silts and clays).

Sample also may be collected using a 300-pound hammer or 3-inch-diameter split-spoon samples at the site. However, use of either of these sample collection devices invalidates standard penetration test results and should be noted in the comments section of the log. The 300-pound hammer should only be used for collection of 3-inch-diameter split-spoon samples. Blow counts should be recorded for collection of samples using either a 3-inch split-spoon, or a 300-pound hammer. An "N" value need not be calculated.

Soil Description. The soil classification should follow the format described in the "Field Classification of Soil" subsection below.

Comments. Include all pertinent observations (changes in drilling fluid color, rod drops, drilling chatter, rod bounce as in driving on a cobble, damaged Shelby tubes, and equipment malfunctions). In addition, note if casing was used, the sizes and depths installed, and if drilling fluid was added or changed. You should instruct the driller to alert you to any significant changes in drilling (changes in material, occurrence of boulders, and loss of drilling fluid). Such information should be attributed to the driller and recorded in this column.

Specific information might include the following:

- The date and the time drilling began and ended each day
- The depth and size of casing and the method of installation
- The date, time, and depth of water level measurements
- Depth of rod chatter
- Depth and percentage of drilling fluid loss
- Depth of hole caving or heaving
- Depth of change in material
- Health and safety monitoring data
- Drilling interval through a boulder

D. Field Classification of Soil

This section presents the format for the field classification of soil. In general, the approach and format for classifying soils should conform to ASTM D 2488, Visual-Manual Procedure for Description and Identification of Soils.

The Unified Soil Classification System is based on numerical values of certain soil properties that are measured by laboratory tests. It is possible, however, to estimate these values in the field with reasonable accuracy using visual-manual procedures (ASTM D 2488). In addition, some elements of a complete soil description, such as the presence of cobbles or boulders, changes in strata, and the relative proportions of soil types in a bedded deposit, can be obtained only in the field.

Soil descriptions should be precise and comprehensive without being verbose. The correct overall impression of the soil should not be distorted by excessive emphasis on insignificant details. In general, similarities rather than differences between consecutive samples should be stressed.

Soil descriptions must be recorded for every soil sample collected. The format and order for soil descriptions should be as follows:

- 1. Soil name (synonymous with ASTM D 2488 Group Name) with appropriate modifiers. Soil name should be in all capitals in the log, for example "POORLY-GRADED SAND."
- 2. Group symbol, in parentheses, for example, "(SP)."
- 3. Color, using Munsell color designation
- 4. Moisture content
- 5. Relative density (coarse grained) or consistency (fine grained)
- 6. Soil structure, mineralogy, or other descriptors

This order follows, in general, the format described in ASTM D 2488.

E. Soil Name

The basic name of a soil should be the ASTM D 2488 Group Name on the basis of visual estimates of gradation and plasticity. The soil name should be capitalized.

Examples of acceptable soil names are illustrated by the following descriptions:

- A soil sample is visually estimated to contain 15 percent gravel, 55 percent sand, and 30 percent fines (passing No. 200 sieve). The fines are estimated as either low or highly plastic silt. This visual classification is SILTY SAND WITH GRAVEL, with a Group Symbol of (SM).
- Another soil sample has the following visual estimate: 10 percent gravel, 30 percent sand, and 60 percent fines (passing the No. 200 sieve). The fines are estimated as low plastic silt. This visual classification is SANDY SILT. The gravel portion is not included in the soil name because the gravel portion was estimated as less than 15 percent. The Group Symbol is (ML).

The gradation of coarse-grained soil (more than 50 percent retained on No. 200 sieve) is included in the specific soil name in accordance with ASTM D 2488. There is no need

to further document the gradation. However, the maximum size and angularity or roundness of gravel and sand-sized particles should be recorded. For fine-grained soil (50 percent or more passing the No. 200 sieve), the name is modified by the appropriate plasticity/elasticity term in accordance with ASTM D 2488.

Interlayered soil should each be described starting with the predominant type. An introductory name, such as "Interlayered Sand and Silt," should be used. In addition, the relative proportion of each soil type should be indicated (see Table 1 for example).

Where helpful, the evaluation of plasticity/elasticity can be justified by describing results from any of the visual-manual procedures for identifying fine-grained soils, such as reaction to shaking, toughness of a soil thread, or dry strength as described in ASTM D 2488.

F. Group Symbol

The appropriate group symbol from ASTM D 2488 must be given after each soil name. The group symbol should be placed in parentheses to indicate that the classification has been estimated.

In accordance with ASTM D 2488, dual symbols (e.g., GP-GM or SW-SC) can be used to indicate that a soil is estimated to have about 10 percent fines. Borderline symbols (e.g., GM/SM or SW/SP) can be used to indicate that a soil sample has been identified as having properties that do not distinctly place the soil into a specific group. Generally, the group name assigned to a soil with a borderline symbol should be the group name for the first symbol. The use of a borderline symbol should not be used indiscriminately. Every effort should be made to first place the soil into a single group.

G. Color

The color of a soil must be given. The color description should be based on the Munsell system. The color name and the hue, value, and chroma should be given.

H. Moisture Content

The degree of moisture present in a soil sample should be defined as dry, moist, or wet. Moisture content can be estimated from the criteria listed on Table 2.

I. Relative Density or Consistency

Relative density of a coarse-grained (cohesionless) soil is based on N-values (ASTM D 1586 [attached]). If the presence of large gravel, disturbance of the sample, or non-standard sample collection makes determination of the in situ relative density or consistency difficult, then this item should be left out of the description and explained in the Comments column of the soil boring log.

Consistency of fine-grained (cohesive) soil is properly based on results of pocket penetrometer or torvane results. In the absence of this information, consistency can be estimated from N-values. Relationships for determining relative density or consistency of soil samples are given in Tables 3 and 4.

J. Soil Structure, Mineralogy, and Other Descriptors

Discontinuities and inclusions are important and should be described. Such features include joints or fissures, slickensides, bedding or laminations, veins, root holes, and wood debris.

Significant mineralogical information such as cementation, abundant mica, or unusual mineralogy should be described.

Other descriptors may include particle size range or percentages, particle angularity or shape, maximum particle size, hardness of large particles, plasticity of fines, dry strength, dilatancy, toughness, reaction to HCl, and staining, as well as other information such as organic debris, odor, or presence of free product.

K. Equipment and Calibration

Before starting the testing, the equipment should be inspected for compliance with the requirements of ASTM D 1586. The split-barrel sampler should measure 2-inch or 3-inch OD, and should have a split tube at least 18 inches long. The minimum size sampler rod allowed is "A" rod (1-5/8-inch OD). A stiffer rod, such as an "N" rod (2-5/8-inch OD), is required for depths greater than 50 feet. The drive weight assembly should consist of a 140-pound or 300-pound hammer weight, a drive head, and a hammer guide that permits a free fall of 30 inches.

IV. Attachments

Soil Boring Log (Sample Soil Boring Log.xls)

Soil Boring Log Form with a completed example (Soil_Log_Examp.pdf)

Tables 1 through 4 (Tables 1-4.pdf)

V. Key Checks and Preventive Maintenance

- Check entries to the soil-boring log and field logbook in the field; because the samples will be disposed of at the end of fieldwork, confirmation and corrections cannot be made later.
- Check that sample numbers and intervals are properly specified.
- Check that drilling and sampling equipment is decontaminated using the procedures defined in SOP Decontamination of Drilling Rigs and Equipment.

VI. References

ASTM D2488-17e1, Standard Practice for Description and Identification of Soils (Visual-Manual Procedures), ASTM International, West Conshohocken, PA, 2017, www.astm.org

ASTM D1586 / D1586M-18, Standard Test Method for Standard Penetration Test (SPT) and Split-Barrel Sampling of Soils, ASTM International, West Conshohocken, PA, 2018, www.astm.org



ROJECT NUMBER	BORING NUMBER

					SOIL BORING LOG					
ROJEC	T :			<u> </u>	LOCATION :					
LEVAT					DRILLIN	NG CONTRAC				
			EQUIPME	NT USED :						
TD WA	TER LEV	EL:			START:		END:			LOGGER:
PTH BE	LOW SURFA	ACE (FT)		STANDARD		SOIL DESCRI	PTION		USCS	COMMENTS
	INTERVAL ((FT) RECOVEF	RY (FT) SAMPLE #/TYPE	PENETRATION TEST RESULTS 6"-6"-6"-6" (N')	MOISTUR	E, USCS GROUI RE CONTENT, RI NSISTENCY, SC MINERALC	ELATIVE DENS DIL STRUCTUR	SITY,		DEPTH OF CASING, DRILLING RATE, DRILLING FLUID LOSS, TESTS, AND INSTRUMENTATION. DRILLING ACTIONS DRILLER COMMENTS PID Readings: Breathing Zone: Above Hole:
				,						_
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BORING NUMBER

SHEET

OF

SOIL BORING LOG

OJECT						LOCATION	
EVATIO	N				_ DRILLING CONTRACTOR	R	
TERL	METH	IOD AND	D EQUIP	PMENT	START	FINISH	LOGGER
				STANDARD	SOIL DESCRI		COMMENTS
SURFACE (FT)	BER TYPE DVERY		PENETRATION - TEST RESULTS 6"-6"-6" (N)	PENETRATION TEST RESULTS SOIL NAME, USCS GROUP SYMBOL, COL MOISTURE CONTENT, RELATIVE DENSI OR CONSISTENCY, SOIL STRUCTURE, MINERALOGY		OR, DEPTH OF CASING, DRILLING RATE,	
							Figure 1 SOIL BORING LOG, FORM D1586



PROJECT NUMBER

DEN 22371. G5

BORING NUMBER

SHEET

OF 3

SOIL BORING LOG

PROJECT Howard Ave (and slide Location Howard 1: 24th Ave, Centennial, CO.

ELEVATION 5/36 Feet DRILLING CONTRACTOR Kendall Explorations, Ashcan, Colorado

DRILLING METHOD AND EQUIPMENT 4"-Inch H.S. Aligers, Mobil B-61 rotary drill rig.

WATER LEVELS 3.2 Feet 8/5/89 STARTAIN BE A 1989 FINISH ALIGN BEB-100 GGER J.A. Michner

	7	SAMPLE		STANDARD PENETRATION	SOIL DESCRIPTION	COMMENTS
SURFACE (FT)	INTERVAL	NUMBER AND TYPE	RECOVERY (FT)	TEST RESULTS 6°-6°-6° (N)	SOIL NAME, USCS GROUP SYMBOL, COLOR, MOISTURE CONTENT, RELATIVE DENSITY OR CONSISTENCY, SOIL STRUCTURE, MINERALOGY	DEPTH OF CASING, DRILLING RATE, DRILLING FLUID LOSS, TESTS AND INSTRUMENTATION
7	2.5				Surface material consist of 4 inches AC underlain by Gindres of 34 inch - minus base rock -	Start Drilling @ 3:00
	4.0	1-5	1:5	2-3-4 (7)	POORLY-GRADED SANDWITH - SILT, (SP-5M), Ane, light brown, wet, loose	Driller notes water at 4 feet
; -	5.0	2-5	0.9	WOH/12"-1	ORGANIC SILT, (OL), very dark, gray to black, wet, very soft; strong H25 odor, many fine roots up to about 1/4 inch	Driller notes very soft drilling 412. dark grey, wet sitty cuttings.
	8.0	3-ST	1.3		ORGANIC SILT, Similar to 2-8, except includes fewer roots - (by volume)	
D -	10.0	10	1.3	2-2-2	SILT, (ML), very dark gray to black, wet, soft	water level @ 3.2 feet on 8/5/89 @ 0730
						Driller notes roughdrilling action and chatter @ 13ft
5-	15.0 15.5	5-5	0.5	60/6"	SIJY GRAVEL, (GM), rounded gravel up to about finch maximum observed Size, wet, very dense	
) -	20.0				I FANCIAY WITH SAND. (CL).	Driller notes smoother, firm drilling @ 19 ft some angular rockchips@ bo t in of its 5 ross balders or n
	23.0		1.0		wery stiff	Driller notes very hard, sto grinding, smooth drilling action from 21 to 23 ft, possibly bedrock
-	23.1	7-5	0	50/1"	NO RECOVERY END SOIL BORING © 23.1 FEET SEE ROCK CORE LOG FOR	
				· · · · · · · · · · · · · · · · · · ·	CONTINUATION OF BL-3	Figure 2 EXAMPLE OF COMPLETE LOG FORM

Table 1 EXAMPLE SOIL DESCRIPTIONS

POORLY GRADED SAND (SP), light brown, moist, loose, fine sand size

FAT CLAY (CH), dark gray, moist, stiff

SILT (ML), light greenish gray, wet, very loose, some mica, lacustrine

WELL-GRADED SAND WITH GRAVEL (SM), reddish brown, moist, dense, subangular gravel to 0.6 inches max

POORLY GRADED SAND WITH SILT (SP-SM), white, wet, medium dense

ORGANIC SOIL WITH SAND (OH), dark brown to black, wet, firm to stiff but spongy undisturbed, becomes soft and sticky when remolded, many fine roots, trace of mica

SILTY GRAVEL WITH SAND (GM), brownish red, moist, very dense, subrounded gravel to 1.2 inches max

INTERLAYERED SILT (60 percent) AND CLAY (40 percent): SILT WITH SAND (ML), medium greenish gray, nonplastic, sudden reaction to shaking, layers mostly 1.5 to 8.3 inches thick; LEAN CLAY (CL), dark gray, firm and brittle undisturbed, becomes very soft and sticky when remolded, layers 0.2 to 1.2 inches thick

SILTY SAND WITH GRAVEL (SM), light yellowish brown, moist, medium dense, weak gravel to 1.0 inches max, very few small particles of coal, fill

SANDY ELASTIC SILT (MH), very light gray to white, wet, stiff, weak calcareous cementation

LEAN CLAY WITH SAND (CL/MH), dark brownish gray, moist, stiff

WELL-GRADED GRAVEL WITH SILT (GW-GM), brown, moist, very dense, rounded gravel to 1.0 inches max

SF032/010.50

Table 2
CRITERIA FOR DESCRIBING MOISTURE CONDITION

Description	Criteria		
Dry	Absence of moisture, dusty, dry to the touch		
Moist	Damp, but no visible water		
Wet	Visible free water, usually soil is below water table		

Table 3
RELATIVE DENSITY OF COARSE-GRAINED SOIL
(Developed from Sowers, 1979)

SILT OUL

LEAN CL

Blows/Ft	Relative Density	Field Test
0-4	Very loose	Easily penetrated with ½-in. steel rod pushed by hand
5-10	Loose	Easily penetrated with ½-in. steel rod pushed by hand
11-30	Medium	Easily penetrated with ½-in. steel rod driven with 5-lb hammer
31-50	Dense	Penetrated a foot with ½-in. steel rod driven with 5-lb hammer
>50	Very dense	Penetrated only a few inches with ½-in. steel rod driven with 5-lb hammer

Table 4
CONSISTENCY OF FINE-GRAINED SOIL
(Developed from Sowers, 1979)

Blows/Ft	Consistency	Pocket Penetrometer (TSF)	Torvane (TSF)	Field Test
<2	Very soft	<0.25	<0.12	Easily penetrated several inches by fist
2-4	Soft	0.25-0.50	0.12-0.25	Easily penetrated several inches by thumb
5-8	Firm	0.50-1.0	0.25-0.5	Can be penetrated several inches by thumb with moderate effort
9-15	Stiff	1.0-2.0	0.5-1.0	Readily indented by thumb, but penetrated only with great effort
16-30	Very stiff	2.0-4.0	1.0-2.0	Readily indented by thumbnail
>30	Hard	>4.0	>2.0	Indented with difficulty by thumbnail

Water-Level Measurements

I. Purpose and Scope

The purpose of this procedure is to provide a guideline for the measurement of the depth to groundwater in piezometers and monitoring wells, even where a second phase of floating liquid (e.g., gasoline) is encountered, and on staff gauges in surfacewater bodies. This SOP includes guidelines for discrete measurements of static water levels and does not cover the use of continuously recording loggers (see SOP *Use of Data Loggers and Pressure Transducers*).

II. Equipment and Materials

- Electronic water-level meter (Solinst® or equivalent) with a minimum 100-foot tape; the tape should have graduations in increments of 0.01 feet or less
- Interface probe (Solinst® Model 122 Interface Meter or equivalent)

III. Procedures and Guidelines

Verify that the unit is turned on and functioning properly. Slowly lower the probe on its cable into the piezometer or well until the probe just contacts the water surface; the unit will respond with a solid tone or light signal. Note the depth from a reference point indicated on the piezometer or well riser. Typically, this is the top of the PVC casing. If no reference is clearly visible, measure the depth to water from the northern edge of the PVC casing. If access to the top of the PVC casing is difficult, sight across the top of the locking casing adjacent to the measuring point, recording the position of the cable when the probe is at the water surface.

Measure the distance from this point to the closest interval marker on the tape and record the water level reading in the logbook. Water levels will be measured to the nearest 0.01-foot. Also, when specified in the project plans, measure and record the depth of the piezometer or well. The depth of the piezometer or well may be measured using the water-level probe with the instrument turned off.

Free product light or dense nonaqueous phase liquid may be present in the piezometer or well. If the presence of free product is suspected, the thickness of the product should be determined using appropriate equipment (e.g., Solinst® Model 122 Interface Meter). The depth to water also is determined with this equipment and the water-level meter should not be used in the piezometer or well as long as product is present. Typically, a constant sound is emitted from the device when free product is encountered and an alternating on/off beep sound is emitted when water is encountered.

1

The apparent elevation of the water level in the well or piezometer is determined by measuring both the apparent depth to water and the thickness of free product. The corrected water-level elevation is calculated by the following equation:

 $WL_c = Wl_a + (Free-product thickness \times 0.80)$

Where WL_c = Corrected water-level elevation

Wl_a = Apparent water-level elevation

0.80 = Typical value for the density of petroleum hydrocarbon products.

If free product is detected on the surface of the water in the piezometer or well, the value of sampling should be reconsidered because of the potential for contaminating the sampling equipment.

Staff gauges may be installed in some surface-water bodies. These facilities typically are constructed by attaching a calibrated, marked staff gage to a wood or metal post, driving the post into the bottom of the surface-water body, and surveying the elevation of the top of the post to a resolution or 0.01-foot. The elevation of the water in the surface-water body then can be determined by reading off the distance the water level is from the top of the post. A shield or other protection may be needed to calm the fluctuations in water level if the gauge is installed at a location exposed to wind or wave.

IV. Attachments

None.

V. Key Checks

- Before each use, verify that the battery is charged by pressing the test button on the water-level meter.
- Verify that the unit is operating correctly by testing the probe in distilled or deionized water. Leave the unit turned off when not in use.

Multi RAE Photoionization Detector (PID)

I. Purpose

The purpose of this SOP is to provide general reference information for using the Multi RAE PID in the field. Calibration and operation, along with field maintenance, will be included in this SOP.

II. Scope

This procedure provides information on the field operation and general maintenance of the Multi RAE PID. Review of the information contained herein will ensure that this type of field monitoring equipment will be properly utilized. Review of the owner's instruction manuals is a necessity for more detailed descriptions.

III. Definitions

Carbon Monoxide Sensor (CO) - Carbon Monoxide concentration in ppm.

Volatile Organic Compound (VOC) – VOC concentration in ppm

Lower Explosive Limit (LEL) - Combustible gas is expressed as a percent of the lower explosive limit.

Hydrogen Sulfide Sensor (H₂S) - Hydrogen Sulfide concentration in ppm.

Oxygen Sensor (OXY) - Oxygen concentration as a percentage.

ppm - parts per million: parts of vapor or gas per million parts of air by volume.

IV. Procedures

The PID operates on the principle that most organic compounds and some inorganic compounds are ionized when they are bombarded by high-energy ultraviolet light. The air sample is drawn across a UV lamp using a pump or a fan. The energy of the lamp determines whether a particular chemical will be ionized. Each chemical compound has a unique photoionization potential (PIP). When the UV light energy is greater than the ionization potential of the chemical, ionization will occur. All PID readings are relative to the calibration gas, usually isobutylene.

It is important to calibrate the PID in the same temperature and elevation that the equipment will be used, and to determine the background concentrations in the field

1

before taking measurements. For environments where background readings are high, factory zero calibration gas should be used.

Note: For volatile and semi-volatile compounds, knowing the PIP is critical in determining the appropriate instrument to use when organic vapor screening. Consult the QAPP and manufacturer's manual to determine that the proper instrument has been selected for the contaminate vapors of interest. If an expected compound at a site has a PIP less than 11.7 eV, it is possible to use a PID. If the ionization potential is greater than 11.7 eV, a flame-ionization detector is required.

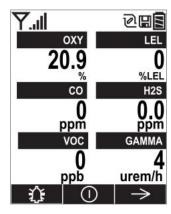
The following subsections will discuss Mini RAE calibration, operation, and maintenance. These sections, however, do not take the place of the instruction manual.

A. Calibration

For Multi RAE configured with O₂, LEL, H₂S, CO, sensors and a 10.6 eV PID Lamp.

Start up Instrument

- Press Mode button
- A RAE Systems logo (or a company name) should appear first. This is followed by a progression of screens that tell you the MultiRAE's current settings:
 - o Product name and model number, air flow type, and serial number
 - Application firmware version, build date, and build time
 - Sensor firmware, build date, build time
 - Installed sensors (including serial number/production/expiration/calibration date and alarm limit settings)
 - Current date, time, temperature, and relative humidity
 - User mode and operation mode
 - Battery type, voltage, shutoff voltage
 - Alarm mode and alarm settings
 - o Datalog period (if it is activated) and interval
 - Policy Enforcement settings (whether calibration and/or bump testing are enforced)
- Then the MultiRAE's main reading screen appears. It may take a few minutes for sensors to show a reading, so if any have not warmed up by the time the main screen is shown, you will see "- -" instead of a numerical value until the sensor provides data (typically less than 2 minutes). Then it displays instantaneous readings similar to the following screen (depending on the sensors installed) and is ready for use.



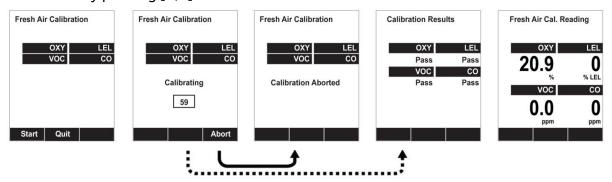
Calibration Check and Adjustment

Zero Calibration

• At the Calibration Menu, select "Fresh Air." Press [Y/+] once to enter the fresh air calibration sub-menu.



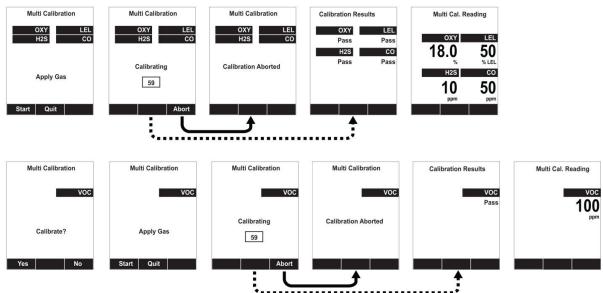
- Press [Y/+] to start fresh air calibration
- A countdown screen appears. You can abort the calibration at any time during the countdown by pressing [N/-].



- Note: Dotted line indicates automatic progression
- If the calibration is not aborted, the display shows the sensor names and tells you
 whether the fresh air calibration passed or failed, followed by the sensors' fresh air
 readings

Multi Sensor Span Calibration

- Depending on the configuration of your MultiRAE and span gas you have, you can
 perform a span calibration simultaneously on multiple sensors. You can define which
 sensors are calibrated together using the Multi Cal Select menu described in section
 8.3.2.9.
- In case all sensors in the instrument cannot be calibrated with the same gas, the MultiRAE will intelligently split the span calibration process into several steps and will provide menu prompts accordingly.
- At the Calibration Menu, select "Multi Sensor Span."
- Install the calibration adapter and connect it to a source of calibration gas.
- Start the flow of calibration gas.
- Press [Y/+] to start calibrating or wait for calibration to start automatically.
- A countdown screen is shown. You can abort the calibration at any time during the countdown by pressing [N/-].

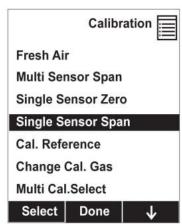


- Note: Dotted line indicates automatic progression
- If the calibration is not aborted, the display shows the sensor names and tells you whether the calibration passed or failed, followed by the sensor readings.

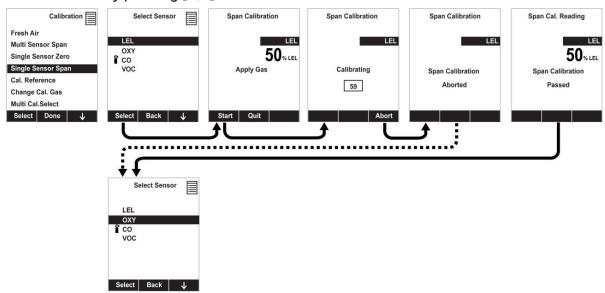
Single Sensor Span Calibration

- Instead of performing a span calibration on multiple sensors simultaneously, you can select a single sensor and perform a span calibration.
- To perform span calibration of an individual sensor, follow these steps:
- At the Calibration Menu, select "Single Sensor Span."
- Select a sensor to calibrate from the list.
- Install the calibration adapter and connect it to a source of calibration gas.
- Verify that the displayed calibration value meets the concentration specified on the gas cylinder.

Start the flow of calibration gas.



- Press [Y/+] to start calibrating or wait for calibration to start automatically.
- A countdown screen appears. You can abort the calibration at any time during the countdown by pressing [N/-].



- Note: Dotted line indicates automatic progression
- Select the done button when calibration is complete.
- CALIBRATION IS COMPLETE!

B. Operation

Due to the Multi RAE having many functions in terms of operation, it is recommended that you follow the operational procedures as outlined in the instruction manual.

C. Site Maintenance

After each use, the meter should be recharged and the outside of the instruments should be wiped clean with a soft cloth.

D. Scheduled Maintenance

<u>Function</u> <u>Frequency</u>

Check alarm and settings Monthly/before each use

Clean screens and gaskets around sensors Monthly

Replace sensors Biannually or when calibration is

unsuccessful

V. Quality Assurance Records

Quality assurance records will be maintained for each air monitoring event. The following information shall be recorded in the field logbook.

- Identification Site name, date, location, CTO number, activity monitored, (surface water sampling, soil sampling, etc), serial number, time, resulting concentration, comments and identity of air monitoring personnel.
- Field observations Appearance of sampled media (if definable).
- Additional remarks (e.g, Multi RAE had wide range fluctuations during air monitoring activities.)

VI. References

Multi RAE User's Guide, RAE Systems, Revision C, May 2013.



Groundwater Sampling for Per- and Polyfluoroalkyl Substances (PFAS)

I. Purpose and Scope

This SOP provides guidelines for groundwater sample collection for samples that will be analyzed for per- and polyfluoroalklyl substances (PFAS) via LC/MS/MS Compliant with the most recent version of the Quality Systems Manual (QSM) for which the lab is certified. This SOP should be used in conjunction with approved region-specific groundwater sampling SOPs which provide methods for general and low-flow groundwater sampling. In cases in which information in this SOP conflicts with region-specific groundwater sampling SOPs, this SOP will supersede the information in the general SOPs.

Standard techniques for collecting representative samples are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program. Materials, equipment, and procedures may vary; refer to the Sampling and Analysis Plan and operator's manuals for specific details.

II. Equipment and Materials

A. Equipment and Materials Required

- If installing wells, ensure driller does not use polytetrafluoroethylene (PFTE)-containing drill lube or other drilling lubes containing PFAS. Biolube has been determined to be an acceptable drilling lube for installing wells where PFAS may be of concern. Additionally, Waterra surge blocks have been confirmed to not contain PFAS and may be used for development.
- Groundwater sampling equipment
 - PFAS-free tubing (avoid Teflon, Viton, PTFE and other fluorinated compounds)
 - High density polyethylene tubing (unlined)
 - If Masterflex tubing is needed for peristaltic pumps, Cole Parmer C-Flex (06424 series) and
 Tygon E-3603 (06509 series) are suitable options
 - PFAS-free Bailer (if using a bailer¹)

¹ Geotech and Waterra offer PFAS free bailer options

- PFAS-free Pump such as:
 - Geotech PFAS-free Portable Bladder Pump (note, most bladder pumps include a Teflon-lined bladder, but Geotech currently has one model which is Teflon-free).
 - Panacea P120 or P125. The P200 Stainless Steel Pump may also be used, but the standard model contains Teflon at the tube connection. If you are using this Panacea model, you must request one with the "PTFE-free thread sealant option."
 - Waterra stainless foot-valve
 - QED Sample Pro
 - Monsoon or Mega Monsoon submersible pump
 - Grundfos Rediflo2 (this pump contains small Teflon components, but has not been shown to leach, it is less preferable than the other options)
 - Peristaltic pump (may be suitable for shallow locations)
- Groundwater sample containers (high density polyethylene [HDPE] bottle with HDPE screwcap), sample bottles should not be glass as glass may sorb PFAS. Sample bottle caps should not contain Teflon. Notify your project manager (PM) if bottles provided by the lab are glass or contain Teflon parts.
- Laboratory prepared deionized, certified PFAS-free water for field blank collection
- PFAS-free shipping supplies (labels [if available]², coolers, and ice)
- Loose leaf paper without waterproof coating or a spiralbound notebook (not waterproof) or tablet (see tablet use notes below)
- Metal clip board (if using loose-leaf paper)
- Pen (not Sharpie)
- Nitrile or latex gloves

B. Equipment and Materials to Avoid During Sampling

Equipment and materials used to collect groundwater samples should not contain any fluorinated compounds, Teflon, or synthetic rubber with fluoropolymer elastomers (e.g., Viton).

Specifically, the following material should be avoided during sampling:

- Gore-Tex brand or similar high-performance outdoor clothing, clothing treated with ScotchGuard brand or similar water repellent, fluoropolymer-coated Tyvek, wrinkle-resistant fabrics, and fireresistant clothing with fluorochemical treatment or anything advertised as water repellant.
- Weather-proof log books with fluorochemical coatings.
- New clothing that has been washed fewer than six times.

² Efforts will be made to obtain PFAS-free labels; however, information on labels is scarce and labels are frequently mounted on PFAS-coated paper to allow for easy removal.

The sample collection area should be clear of the following items:

- Pre-packaged food wrappers (e.g., fast food sandwich wrappers, pizza boxes, etc.)
- Microwave popcorn bags
- Blue ice containers
- Non-Stick aluminum foil
- Kim-Wipes
- Sunscreen, insect repellant and other personal hygiene products that may contain PFAS

Research which has not yet been published has allowed us to generate a list of sunscreens and insect repellents which do not contain fluorine. Check with Bill Diguiseppi or Laura Cook on recommendations (because the research is not ours, it cannot be released externally at this time).

The use of electronics (e.g., cell phones and tablets) should be avoided without the implementation of precautionary measures outlined below:

• All devices should be used with clean, ungloved hands and an approved stylus (if desired).

Following the use of a device, hands must be washed with soap and water and clean gloves should be used prior to contact with sampling equipment (bottleware, tubing, etc.).

III. Procedures and Guidelines

Wash hands with dish detergent before sampling and don nitrile gloves. Do not use Kleen Guard powder free nitrile gloves which were shown in research to contain fluorine

Follow Navy CLEAN SOPs for low-flow or conventional groundwater sample collection, depending on site requirements.

A. Sample Collection

Once water quality parameters have stabilized for low-flow purging, samples can be collected. For conventional purging, if water quality parameters do not stabilize, a minimum of 3 well volumes must be purged prior to sample collection.

The steps to be followed for sample collection are as follows:

- 1. Ensure that the end of the tubing does not touch the ground or equipment. Remove the cap from the sample bottle. Position the sample bottle under the end of the tubing.
- 2. Fill the bottle. Do not fill the bottle past the middle of the bottle shoulder. Samples do not need to be collected headspace free.
- 3. Affix labels after bottles have been closed; collect only one sample at a time to avoid mislabeling. Pack the sample on ice immediately for shipment to the offsite laboratory. Avoid packing materials that may contain fluorine. Unpublished research has allowed us to generate a list of packing materials which do not contain fluorine. Please contact Bill Diguiseppi or Laura Cook for recommendations (because the research is not ours, it cannot be released externally at this time).

B. Equipment Decontamination

Whenever possible, use disposable equipment when collecting groundwater samples. If reusable equipment must be used, the equipment must be cleaned/decontaminated between uses. Alconox and Liquinox soap are acceptable for cleaning/decontaminating reusable equipment at PFAS sites. Any water

used for cleaning/decontamination must be certified PFAS-free by a laboratory. Consider triple-rinsing. Once decontaminated, wrap equipment in plastic bags (such as Ziploc) or un-coated aluminum foil, and store away from potential PFAS sources.

Use of Water Quality Equipment and Water Level Indicators

Water quality meters typically do not contain PFAS. However, consistent with general sampling SOPs, disconnect the water quality meter prior to sampling. Some water level indicators do contain small polyvinylidene fluoride (a PFAS constituent for which we do not currently monitor) or less frequently, Teflon, components, but we have not noted cross contamination from water level indicators at any sites. The Durham Geoslope Water Level Indicators and the Solinst Model 101 with the P2 meter have been shown to be fluorine free.

IV. References

United States Environmental Protection Agency (USEPA), 2009. *Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/ Tandem Mass Spectrometry (LC/MS/MS)*. September.

United States Navy, 2020. Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/November 2020 Update. November.

United States Navy, 2015. Navy Drinking Water Sampling Policy for Perfluorochemicals: Perfluoroctane Sulfonate and Perfluoroctanoic Acid. September.



DRAFT FINAL STANDARD OPERATING PROCEDURE - Navy CLEAN PROGRAM

Rotosonic Groundwater Sample Collection for Per- and Polyfluoroalkyl Substances

I. Purpose and Scope

This SOP provides guidelines for groundwater sample collection using rotosonic drilling technology for samples that will be analyzed for per- and polyfluoroalklyl substances (PFAS) via LC/MS/MS Compliant with QSM 5.1 Table B-15.

Standard techniques for collecting representative samples are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program under Contract N62470-16-D-9000. Materials, equipment, and procedures may vary; refer to the Sampling and Analysis Plan and operator's manuals for specific details.

II. Equipment and Materials

Equipment and Materials Required

- Sonic drill sampling rods and retractable stainless-steel screen without PFAS-containing components (Avoid Teflon, Viton, PTFE and all other fluorinated compounds)
- PFAS-free tubing (avoid Teflon, Viton, PTFE and other fluorinated compounds)
 - High density polyethylene tubing (unlined)
 - Masterflex tubing, Cole Parmer C-Flex (06424 series) and Tygon E-3603 (06509 series) are suitable options
- Water quality meter (Horiba® or YSI®)
- PFAS-free bladder pump
- Pre-cleaned sample containers
- Air monitoring and water quality instruments (as needed)
- Personal protective equipment
- Groundwater sample containers (high density polyethylene [HDPE] with HDPE screw cap [no Teflon caps])
- PFAS-free shipping labels (if available¹) materials
- Loose leaf paper or a wire-bound notebook without waterproof coating
- Metal clipboard
- Pen (not Sharpie)
- Nitrile or Latex gloves
- Laboratory prepared deionized, certified PFAS-free water for field blank collection
- PVC casing and screen for temporary well

¹ Efforts will be made to obtain PFAS-free labels; however, information on labels is scarce and labels are frequently mounted on PFAS-coated paper to allow for easy removal.



Ensure the driller has not used and will not use drilling lube containing polytetrafluoroethylene (PFTE) or any other fluorine-containing substance. Biolube has been determined to be an acceptable substitute.

Equipment and Materials to Avoid During Sampling

Equipment and materials used to collect groundwater samples should not contain any fluorinated compounds, Teflon, or synthetic rubber with fluoropolymer elastomers (e.g., Viton).

Specifically, the following material should be avoided during sampling:

- Gore-Tex brand or similar high-performance outdoor clothing, clothing treated with ScotchGuard brand or similar water repellent, fluoropolymer-coated Tyvek, wrinkleresistant fabrics, and fire-resistant clothing with fluorochemical treatment or anything advertised as water repellant.
- Weather-proof log books with fluorochemical coatings
- New clothing that has been washed fewer than six times

The sample collection area should be clear of the following items:

- Pre-packaged food wrappers (e.g., fast food sandwich wrappers, pizza boxes, etc.)
- Microwave popcorn bags
- Blue ice containers
- Non-stick aluminum foil
- Kim-Wipes
- Sunscreen, insect repellant and other personal hygiene products that may contain PFAS

Research which has not yet been published has allowed us to generate a list of sunscreens and insect repellents which do not contain fluorine. Check with Bill Diguiseppi or Laura Cook on recommendations (because the research is not ours, it cannot be released externally at this time).

III. Procedures and Guidelines

Wash hands with dish detergent before sampling and don nitrile gloves. Do not use Kleen Guard powder free nitrile gloves which were shown in research to contain fluorine.

Once the area has been determined to be free of materials potentially containing PFAS, these steps can be followed to collect the sonic groundwater sample:

- 1) Decontaminate slotted lead rod and other downhole equipment in accordance with SOP *Decontamination of Personnel and Equipment* and this SOP.
- 2) Advance the drive-ahead sampler to the desired sampling depth and withdraw the rods 2 to 3 feet to expose the retractable screen to the aquifer formation. Ensure that the screened lead rod has been inserted to the desired sampling depth. If using temporary well casing, deploy PVC screen and casing to desired sampling depth.
- 3) Deploy your sampling pump to mid-screen.
- 4) Purge the volume of water added to the well during drilling. If no water has been used, purge three well volumes from the casing. When using the drive-ahead sampler, calculate



the well volume as the volume of water in the drill rod (bullet a). When using a temporary well, calculate the well volume as the volume of water in the casing plus the volume of water in the sand pack (bullet b).

- a) To calculate the volume of water in the drill rod, use the appropriate coefficient for the rod diameter given under "Well Casing Volume per Foot of Depth" on the attached reference sheet (Geotech, 2019). Then use the following formula:
 - V (gallons) = coefficient * h (where h is the height of the water column in feet)
- b) To calculate the volume of the temporary well casing and sand pack, first calculate the volume of the casing as above. Then calculate the volume of the annulus using the coefficients given under "Volume of Annulus" on the reference sheet (Geotech, 2019). The volume of the annulus is then multiplied by the porosity of the sand pack (a porosity of 0.35 can be assumed for this case). This is summarized in the following formula:
 - V (gallons) = V_casing + (0.35 * V_annulus)
- c) Note: If the sample interval / well is unable to produce enough water to generate three well volumes or if it goes dry, then the volume purge method will not be used. In this case, the casing will be pumped dry and allowed to recharge to 90% of the static water level (time permitting) before sampling.
- Collect and record one set of water quality parameters prior to sampling.
- 6) Fill all sample containers. Samples should be collected in accordance with SOP *Groundwater Sampling when Analyzing for Per- and Polyfluoroalkyl Substances (PFAS)*. Affix labels after bottles have been closed; collect only one sample at a time to avoid mislabeling.
- 7) Remove and discard polyethylene sampling tubing from the rods. Withdraw PVC temporary PVC screen and casing.

Equipment Decontamination

Ensure that the sonic rig operator thoroughly completes the decontamination process between sampling locations. Do not use water from the facility (e.g. fire hydrants) if there is a possibility that the water available is contaminated with PFAS.

Whenever possible, use disposable equipment when collecting groundwater samples. If reusable equipment must be used, the equipment must be cleaned/decontaminated between uses. Alconox and Liquinox soap are acceptable for cleaning/decontaminating reusable equipment at PFAS sites. Any water used for cleaning/decontamination must be certified PFAS-free by a laboratory. Consider triple-rinsing. Once decontaminated, wrap equipment in plastic bags (such as Ziploc), and store away from potential PFAS sources.

Use of Water Quality Equipment

Water quality meters typically do not contain PFAS. However, consistent with general sampling SOPs, disconnect the water quality meter prior to sampling.



References

Geotech, 2019. Water Measure Tables. Web. 28 March 2019.

United States Environmental Protection Agency (USEPA), 2009. *Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS)*. September.

United States Navy, 2015. *Navy Drinking Water Sampling Policy for Perfluorochemicals: Perfluoroctane Sulfonate and Perfluoroctanoic acid.* September.



Soil Sampling for Per- and Polyfluoroalkyl Substances

I. Purpose

This SOP provides guidelines for soil sample collection and handling for samples that will be analyzed for per- and polyfluoroalkyl substances (PFAS) via LC/MS/MS Compliant with the most recent version of the Quality Systems Manual (QSM) for which the lab is certified at the time of analysis. Standard techniques for collecting representative samples are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program. Materials, equipment, and procedures may vary; refer to the Sampling and Analysis Plan and operator's manuals for specific details.

II. Equipment and Materials

A. Equipment and Materials Required

A hand auger or other device that can be used to remove the soil from the ground. Stainless steel tools, carbon steel tools, or steel DPT tooling with acetate sleeves are preferred for PFAS sampling. Avoid any sampling materials containing PFAS (such as Teflon, Viton, PTFE, or other fluorinated compounds). Any plastic sampling materials should be evaluated thoroughly before selection to ensure they are fluorine-free.

A stainless steel spatula or fluorine-free disposable plastic scoop should be used to remove material from the sampling device.

Unpainted wooden stakes or pin flags

Fiberglass measuring tape (at least 200 feet in length)

GPS Unit

- PFAS-free labels (if available¹) shipping materials
- Loose leaf paper or a wire-bound notebook without waterproof coating or tablet (see notes on tablet use below)
- Metal clipboard (if using loose-leaf paper)
- Pen (not Sharpie)

Personal protection equipment (rubber or latex gloves, boots, etc.). Check with your SME prior to selecting PPE to ensure there are no fluorine-containing components.

¹ Efforts will be made to obtain PFAS-free labels; however, information on labels is scarce and labels are frequently mounted on PFAS-coated paper to allow for easy removal.

Sample jars (sample jars should be made of high density polyethylene (HDPE) as glass jars may sorb PFAS, please notify the project manager [PM] if glass jars are provided by the lab). Sample containers should not contain Teflon lids.

Laboratory-prepared deionized, certified PFAS-free water for field blank collection

B. Equipment and Materials to Avoid During Sampling

Equipment and materials used to collect soil samples should not contain any fluorinated compounds including Teflon or synthetic rubber with fluoropolymer elastomers (e.g. Viton).

If a driller is supporting collection of soil samples in split spoons or acetate DPT sleeves, ensure the driller has not used and will not use drilling lube containing polytetrafluoroethylene (PTFE) or any other fluorine-containing substance. Biolube has been determined to be an acceptable substitute.

Specifically, the following material should be avoided during sampling:

- Gore-Tex brand or similar high-performance outdoor clothing, clothing treated with ScotchGuard brand or similar water repellent, fluoropolymer-coated Tyvek, wrinkle-resistant fabrics, and fire-resistant clothing with fluorochemical treatment or anything advertised as water repellant.
- Weather-proof log books with fluorochemical coatings.
- New clothing that has been washed fewer than six times.

The sample collection area should be clear of the following items:

- Pre-packaged food wrappers (e.g., fast food sandwich wrappers, pizza boxes, etc.)
- Microwave popcorn bags
- Blue ice containers
- Non-stick aluminum foil
- Kim-Wipes
- Sunscreen, insect repellant and other personal hygiene products that may contain PFAS

Contact your PFAS SME for an approved list of sunscreens and insect repellants.

The use of electronics (e.g., cell phones and tablets) should be avoided without the implementation of precautionary measures outlined below:

• All devices should be used with clean, ungloved hands and an approved stylus (if desired).

Following the use of a device, hands must be washed with soap and water and clean gloves should be used prior to contact with sampling equipment (bottleware, tubing, etc.).

III. Procedures and Guidelines

Once the area has been determined to be free of materials potentially containing PFAS, these steps can be followed to collect the soil samples:

Wear protective gear, as specified in the Health and Safety Plan.

To locate samples, identify the correct location using the pin flags or stakes. Proceed to collect a sample from the undisturbed soil adjacent to the marker following steps C and D. If markers are not present, the following procedures will be used.

A. Samples On A Grid

- 1. Use measuring tape to locate each sampling point on the first grid line as prescribed in the sampling plan. As each point is located, drive a numbered stake in the ground and record its location on the site map and in the field notebook/clipboard.
- 2. Proceed to sample the points on the grid line.
- 3. Measure to location where next grid line is to start and stake first sample. For subsequent samples on the line take two orthogonal measurements: one to the previous grid line, and one to the previous sample on the same grid line.
- 4. Proceed to sample the points on the grid line as described in Section C below.
- 5. Make sure to stake location after sample collection in case professional surveying is to be completed.
- 6. Repeat 1c and 1e above until all samples are collected from the area.
- 7. Or, a GPS unit can be used to identify each location based on map coordinates, if available.

B. Non-Grid Samples

- 1. Use measuring tape to position sampling point at location described in the sampling plan by taking two measurements from fixed landmarks (e.g., corner of house and fence post).
- 2. Note measurements, landmarks, and sampling point on a sketch in the field notebook, and on a site location map.
- 3. Proceed to sample as described in Section C below.
- 4. Make sure to stake location after sample collection in case professional surveying is to be completed.
- 5. Repeat 2a through 2d above until all samples are collected from the area.
- 6. Or, a GPS unit can be used to identify each location based on map coordinated, if available.

To the extent possible, differentiate between fill and natural soil. If both are encountered at a boring location, sample both as prescribed in the field sampling plan. Do not locate samples in debris, tree roots, or standing water. In residential areas, do not sample in areas where residents' activities may impact the sample (e.g., barbecue areas, beneath eaves of roofs, driveways, garbage areas). If an obstacle prevents sampling at a measured grid point, move as close as possible, but up to a distance of one half the grid spacing in any direction to locate an appropriate sample. If an appropriate location cannot be found, consult with the Field Team Leader (FTL). If the FTL concurs, the sampling point may be deleted from the program. The FTL will contact the CH2M HILL PM immediately. The PM and Navy Technical Representative (NTR) will discuss whether the point should be deleted from the program. If it is deleted, the PM will follow-up with the NTR in writing.

C. Collecting Samples Using Hand Tools

1. Use a decontaminated stainless steel scoop/trowel or disposable plastic scoop to scrape away surficial organic material (grass, leaves, etc.) adjacent to the stake. New disposable scoops or trowels may also be used to reduce the need for equipment blanks if the disposable scoops have been confirmed by your project PFAS subject matter expert (SME) to be PFAS free.

2. If sampling:

- Surface soil: Obtain soil sample by scooping soil using the augering scoop/trowel, starting from the surface and digging down to a depth of about 6 inches, or the depth specified in the workplan.
- Subsurface soil: Obtain the subsurface soil sample using an auger down to the depths prescribed in the field sampling plan.
- 3. Record lithologic description and any pertinent observations (such as discoloration) in the field notebook/clipboard.
- 4. Empty the contents of the scoop/trowel into a decontaminated stainless steel pan or dedicated sealable bag.
- 5. Repeat this procedure until sufficient soil is collected to meet volume requirements.
- 6. Homogenize cuttings in the pan using a decontaminated stainless steel utensil.
- 7. Transfer sample for analysis into appropriate containers with a decontaminated utensil. Affix labels after bottles have been closed; collect only one sample at a time to avoid mislabeling.
- 8. Immediately upon collection, all samples for chemical analysis are to be placed in a closed container on ice unless it is not possible to do so. Although unusual and uncommon, there may be instances where it is not possible to have containers with ice at the sample location. In these instances, the samples should be placed on ice as soon as practical and during the time between collection and placing the samples on ice, the samples should be kept as cool as possible.
- 9. Backfill the hole with soil removed from the borehole. To the extent possible, replace topsoil and grass and attempt to return appearance of sampling area to its pre-sampled condition. For samples in non-residential, unmowed areas, mark the sample number on the stake and leave stake in place. In mowed areas, remove stake.

To Collect Samples Using DPT Methods

- 1. Decontaminate sampling tubes and other non-dedicated downhole equipment in accordance with SOP *Decontamination of Personnel and Equipment*. Ensure that decontamination water used is PFAS free (do not use water from fire hydrants on-base for steam cleaning unless the water has been demonstrated to be free of PFAS). Additionally, ensure that all decontaminated tools are wrapped in un-coated aluminum foil as needed.
- 2. Drive sampling tube to the desired sampling depth using the truck-mounted hydraulic percussion hammer. If soil above the desired depth is not to be sampled, first drive the lead rod, without a sampling tube, to the top of the desired depth.
- 3. Remove the rods and sampling tube from the borehole and remove the sampling tube from the lead rod.
- 4. Cut open the acetate liner using a specific knife designed to slice the acetate liners (see below).





- 5. Fill all sample containers, using a decontaminated or dedicated sampling implement. Label the containers and immediately place samples on ice for shipment to the laboratory.
- 6. Decontaminate all non-dedicated downhole equipment (rods, sampling tubes, etc.) in accordance with SOP Decontamination of Personnel and Equipment and ensure decontamination water is from a PFAS-free water source and if equipment is wrapped in aluminum foil that un-coated aluminum foil is used.
- 7. Backfill borehole at each sampling location with grout or bentonite and repair the surface with like material (bentonite, asphalt patch, concrete, etc.), as required.

D. Equipment Decontamination

Whenever possible, use disposable equipment when collecting soil samples. If reusable equipment must be used, the equipment must be cleaned/decontaminated between uses. Alconox and Liquinox soap are acceptable for cleaning/decontaminating reusable equipment at PFAS sites. Any water used for cleaning/decontamination must be certified PFAS-free by a laboratory. Consider triple-rinsing. Once decontaminated, wrap equipment in plastic bags (such as Ziploc) or un-coated aluminum foil, and store away from potential PFAS sources.

IV. References

United States Environmental Protection Agency (USEPA), 2009. *Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS)*. September.

United States Navy, 2020. Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/November 2020 Update. November.

United States Navy, 2015. Navy Drinking Water Sampling Policy for Perfluorochemicals: Perfluoroctane Sulfonate and Perfluoroctanoic Acid. September



STANDARD OPERATING PROCEDURE - Navy CLEAN PROGRAM

Management of Liquid Waste Containing Per- and Polyfluoroalkyl Substances (PFAS)

I. Purpose and Scope

This SOP provides guidelines for managing waste containing per- and polyfluoroalkyl substances (PFAS) in accordance with the *Interim Per- and Polyfluoroalkyl Substances (PFAS) Site Guidance for NAVFAC Remedial Project Managers (RPMs)/November 2020 Update* (guidance). This SOP should be used in conjunction with an Environmental and/or Waste Management Plan (EMP and/or WMP) approved by your Environmental Manager (EM). If you do not have a site-specific EMP, please contact your EM.

Standard procedures for managing liquid waste during PFAS investigation are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program.

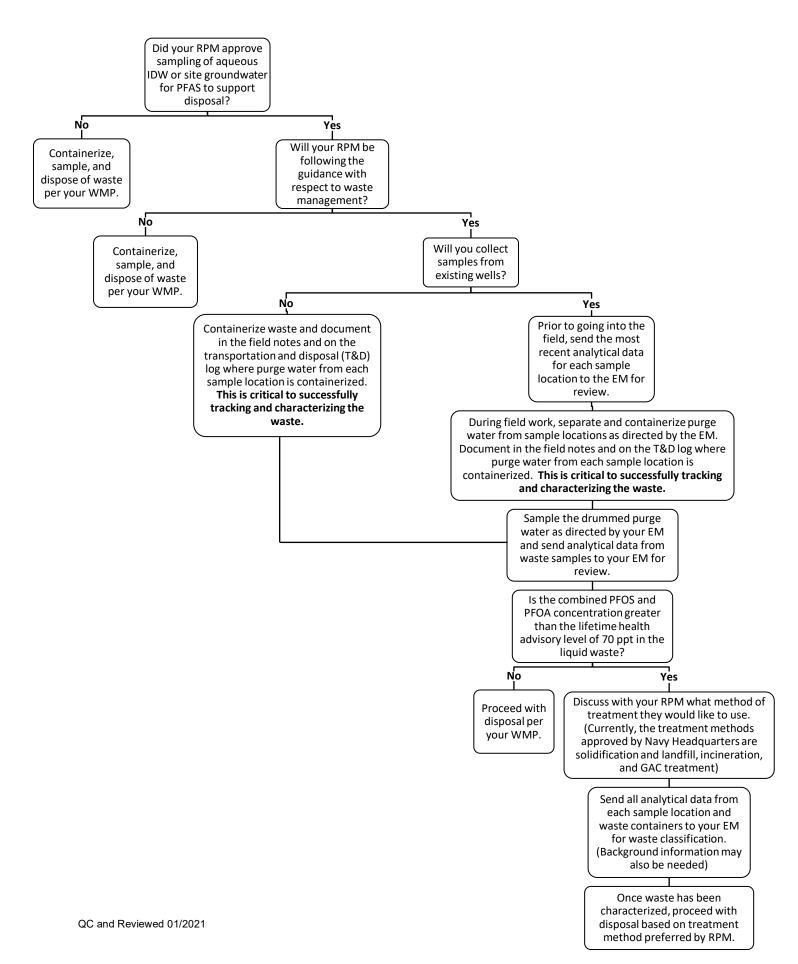
Currently, PFAS are not regulated as a hazardous waste in US EPA regulations (state and territory rules may vary). Treatment of liquid waste containing PFAS, as recommended by the guidance, is a client directed action. When and how it is implemented will be left to the discretion of the individual RPMs. These project specific actions will be communicated with the Project Manager (PM) and/or Activity Manager (AM).

II. Procedures and Guidelines

The following flowchart outlines the procedures required to manage liquid waste during PFAS investigations. Any deviations from this procedure must be approved by the EM.

QC and Reviewed 01/2021







Surface Water Sampling for Per- and Polyfluoroalkyl Substances

Purpose and Scope

This SOP provides guidelines for surface water sample collection for samples that will be analyzed for per- and polyfluoroalklyl substances (PFAS) via LC/MS/MS Compliant with the most recent version of the Quality Systems Manual (QSM) for which the lab is certified at the time of analysis. Standard techniques for collecting representative samples are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program. Materials, equipment, and procedures may vary; refer to the Sampling and Analysis Plan and operator's manuals for specific details.

II. Materials and Equipment

A. Equipment and Materials Required

- Open tube sampler
- Dip sampler
- Weighted bottle sampler (no glass)
- Hand pump without Teflon components
- Van Dorn sampler (Kemmerer cannot be used as it has Teflon caps)
- Depth-integrating sampler
- Peristaltic pump and PFAS-free tubing
- High density polyethylene tubing (unlined)
- Masterflex tubing such as Cole Parmer C-Flex (06424 series) and Tygon E-3603 (06509 series) are suitable options
- Sample containers (high density polyethylene [HDPE] with HDPE screw cap [no Teflon caps])
- PFAS-free labels (if available¹) and shipping materials
- Loose leaf paper or a wire-bound notebook without waterproof coating or tablet (see notes about tablet use below)

¹ Efforts will be made to obtain PFAS-free labels; however, information on labels is scarce and labels are frequently mounted on PFAS-coated paper to allow for easy removal.

- Metal clipboard (if using loose leaf paper)
- Pen (not Sharpie)
- Nitrile or Latex gloves (Do not use Kleen Guard powder free nitrile gloves which were shown in research to contain fluorine)
- Meters for specific conductance, temperature, pH, and dissolved oxygen

Equipment and materials used to collect surface water samples should not contain any fluorinated compounds including Teflon or synthetic rubber with fluoropolymer elastomers (e.g. Viton). Neoprene and rubber waders should not be an issue, however, check with your PFAS subject matter expert (SME) during field preparation to ensure all equipment is free of fluorine-containing components.

B. Equipment and Materials to Avoid During Sampling

Equipment and materials used to collect groundwater samples should not contain any fluorinated compounds, Teflon, or synthetic rubber with fluoropolymer elastomers (e.g., Viton).

Specifically, the following material should be avoided during sampling:

- Gore-Tex brand or similar high-performance outdoor clothing, clothing treated with ScotchGuard brand or similar water repellent, fluoropolymer-coated Tyvek, wrinkle-resistant fabrics, and fireresistant clothing with fluorochemical treatment or anything advertised as water repellant.
- Weather-proof log books with fluorochemical coatings.
- New clothing that has been washed fewer than six times.

The sample collection area should be clear of the following items:

- Pre-packaged food wrappers (e.g., fast food sandwich wrappers, pizza boxes, etc.)
- Microwave popcorn bags
- Blue ice containers
- Non-stick aluminum foil
- Kim-Wipes
- Sunscreen, insect repellant and other personal hygiene products that may contain PFAS

Contact your PFAS SME for an approved list of sunscreens and insect repellants.

The use of electronics (e.g., cell phones and tablets) should be avoided without the implementation of precautionary measures outlined below:

All devices should be used with clean, ungloved hands and an approved stylus (if desired).

Following the use of a device, hands must be washed with soap and water and clean gloves should be used prior to contact with sampling equipment (bottleware, tubing, etc.).

III. Procedures and Guidelines

Wash hands with dish detergent before sampling and don nitrile gloves. Do not use Kleen Guard powder free nitrile gloves which were shown in research to contain fluorine.

Before surface water samples are taken, all sampler assemblies and sample containers are cleaned and decontaminated as described in SOP *Decontamination of Personnel and Equipment* as well as this SOP (see below). Surface water samples collected from water bodies tidally influenced should be collected at

low tide and under low flow conditions to minimize the dilution of potential contaminants. Once the area has been determined to be free of materials potentially containing PFAS, follow the methods for surface water sample collection described below.

Surface water samples are collected manually by submerging a clean stainless steel or polypropylene container into the water body. Samples may be collected at depth with a covered bottle that can be removed with a tripline provided the bottle or bottle cap does not contain Teflon. The most common sampler types are sealable bottles, pond samplers, peristaltic pumps, and weighted bottle samplers. Pond samplers have a fixed or telescoping pole attached to the sample container. Weighted bottle samplers are lowered below water surface, where the attached bottle is opened, allowed to fill, and pulled out of the water. When retrieved, the bottle is tightly capped and removed from the sampler assembly. A specific type of weighted bottle sampler is the Van Dorn and is acceptable in most instances. The Kemmerer weighted bottle sampler cannot be used for PFAS sampling due to the Teflon caps.

A sample is taken with the following specific steps:

- 1. The location and desired depth for water sampling are selected.
- 2. The sample site is approached from downstream in a manner that avoids disturbance of bottom sediments as much as possible. The sample bottle is gently submerged with the mouth pointed upstream and the bottle tilted slightly downstream. Bubbles and floating materials should be prevented from entering the bottle. If using a Peristaltic pump, lower the tubing into the water to the desired depth.
- 3. For weighted bottle samplers, the assembly is slowly lowered to the desired depth. The bottle stopper is unseated with a sharp tug and the bottle is allowed to fill until bubbles stop rising to the surface.
- 4. When the bottle is full, it is gently removed from the water. If sample transfer is required, it should be performed at this time. Fill all sample containers to the middle of the bottle shoulder. Do not fill bottles completely. Affix labels after sample containers are closed; collect only one sample at a time to avoid mislabeling.
- 5. Measure dissolved oxygen, specific conductance, temperature, and pH at the sampling location.

A. Equipment Decontamination

Whenever possible, use disposable equipment when collecting surface water samples. If reusable equipment must be used, the equipment must be cleaned/decontaminated between uses. Alconox and Liquinox soap are acceptable for cleaning/decontaminating reusable equipment at PFAS sites. Any water used for cleaning/decontamination must be certified PFAS-free by a laboratory. Consider triple-rinsing. Once decontaminated, wrap equipment in plastic bags (such as Ziploc) or un-coated aluminum foil, and store away from potential PFAS sources.

Use of Water Quality Equipment

Water quality meters typically do not contain PFAS. However, consistent with general sampling SOPs, disconnect the water quality meter prior to sampling.

IV. References

United States Environmental Protection Agency (USEPA), 2009. *Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS)*. September.

United States Navy, 2015. Navy Drinking Water Sampling Policy for Perfluorochemicals: Perfluoroctane Sulfonate and Perfluoroctanoic Acid. September.



Sediment Sampling for Per- and Polyfluoroalkyl Substances

Purpose and Scope

This SOP provides guidelines for sediment sample collection and handling for samples that will be analyzed for per- and polyfluoroalklyl substances (PFAS) via LC/MS/MS Compliant with the most recent version of the Quality Systems Manual (QSM) for which the lab is certified at the time of analysis. Standard techniques for collecting representative samples are summarized. These procedures are specific to the Navy Comprehensive Long-term Environmental Action Navy (CLEAN) Program. Materials, equipment, and procedures may vary; refer to the Sampling and Analysis Plan and operator's manuals for specific details.

II. Equipment and Materials

A. Equipment and Materials Required

- Sample collection device (hand corer, scoop, dredge, grab sampler, or other suitable device). Check with your PFAS subject matter expert (SME) during field preparation to ensure all equipment is free of fluorine-containing components.
- Stainless steel spoon or spatula or fluorine-free plastic disposable scoop for media transfer
- Measuring tape
- GPS Unit
- PFAS-free labels (if available¹) shipping materials
- Loose leaf paper or a wire-bound notebook without waterproof coating
- Metal clipboard (if using loose-leaf paper)
- Pen (not Sharpie)
- Personal protection equipment (rubber or latex gloves, boots, hip waders, etc.). Check with your SME prior to selecting PPE to ensure there are no fluorine-containing components.
- Materials for classifying soils, particularly the percentage of fines
- Sample jars (high density polyethylene [HDPE] with HDPE screw cap [no Teflon caps])
- Laboratory prepared deionized, certified PFAS-free water for field blank collection

¹ Efforts will be made to obtain PFAS-free labels; however, information on labels is scarce and labels are frequently mounted on PFAS-coated paper to allow for easy removal.

B. Equipment and Materials to Avoid During Sampling

Equipment and materials used to collect soil samples should not contain any fluorinated compounds including Teflon or synthetic rubber with fluoropolymer elastomers (e.g. Viton).

Specifically, the following material should be avoided during sampling:

- Gore-Tex brand or similar high-performance outdoor clothing, clothing treated with ScotchGuard brand or similar water repellent, fluoropolymer-coated Tyvek, wrinkle-resistant fabrics, and fireresistant clothing with fluorochemical treatment or anything advertised as water repellant.
- Weather-proof log books with fluorochemical coatings.
- New clothing that has been washed fewer than six times.

The sample collection area should be clear of the following items:

- Pre-packaged food wrappers (e.g., fast food sandwich wrappers, pizza boxes, etc.)
- Microwave popcorn bags
- Blue ice containers
- Non-Stick aluminum foil
- Kim-Wipes
- Sunscreen, insect repellant and other personal hygiene products that may contain PFAS

Contact your PFAS SME for an approved list of sunscreens and insect repellants.

The use of electronics (e.g., cell phones and tablets) should be avoided without the implementation of precautionary measures outlined below:

• All devices should be used with clean, ungloved hands and an approved stylus (if desired).

Following the use of a device, hands must be washed with soap and water and clean gloves should be used prior to contact with sampling equipment (bottleware, tubing, etc.).

III. Procedures and Guidelines

Wash hands with dish detergent before sampling and don nitrile gloves. Do not use Kleen Guard powder free nitrile gloves which were shown in research to contain fluorine.

Once the area has been determined to be free of materials potentially containing PFAS, these steps can be followed to collect the sediment samples:

- 1. Field personnel will start downstream and work upstream to prevent contamination of unsampled areas. In surface water bodies that are tidally influenced, sampling will be performed at low tide and under low flow conditions to minimize the dilution of possible contaminants. Sediment sampling activities will not occur immediately after periods of heavy rainfall.
- 2. Make a sketch of the sample area that shows important nearby river features and permanent structures that can be used to locate the sample points on a map. Whenever possible, include measured distances from such identifying features. Also include depth and width of waterway, rate of flow, type and consistency of sediment, and point and depth of sample removal (along shore, mid-channel, etc.).
- 3. Note in the field book any possible outside sources of contamination; for example, the outlet to a drainage culvert in the water body near your sampling location.

- 4. Transfer sample into appropriate sample jars with a stainless steel utensil or plastic disposable scoop confirmed to be fluorine-free. Be especially careful to avoid the loss of the very fine clay/silt particles when collecting the sample. The fine particles have a higher adsorption capacity than larger particles. Minimize the amount of water that is collected within the sample matrix. Decant the water off the sample slowly and carefully to maximize retention of the very fine particles. The sampler's fingers should never touch the sediment. Classify the soil type of the sample using the Unified Soil Classification System, noting particularly the percentage of silt and clay. Affix the sample label to the container after the container has been closed; collect only one sample at a time to avoid mislabeling.
- 5. Rocks and other debris should be removed before placement in jars.
- 6. For channel sampling, be on the alert for submerged hazards (rocks, tree roots, drop-offs, loess silt and muck) which can make wading difficult.
- 7. Immediately upon collection, all samples are to be placed in a closed container on ice. Although unusual and uncommon, there may be instances where it is not possible to have containers with ice at the sample location. In these instances, the samples should be placed on ice as soon as practical and during the time between collection and placing the samples on ice, the samples should be kept as cool as possible.
- 8. Equipment Decontamination
 - Whenever possible, use disposable equipment when collecting sediment samples. If reusable
 equipment must be used, the equipment must be cleaned/decontaminated between uses.
 Alconox and Liquinox soap are acceptable for cleaning/decontaminating reusable equipment at
 PFAS sites. Any water used for cleaning/decontamination must be certified PFAS-free by a
 laboratory. Consider triple-rinsing. Once decontaminated, wrap equipment in plastic bags (such
 as Ziploc) or un-coated aluminum foil, and store away from potential PFAS sources.

IV. References

United States Environmental Protection Agency (USEPA), 2009. *Determination of Selected Perfluorinated Alkyl Acids in Drinking Water by Solid Phase Extraction and Liquid Chromatography/Tandem Mass Spectrometry (LC/MS/MS)*. September.

United States Navy, 2015. Navy Drinking Water Sampling Policy for Perfluorochemicals: Perfluoroctane Sulfonate and Perfluoroctanoic Acid. September.



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PLANNING FIELD SAMPLING ACTIVITIES

1.0 PURPOSE

This section sets forth standard operating procedures (SOPs) for planning and scheduling field sampling activities. This SOP shall also be used to determine the number and type of laboratory and field Quality Control (QC) samples required while working on U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) sites/projects, and to prepare and implement Task Order Field Sampling Plans (FSP). For information on the number and type of QC samples required for the various QC Levels, see SOPs III-A, Laboratory QC Samples (Water and Soil), III-B, Field QC Samples (Water and Soil), III-C Field and Laboratory QC Samples (Air).

2.0 PROCEDURES

To prepare a field sampling plan, designated personnel must identify the objectives of the sampling program, determine the number of samples to be collected for each matrix (see SOP I-A-2, *Development of Data Quality Objectives*), and select the analyses to be performed on each sample (see SOPs I-A-3, *Selection of Analytes* and I-A-4, *Analytical Methods Selection*). The duration of sampling for each matrix, the preferred sampling method, the method of shipment, and the type and quantity of supplies (such as coolers, coolant and packing material that will be needed for sample storage and transport) must also be determined. Finally, the number and type of decontamination water sources to be used for each phase of sampling must be identified. The methods of determining each of these elements are addressed below.

2.1 NUMBER OF SAMPLES

Designated project personnel shall determine the number of samples to be collected from each sample matrix (e.g., soil, water), and specify the type of sample analysis. SOPs I-A-2, *Development of Data Quality Objectives*, I-A-3, *Selection of Analytes*, and I-A-4, *Analytical Methods Selection*, shall be used to determine numbers and locations of samples, as well as appropriate analytical methods. These figures will be used to estimate the costs of sample analysis. They will also help determine the number and types of sample containers required; number of field duplicates, field replicates, equipment rinsates, performance evaluation (PE) samples, matrix spike/matrix spike duplicates (MS/MSD), and trip blanks to be collected, and the analyses to be performed on them for each matrix and analytical method; and the number of days required to perform sampling activities.

Sampling intervals for soil borings shall be selected on the basis of potential sources of contamination, the geologic and hydrologic complexity of the site, and the objectives of the sampling program. Areas of high contamination (for example, contamination in the capillary fringe) or complex geology or hydrogeology may require continuous sampling.

2.2 DURATION OF SAMPLING ACTIVITIES

The anticipated number of working days needed to complete field sampling activities shall be determined before fieldwork commences. A schedule should be developed that outlines the approximate number of samples to be collected each day, categorized by sample matrix, method of sample collection, and sample analysis (e.g., 28 soil samples collected using a hand auger and analyzed for organochlorine pesticides and chlorinated herbicides; 15 water samples collected using a bailer—7 analyzed for volatile organics and 8 analyzed for organic lead). This information will be used to determine the number of field equipment rinsate samples that will be collected (if any), the types of analyses to be performed on them, the number of MS/MSDs and field duplicates, equipment needs, and personnel.

2.3 NUMBER OF SAMPLES TO BE ANALYZED FOR VOLATILE ORGANICS

Prior to initiation of site sampling activities, designated personnel shall determine the number of samples to be analyzed for volatile organic compounds (VOCs). This information will be used to determine the approximate number of coolers that will contain samples to be analyzed for VOCs, which will in turn, dictate the number of VOC trip blanks needed, as specified in SOP III-B, *Field QC Samples (Water, Soil)*.

2.4 DECONTAMINATION WATER SOURCES

Prior to initiation of sampling activities, designated personnel shall determine the number and type of decontamination water sources. Decontamination water includes both potable water used for equipment washing, and deionized or distilled water used during the final equipment rinse. The locations of potable water supplies for field decontamination activities shall be identified and designated as the only sources to be used during site sampling activities. Similarly, the source(s) of deionized or distilled water shall be identified and designated as the only source(s) to be used during site sampling activities. The intent of this procedure is to reduce variability in equipment decontamination procedures and to make it possible to easily identify the source of contamination in the event that analysis of field blanks reveals the presence of contaminants of concern.

3.0 DOCUMENTATION

The number of samples to be collected, the proposed duration of sampling activities, the number of samples that will be analyzed for VOCs, and the number and type of decontamination water sources that will be used for field activities will be specified in the FSP and QAPP portions of the Work Plan prepared for each NAVFAC NW Task Order. Records of how this information is actually implemented during field activities will be maintained in field logbooks, as specified in SOP III-D, *Logbooks*.

4.0 REFERENCES

SOP I-A-2, Development of Data Quality Objectives

SOP I-A-3, Selection of Analytes

SOP I-A-4, Analytical Methods Selection

SOP II-B, Field QC Samples (Water and Soil)

SOP III-A, Laboratory QC Samples (Water and Soil)

SOP III-B, Field QC Samples (Water, Soil)

SOP III-C Field and Laboratory QC Samples (Air)

SOP III-D, Logbooks

5.0 ATTACHMENTS

None.



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IDW MANAGEMENT

1.0 PURPOSE

This standard operating procedure (SOP) describes the activities and responsibilities of the U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) and their subcontractors with regard to management of investigation-derived waste (IDW). The purpose of this procedure provides guidance for the minimization, handling, labeling, temporary storage, and inventory of IDW generated during site investigations and remediation projects conducted under the direction of NAVFAC NW. **Each base may have specific required procedures**. These procedures are made available to the contractor through the NAVFAC Naval Technical Representative (NTR) or other government point of contact. This SOP is also applicable to personal protective equipment (PPE), sampling equipment, decontamination fluids, non-IDW trash, non-indigenous IDW, and hazardous waste and other regulated wastes generated during implementation of site investigations and removal or remedial actions. The information presented will be used to prepare and implement Work Plans (WP), Field Sampling Plans (FSP), and Waste Management Plans (WMPs) for IDW-related field activities.

2.0 PROCEDURES

The procedures for IDW management in the field are described below in Sections 2.1 to 2.5. The implementation of these procedures requires Remedial Project Managers (RPMs), Field Managers, their designates and subcontractors to perform the following tasks:

- Minimize generation of IDW,
- Segregate IDW,
- Properly handle IDW containers,
- Properly label IDW containers,
- Apply good management practices in storing IDW drums and containers,
- Prepare IDW drum inventories,
- Update and Report changes to IDW drum inventories,
- Perform inspections of IDW containers and storage areas, as required,
- Prepare IDW containers for proper off-site transportation and disposition, as required.

2.1 IDW MINIMIZATION

Field Managers and their designates shall minimize the generation of onsite IDW to reduce the need for special storage or disposal requirements that may result in substantial additional costs and provide little or no reduction in site risks (EPA 1992). The volume of IDW shall be reduced, by applying minimization

practices throughout the course of site investigation activities. These minimization strategies include:
1) material substitution; 2) using proper low-volume drilling techniques; 3) using disposable sampling and PPE; 4) using bucket and drum liners; and 5) segregating non-contaminated IDW and trash from contaminated IDW. Waste minimization strategies and types of IDW expected to be generated shall be documented in the appropriate project plans.

2.1.1 Material Substitution

Material substitution consists of selecting materials that degrade readily or have reduced potential for chemical impacts to the site and the environment. An example of this practice is the use of biodegradable detergents (e.g., Alconox® or non-phosphate detergents) for decontamination of non-consumable PPE and sampling equipment. In addition, field equipment decontamination can be conducted using isopropyl alcohol rather than hexane or other solvents (for most analytes of concern), to reduce the potential onsite chemical impacts of the decontamination solvent. Decontamination solvents shall be selected carefully so that solvents, and their known decomposition products, do not result in generation of RCRA hazardous waste.

2.1.2 Drilling Methods

Drilling methods that minimize potential IDW generation should be given priority. Sonic, Hollow stem auger and air rotary methods should be selected, where feasible, over mud rotary methods. Mud rotary drilling produces waste drilling mud, while hollow stem and air rotary drilling methods produce relatively low volumes of soil waste. Sonic drilling produces the least amount of waste. Small diameter borings and cores shall be used when soil is the only matrix to be sampled at the boring location; the installation of monitoring wells requires the use of larger diameter borings.

Soil, sludge, or sediment removed from borings, containment areas, and shallow test trenches shall not be returned to the source, unless allowed by regulation and included in the approved WP, FSP, or WMP.

2.1.3 Decontamination Fluids

The use of disposable sampling equipment, such as plastic bailers, trowels, and drum thieves (which do not require decontamination) minimizes the quantity of decontamination fluids generated. In general, decontamination fluids, and well development and purge water, should not be minimized because the integrity of the associated analytical data may be affected.

2.1.4 PPE and Disposable Sampling Equipment

Visibly soiled PPE and disposable sampling equipment shall be segregated from non-visibly soiled PPE and sampling equipment. Where investigation involves potentially hazardous waste or other regulated wastes, visibly soiled PPE and disposable sampling equipment may require decontamination. The Field Manager shall use best professional judgment to determine if decontamination is appropriate. This determination should be included in the approved WP, FSP, or WMP. If decontamination is performed, PPE and disposable sampling equipment generated in the decontamination process may be double-bagged and disposed of as non-hazardous waste.

2.1.5 Liners

Bucket liners can be used in the decontamination process to reduce the volume of solid IDW-generated and reduce costs on larger projects. The plastic bucket liners can be crushed into a smaller volume than the buckets, and only a small number of plastic decontamination buckets are required for the entire project. Larger, heavy-duty, 55-gallon drum liners can be used for heavily contaminated IDW to provide secondary containment, and reduce the costs of disposal and drum recycling. Drum liners may extend the containment life of the drums in severe climates and will reduce the costs of cleaning out the drums prior to recycling.

2.1.6 Segregation of non-IDW

All waste materials generated in the support zone are considered non-IDW trash. To minimize the total volume of IDW, all trash shall be separated from IDW, sealed in garbage bags, and properly disposed of offsite as municipal waste.

2.1.7 Monitoring Well Construction

Excess cement, sand, and bentonite grout prepared for monitoring well construction shall be kept to a minimum. Well construction shall be observed by Field Managers to ensure that a sufficient, but not excessive, volume of grout is prepared. Some excess grout may be produced. Unused grout that has not come in contact with potentially contaminated soil or ground water shall be considered non-hazardous trash and shall be disposed of offsite by the drilling subcontractor. Surplus materials from monitoring well installation, such as scrap PVC sections, used bentonite buckets, and cement/sand bags that do not come in contact with potentially contaminated soil, shall be considered non-IDW trash and shall be disposed of offsite by the drilling subcontractor.

2.1.8 Field Analytical Test Kits

IDW generated from the use of field analytical test kits consists of those parts of the kit that have been used and/or come into contact with potentially contaminated site media, or excess extracting solvents and other reagents. Potentially contaminated solid test kit IDW shall be contained in plastic bags and stored with PPE or disposable sampling equipment IDW from the same source area as soil material used for the analyses. The small volumes of waste solvents, reagents, and water samples used in field test kits should be segregated, and disposed of accordingly (based upon the characteristics of the materials, MSDS sheets, and as described in the WMP). Most other test kit materials should be considered non-IDW trash, and be disposed of as municipal waste.

2.2 SEGREGATION OF IDW BY MATRIX AND LOCATION

To facilitate subsequent IDW screening, sampling, classification and/or disposal, IDW shall generally be segregated by matrix and source location at the time it is generated. Each drum of solid IDW shall be completely filled, when possible. For liquid IDW, drums should be left with headspace of approximately 5% by volume to allow for expansion of the liquid and potential volatile contaminants. IDW from each distinct matrix shall be stored in a single drum (e.g., soil, water or PPE shall not be mixed in one drum). In general, IDW from separate sources should not be combined in a single drum.

It is possible that monitoring well development and purge water will contain suspended solids, which will settle to the bottom of the storage drum as sediment. Significant observations on the turbidity or sediment load of the development or purge water shall be included in the logbook and reported in attachments to

the quarterly drum inventory report (see SOP III-D, *Logbooks* and Section 2.5). To avoid having mixed matrices in a single drum (i.e., sediment and water), it may be necessary to decant the liquids into a separate drum, after the sediments have settled out. This segregation may be accomplished during subsequent IDW sampling activities or during consolidation in a holding tank prior to disposal. Disposal of liquid IDW into the sanitary sewer shall only occur if approved by the appropriate regulatory agencies, municipal entities, and Naval installation. Appropriate precautions per the approved Health and Safety Plan (HASP) shall be implemented to ensure worker protection during these activities.

Potentially contaminated well construction material shall be placed in separate containers. Soil, sediment, sludge, or liquid IDW shall be segregated from potentially contaminated waste well construction materials. Potentially contaminated well construction materials from different monitoring wells shall not be commingled.

Potentially hazardous PPE and disposable sampling equipment shall be segregated from other IDW. PPE from generally clean field activities, such as water sampling, shall be segregated from visibly soiled PPE, double-bagged and disposed of offsite as municipal waste. Disposable sampling equipment from activities such as soil, sediment, and sludge sampling includes plastic sheeting used as liner material in containment areas around drilling rigs and waste storage areas; disposable sampling equipment; and soiled decontamination equipment. Where investigation involves potentially hazardous waste, visibly soiled PPE and disposable sampling equipment may require decontamination. The Field Manager shall use best professional judgment to determine if decontamination is appropriate. If decontamination is performed, PPE and disposable sampling equipment generated in the decontamination process may be double-bagged and disposed of as non-hazardous waste. PPE and disposable sampling equipment generated on separate days may be commingled.

Decontamination fluids shall be stored in drums separate from other IDW. If practical, decontamination fluids generated from different sources should not be stored in the same drum. If decontamination fluids generated over several days or from different sources are stored in a single container, information regarding dates of generation and sources shall be recorded in the field notebook, on the drum label (Section 2.3.2), and in the drum inventory (Section 2.5).

Liquid and sediment portions of the equipment decontamination fluid in the containment unit used by the drilling or excavation field crew should be separated. The contents of this unit normally consist of turbid decontamination fluid above a layer of predominantly coarse-grained sediment. When the contents of the containment unit are to be stored in IDW containers, the Field Manager shall direct the placement of as much liquid into drums as possible and transfer the remaining solids into separate drums. Observations of the turbidity and sediment load of the liquid IDW should be noted in the field notebook, on the drum label (Section 2.3.2), and in attachments to the drum inventory (see Section 2.5). It is likely that decontamination fluids will contain minor amounts of suspended solids that will settle out of suspension to become sediment at the bottom of IDW storage drums. As noted above, it may be necessary to segregate the drummed water from sediment during subsequent IDW sampling or disposal activities.

2.3 DRUM HANDLING AND LABELING

Drum handling consists of those actions necessary to prepare an IDW drum for labeling. Drum labeling consists of those actions required to legibly and permanently identify the contents of an IDW drum. Specific handling, storage, and labeling requirements may differ with the Naval installation or oversight entity. Specific requirements should be determined at the planning stage and documented in the WMP. General requirements are provided in the following sections.

2.3.1 Drum Handling

The drums used for containing IDW shall be approved by the United States Department of Transportation (DOT, 49 CFR 172). The drums shall be made of steel or plastic, have a 55-gallon capacity, be completely painted or opaque, and have removable lids (i.e., 1A1 or 1A2). New steel drums are preferred over recycled drums. For short-term storage of liquid IDW prior to discharge, double-walled bulk steel or plastic storage tanks may be used. Consideration must be given to scheduling and cost-effectiveness of bulk storage, treatment, and discharge system versus longer-term drum storage.

For long-term IDW storage, the DOT-approved drums with removable lids are recommended. The integrity of the foam or rubber sealing ring located on the underside of some drum lids shall be verified prior to sealing drums containing IDW liquids. If the ring is only partially attached to the drum lid, or if a portion of the ring is missing, a drum lid with sealing ring that is in good condition must be used. At some facilities, drums containing liquid IDW will be required to be stored in protective overpacks.

To prepare IDW drums for labeling, the outer wall surfaces and drum lids shall be wiped clean of all material that may prevent legible and permanent labeling. If potentially contaminated material adheres to the outer surface of a drum, that material shall be wiped from the drum, and the paper towel or rag used to remove the material shall be segregated with visibly soiled PPE and disposable sampling equipment.

2.3.2 Drum Labeling

Proper labeling of IDW drums is essential to the success and cost-effectiveness of subsequent waste screening and disposal activities. Labels shall be permanent and descriptive to facilitate correlation of field analytical data with the contents of individual IDW drums.

2.3.2.1 Preprinted Labels

A preprinted drum label as required by the appropriate Naval installation and/or regulatory agency shall be completed. The label will be affixed to the outside of the drum (or overpack if required) with the label easily readable for inspections and inventory. Label requirements may vary based on the site.

The requested information shall be printed legibly on the drum labels in black, indelible ink. Instructions for entering the required drum-specific information for each label field are provided by the Naval installation.

Painted Labels

An alternative method for labeling drums, if acceptable for the project, is to paint label information directly on the outer surface of the drum. At a minimum, the information placed on the drum shall include the contract/delivery order number, a drum number, the source identification type and number, the type of IDW, the generation date(s), and the government point of contact and telephone number. The drum surface shall be dry and free of material that could prevent legible labeling. Label information shall be confined to the upper two-thirds of the total drum height. The printing on the drum shall be large enough to be easily legible. Yellow, white, or red paint markers (oil-based enamel paint) that are non-photodegradable are recommended to provide maximum durability and contrast with the drum surface.

2.3.2.2 Regulatory Marking and Labeling

Federal and State regulations may require specific labeling for IDW generated (i.e., RCRA, TSCA, NESHAPs). Pre-printed labels shall be used as appropriate and completed in accordance with the specific regulatory requirement. These requirements will be identified in the approved project plans. Once determined to be hazardous, weekly inspections must also be conducted to ensure that labels and markings are in good conditions and to ensure the integrity of containers.

In addition, prior to off-site transportation USDOT requirements for marking and labeling of regulated DOT materials must be complied with. These requirements will be identified in the approved project plans or otherwise coordinated with the Field Manager after the IDW has been characterized and off-site disposition is being planned. Note that personnel (i.e., contractors or subcontractors) who perform USDOT functions must be properly trained in accordance with 49 CFR 172, Subpart G.

2.4 DRUM STORAGE

Drum storage procedures shall be implemented to minimize potential human contact with the stored IDW and prevent extreme weathering of the stored drums. Waste accumulation areas will be pre-designated by NAVFAC NW prior to the start of site work. IDW drums should be placed on pallets. Good management practices should be used in storing drums which include: containers shall be in good condition and closed during storage; wastes must be compatible with containers; where liquids are stored, storage areas should have secondary containment; and spill or leaks should be removed as soon as possible. These good management practices are mandatory requirements where RCRA hazardous wastes are stored.

Waste accumulation areas shall be maintained as prescribed by local regulatory entities and the appropriate Naval installation. In general, drums of IDW shall be stored within the Area of Concern (AOC) so that the site can utilize RCRA regulatory flexibility (i.e., administrative requirements, such as 90-day storage, may not be triggered; and LDRs will not be triggered if IDW is placed back in AOC). If IDW is determined to be RCRA hazardous waste, then RCRA storage, transportation and disposal requirements must be met.

Drums shall be stored at identified waste accumulation areas. All IDW drums generated during field activities at a single AOC shall be placed together, in a secure, fenced onsite area to prevent access to the drums by unauthorized personnel. When a secure area is not available, drums shall be placed in an area of the site with the least volume of human traffic. Plastic sheeting (or individual drum covers) and yellow caution tape shall be placed around the stored drums. Drums from projects involving multiple AOCs should remain at the respective source areas where the IDW was generated. IDW should not be transferred offsite for storage elsewhere, except under rare circumstances, such as the lack of a secure storage area onsite.

Proper drum storage practices shall be implemented to minimize damage to the drums from weathering and possible exposure to humans or the environment. When possible, drums shall be stored in dry, shaded areas and covered with impervious plastic sheeting or tarpaulin material. Every effort shall be made to protect the preprinted drum labels from direct exposure to sunlight, which causes ink on the labels to fade. In addition, drums shall be stored in areas that are not prone to flooding. The impervious drum covers shall be appropriately secured to prevent dislodging by the wind. It may be possible to obtain impervious plastic covers designed to fit over individual drums; however, the labeling information shall be repeated on the outside of these opaque covers.

Drums in storage shall be placed with sufficient space between rows of drum pallets and shall not be stacked, such that authorized personnel may access all drums for inspection. Proper placement will also render subsequent IDW screening, sampling, and disposal more efficient. It is recommended that IDW drums be segregated in separate rows/areas by matrix (i.e., soil, liquid or PPE/other).

If repeated visits are made to the project site, the IDW drums shall be inspected to clear encroaching vegetation, check the condition and integrity of each drum, check and replace labels as necessary, and replace or restore protective covers.

2.5 DRUM INVENTORY

Accurate preparation of an IDW drum inventory is essential to all subsequent activities associated with IDW drum tracking and disposal. An inventory shall be prepared for each project in which IDW is generated, stored, and disposed of. Naval installations and local regulatory authorities may have specific requirements associated with waste inventory and these requirements should be included in the planning process and documented in the WP, FSP, and WMP.

The drum inventory information shall include 11 elements that identify drum contents and indicate their fate.

2.5.1 Navy Activity (Generator)/Site Name

Inventory data shall include the Navy activity and the site name where the IDW was generated (e.g., NASWI, NBK Bangor, etc.).

2.5.2 DO Number

Inventory data shall include the contract and delivery order number associated with each drum (e.g., 0089).

2.5.3 Drum Number

The drum number assigned to each drum shall be included in the inventory database.

2.5.4 Storage Location Prior to Disposal

The storage location of each drum prior to disposal shall be included in the inventory (e.g., Building 394 Battery Disassembly Area, or Adjacent to West end of Building 54).

2.5.5 Origin of Contents

The source identification of the contents of each IDW drum shall be specified in the inventory (e.g., soil boring number, monitoring well number, sediment sampling location, or the multiple sources for PPE- or rinse water-generating activities).

2.5.6 IDW Type

Inventory data shall include the type of IDW in each drum (e.g., soil, PPE, disposable sampling equipment, sludge, sediment, development water, steam cleaning water, decontamination rinse water).

2.5.7 Waste Volume

The amount of waste in each drum shall be specified in the inventory as a percentage of the total drum volume or an estimated percentage-filled level (e.g., 95% maximum for liquid IDW).

2.5.8 Recommended Analytical Methods and Test Results Compared with Applicable Regulatory Standards

The recommended EPA analytical methods that adequately characterize IDW contained in each drum will be summarized in a tabular format and attached to the quarterly IDW drum inventory report (see Attachment I-A-7-1). The methodology for sampling and characterizing IDW shall be specified in the appropriate project plans.

2.5.9 Recommended or Actual Disposition of IDW Drum Contents

The recommended means of IDW disposal for each drum shall be summarized in a tabular format (e.g., Offsite, Encapsulated Onsite, Treatment/Sewer, Offsite Incinerator) and attached to the quarterly IDW drum inventory report (see Attachment I-A-7-1). Additional narrative discussion of the rationale for the recommended disposal option shall be attached to the quarterly IDW drum inventory report as data become available.

2.5.10 Generation Date

Inventory data shall include the date IDW was placed in each drum. If a drum contains IDW-generated over more than one day, the start date for the period shall be specified in dd-month-yy format. This date is <u>not</u> to be confused with an RCRA hazardous waste accumulation date (40 CFR 262). The accumulation start date, if required for RCRA wastes, shall be included on the hazardous waste drum label (Section 2.3.2.2).

2.5.11 Expected Disposal Date

The expected date each drum is to be disposed of shall be specified as part of the inventory in month-yy format. This date is for informational purposes only for the Navy, and shall not be considered contractually binding.

2.5.12 Actual Disposal Date

The actual drum disposal date occurs at the time of onsite disposal, or acceptance by the offsite treatment or disposal facility. It shall only be entered in the drum inventory database when such a date is available in dd-month-yy format.

In order to provide information for all 11 of the inventory elements of the quarterly inventory report described above, the main source of information will be provided by RPMs, or their designees, and summarized in Attachment I-A-7-1.

The recommended analytical test methods and actual test results (compared to applicable regulatory standards) will be provided to the appropriate Navy groups, by the RPM, or their designees, when such data are available. Testing methods shall be documented in the associated project plans. Recommended disposal options or actual disposition of the IDW drum contents will also be provided by RPMs as data become available. The NAVFAC Northwest RPM will forward all IDW data to the appropriate Navy

authority as attachments to the quarterly IDW drum inventory report. This information constitutes the results of preparing and implementing an IDW screening, sampling, classification, and disposal program for each site.

3.0 DOCUMENTATION

The RPM or designee is responsible for completing and updating the site-specific IDW drum inventory spreadsheet and submitting it as needed. The RPM is also responsible for submitting backup documentation to the U.S. Navy Program Management Office (PMO) about the analytical methods recommended to adequately characterize the IDW in each drum (Section 2.5.8). In addition, actual site or drum sampling results shall be forwarded to the PMO, along with a comparison to the applicable regulatory standards, for inclusion as attachments to the quarterly IDW drum inventory. As necessary, the backup documentation to the quarterly IDW drum inventory report shall also include the recommended means for IDW disposal for each drum (Section 2.5.9). After disposal, the actual means and/or location of disposal shall be indicated in tabular format with supporting narrative.

Field Managers and designates are responsible for documenting all IDW-related field activities in the field notebook, including most elements of the IDW drum inventory spreadsheet. The correct methods for developing and maintaining a field notebook are presented in SOP III-D, *Logbooks*.

Upon receipt of analytical data from the investigation, the information will be forwarded to the appropriate Naval authority for comparison to regulatory waste criteria. The Navy will designate the IDW and disposal options will be assessed based on the waste designation, approved transport/disposal facilities, and schedule for disposal. Naval installations may have additional requirements for reviewing analytical data, characterizing waste materials, transporting and off-site disposal. The RPM shall coordinate with the Naval installation early in the planning process to ensure that these requirements are properly identified, incorporated into the approved project plans, as available, and implemented in the field.

The disposal of IDW must be approved by the Navy and, in some cases, pertinent regulatory agencies. The disposal must be documented.

4.0 REFERENCES

Department of Transportation (DOT), Hazardous Materials Transportation Regulations, 49 CFR Parts 171 – 179.

EPA. 1998. EPA530-F-98-026, Management of Remediation Waste Under RCRA

EPA. 1991. Management of Investigative-Derived Wastes During Site Inspections. U.S. Environmental Protection Agency/540/G-91/009. May.

EPA. 1992. Guide to Management of Investigative-Derived Wastes. Quick Reference Guide. U.S. Environmental Protection Agency: 9345.3-03FS. January.

5.0 ATTACHMENTS

Attachment IA71 Example Format – Quarterly IDW Drum Inventory Updates

Attachment I-A-7-1 Quarterly IDW Drum Inventory Updates

Navy Activity / Site Name (Generator Site)	DO Number (0bbb)	Drum Number (xxxx-AA-Dzzz)	Drum Storage Location	Origin of Contents (Source ID#)	IDW Type	Waste Volume (Fill level %)	Waste Generation Date (dd-mm-yy)	Expected Disposal Date (mm-yy)	Actual Disposal Date (dd-mm-yy)
NSC Pearl Harbor/ Landfill	0068	0068-LF-D001	NSC, Bldg 7	SB-1	Soil Cuttings	100	16-Dec-92	Dec-93	Na
		0068-LF-D002	NA	MW-1	Purge Water	75	20-Dec-92	Jul 93	26-Jul-93
				MW-2	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,				
				MW-3					
		0068-LF-D003	NA	MW-1	Decon Water	95	20-Dec-92	Jul-93	26-Jul-93
				MW-2					
				MW-3					
		0068-LF-D004	NSC, Bldg.16	SB-1	PPE	50	16-Dec-92	Oct-93	NA
				SB-2					
				SB-3					
				SB-4					
				MW-1					
				MW-2					
				MW-3					
NAVSTA Guam/ Drum Storage	0047	0047-DS-001	Hazmat Storage Area	SB-1	Soil Cuttings	100	18-Feb-93	Sep-93	NA
				SB-2					

NA = Not Applicable



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GENERAL FIELD OPERATION

1.0 PURPOSE

This standard operating procedure (SOP) defines the general field organization and the field structure of sample collection, sample identification, record keeping, field measurements, and data collection. These SOPs are used to ensure the activities used to document sampling and field operations provide standardized background information and identities.

2.0 PROCEDURES

2.1 MOBILIZATION/DEMOBILIZATION

The SM or designee ensures that all purchase requests have been reviewed and approved by the PM. Then, the SM and PM assemble the project team in order to review the scope of work, disseminate the project plans, and complete the field equipment checklist (provided as Attachment I-A-9-1). After review by the project team, if additional items are required, additional purchase requests are prepared and approved by the PM.

The SM and project team upon arrival at the site inspects all equipment. Packing slips, bills of lading, or other documentation received with the shipment are initialed and returned to the purchasing department and a copy placed into the field file. Quantities, types, and makes of items received are checked against the original purchase requests to validate the shipment. Prior to validation of the shipping receipt, equipment is inspected to ensure all components are present and that the equipment calibrates and is fully functional. Any equipment received that is not fully functional is returned immediately and the vendor contacted to arrange a replacement.

The SM provides copies of the appropriate SOPs to the project team prior to the start of field activities. The most current versions of the SOPs are brought to the field. Any revisions to the SOPs must be approved by the PM and recorded in the field logbook.

It is imperative that rental equipment be cleaned (decontaminated), packaged, and returned immediately following the completion of a task. If any problems occurred on site with any equipment, the problems should be noted in detail in the field logbook and the SM notified. The SM will forward this information to the purchasing department and the vendor.

2.2 SHIPPING

If it is possible and /or practical, equipment and supplies should be shipped directly to the field site. If sensitive field equipment is to be shipped to the site, care shall be taken to ensure the equipment is not damaged en route. All original packaging material should be retained for return shipment of the equipment. Additional packing material (e.g., bubble wrap, bubble bags) may be required to provide additional protection for the shipped items. Equipment should always be shipped in its original carrying case. Each piece being shipped must have an address label on the shipping container separate from the shipping air bill.

2.3 CHAIN OF COMMAND

Chain of command protocols are implemented by the PM. These protocols should be strictly followed while performing field tasks. All decisions concerning priorities, project team assignments, sampling procedures, equipment management, and task approach are made by the PM, the SM, or an approved appointee. The SM or an approved designee will conduct a daily meeting prior to the start of field activities to discuss individual responsibilities. The meeting will also address potential contaminants that may be encountered, safety items (such as use of heavy equipment or protection against noise), special sampling requirements, and site control(s) to be employed to prevent injuries or exposure.

2.4 SAMPLING ORGANIZATION

The SM ensures the sampling design, outlined in project plans, is followed during all phases of the sampling activities at the site. For each sampling activity, field personnel record the information required by the applicable SOPs in their logbooks and on the exhibits provided in the SOPs.

2.5 REVIEW

The PM, SM, and, on occasion, the QAO or an approved designee checks field logbooks, daily logs, and all other documents that result from field operations for completeness and accuracy. Any discrepancies on these documents are noted and returned to the originator for correction. The reviewer acknowledges that review comments have been incorporated into the document by signing and dating the applicable reviewed documents.

3.0 DOCUMENTATION

Project activities shall be recorded in the field logbooks. The logbooks shall be kept current for the daily activities including documentation of all samples collected and the information relevant to the sample collection. All project required field forms shall be completed within a timely manner upon completion of the field task. All required field forms and specific logbook notations should be detailed in the field sampling plan.

4.0 REFERENCES

None.

5.0 ATTACHMENTS

Attachment IA91 Field Equipment Checklist.

Attachment I-A-9-1 Field Equipment Checklist

<u>General</u>		7.	Duct tape
1.	Health and Safety Plan	8.	Strapping tape
2.	Site base map	9.	Paper towels
3.	Hand calculator	10.	Bubble pack, foam pellets, or shredded paper
4.	Brunton compass	11.	Vermiculite
5.	Personal clothing and equipment	12.	Cooler labels ("This Side Up,"
6.	Personal Protective Equipment (First Aid kit)		"Hazardous Material," "Fragile")
7.	Cell or radio telephone	13.	Federal Express/DHL labels
Environme	ental Monitoring Equipment		
1.	Shovels		
2.	Keys to well caps		
3.	pH meter (with calibrating solutions)		
4.	pH paper		
5.	Thermometer		
6.	Conductivity meter (with calibrating solution)		
7.	Organic vapor analyzer or photoionization detector with calibration gas		
8.	H2S, O2, combustible gas indicator		
9.	Draeger tubes		
Shipping S	Supplies		
1.	Sample preservatives (nitric, hydrochloric, sulfuric acid/sodium hydroxide)		
2.	Heavy-duty aluminum foil		
3.	Coolers		
4.	Ice packs		
5.	Large zipper locking plastic bags		
6.	Heavy-duty garbage bags		

Sampling 1	<u>Equipment</u>	26.	0.45-micron filters (prepackaged in holders)
1.	Tool box with assorted tools (pipe wrenches, screwdrivers, socket set	27.	Stainless steel bowls
	and driver, open and box end	28.	SW scoop
	wrenches, hacksaw, hammer, vice grips)	29.	Peristaltic pump/tubing
2.	Geologic hammer	30.	Sample tags
3.	Trowel	31.	SOPs, HAZWOPER training
4.	Stainless steel and/or Teflon spatula		certificates, MSDs, FSP, QAPP
5.	Hand auger	<u>Decontam</u>	ination Equipment
6.	Engineer's tape	1.	Non-phosphate laboratory-grade detergent
7.	Steel tape	2.	Selected high purity, contaminant
8.	Electric water level sounder		free solvents
9.	Petroleum Interface Probe	3.	Long-handled brushes
10.	Batteries	4.	Drop cloths (plastic sheeting)
11.	Bailers (Teflon, stainless steel,	5.	Trash container
12.	acrylic, PVC) Slug test water displacement tube	6.	Galvanized tubs or equivalent (e.g., baby pools)
13.	Vacuum hand pump	7.	Tap Water
14.	Electric vacuum pump	8.	Contaminant free distilled/deionized
15.	Displacement hand pump		water
16.	Mechanical pump (centrifugal, submersible, bladder)	9.	Metal/plastic container for storage and disposal of contaminated wash solutions
17.	Portable generator	10.	Pressurized sprayers, H ₂ O
18.	Gasoline for generator	11.	Pressurized sprayers, solvents
19.	Hose	12.	Aluminum foil
20.	Calibrated buckets	13.	Sample containers
21.	Stop watch	14.	Emergency eyewash bottle
22.	Orifice plate or equivalent flow meter	15.	Documentation Supplies
23.	Data logger and pressure	Document	ation Supplies
	transducers	1.	Weatherproof, bound field logbooks
24.	Strip chart recorders	2	with numbered pages
25.	Sample bottles	2.	Daily Drilling Report forms

3.	Field Borehole Log forms
4.	Monitoring Well Installation Log forms
5.	Well Development Data forms
6.	Groundwater Sampling Log forms
7.	Aquifer Test Data forms
8.	Sample Chain-of-Custody forms
9.	Custody seals
10.	Communication Record forms
11.	Documentation of Change forms
12.	Camera and film
13.	Paper
14.	Permanent/indelible ink pens
15.	Felt tip markers (indelible ink)
16.	Munsell Soil Color Charts



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MONITORING/SAMPLING LOCATION RECORDING

1.0 PURPOSE

This standard operating procedure (SOP) describes the guidelines for generating the descriptions and information to be recorded for each physical location where monitoring, or sampling is conducted.

2.0 PROCEDURES

2.1 SAMPLING LOCATION MARKING

Sampling locations are based on criteria presented in the SAP. Whenever possible, each sampling location will be marked by a wooden lathe stake, directly marking the surface with marking paint, or with surveyors flagging. Each should be labeled with the location identifier outlined in the SAP. This should be done during the site visit or as soon as is feasible during field activities. This is to give the utility locators a better idea of the specific area to be cleared. Having the locations marked will also assist the field crew gain a better perspective of the locations to be worked

2.2 PHOTOGRAPHIC DOCUMENTATION

Site photographs showing monitoring/sampling locations with respect to structures or the site in general are encouraged. At certain installations, photography must be approved by the Navy. Prior to commencing work, the Navy must be notified to determine if cameras are allowed at the installation. The Note that the Navy will likely inspect your camera and may purge/delete some pictures if they feel there is a security issue. When possible, a menu board included in the photograph can be used to give relative information regarding the project and location.

For each photograph, record the following information in the field logbook:

- Photo number
- Date and time of the photo
- Orientation of the photo (direction facing)
- Subject-a description of what is contained within the photo. Others may be using the photos that are unfamiliar with the site and locations.

A detailed description of field logbook entries can be found in SOP III-D, *Logbooks*.

2.3 MONITORING/SAMPLING LOCATION INFORMATION FORM

A Monitoring/Sampling Location Information form must be filled out to establish each new sampling location. This form must be provided to the Navy for inclusion into the NAVFAC NW NIRIS Database. Established locations should not be re-established unless new information (such as survey information) is recorded about a location. A location description may be provided about a sampling location. It should contain detailed information regarding the physical features surrounding the location, including relevant

site information (i.e., obvious contamination, measurements to physical features, topographical relief, etc.). This description may be a copy of the field logbook or notes on project plan maps. These descriptions shall be attached to the field form. The PM is responsible for insuring that the project personnel have and use consistent terminology and descriptions as established in the SAP. The reverse of the field form contains a brief discussion of the form and descriptions of the information requested on the front.

3.0 DOCUMENTATION

None.

4.0 REFERENCES

SOP III-D, Logbooks

5.0 ATTACHMENTS

Attachment IA101 Example Monitoring/Sampling Location Information Form

FORM 11-1A MONITORING/SAMPLING LOCATION SUMMARY									
Installation ID:	Establi	ishing Contra	act ID:			Prime Contractor N	ame:		
Site Name:			DO/CTO:			Establishing Phase:		Date Established:	
Survey Contractor:			Local System Description:						
Location Name		Location Type	Projection Specification			Coordinates Northing (feet) East		ing (feet)	Ground Elevation (feet msl)
							1		

Location Types

ACID	Acid Pit	DU Decision Unit DW Domestic well	OUTFALLOu		SWS nonspecific	Surface water body -	WLBM	Bedrock Monitoring Well
ADIT	Adit	D_RIG_W Drill Rig Fluid		-Water	•		WLE	Extraction well
AGT	Above ground tank	Container EC Electrode	Separator PARK		SWSD Water/Sed	Surface iment	WLEA	Alluvial Extraction Well
AIR	Air (not inside a	ECT Electrode EF System effluent		antation/park/fore	SWWP	Wipe	WLEB	Bedrock Extraction Well
•	- ambient conditions)	EF System effluent EVAP EVAPORATION	st		SYSTEM	Treatment system air or	WLHM	Hybrid Monitoring Well
AMB	Ambient drinking juifer monitoring well	POND	PC Pai	int chip	water	Treatment system all of	WLI	Injection well
		EXCV Excavation FAGT Former above	PIPE Pip	peline	T	Trench	WLIA	Alluvial Injection Well
vapor m	Ambient organic onitor	ground tank location	PUBW Pul	blic drinking	TAA	Temporary	WLIM	Interface Monitoring Well
•	Asbestos-Containing	FL Fuel line FLOOD Flood Plain	water well	· ·	accumulati		WLL	Leaching Well
Area	7 lobe oto 5 Containing	FLOOD Flood Plain	PUMP_STAT	N ing station	TAIL	Mine tailings pile		· ·
BAY	Bay	Flood Control Gate	RAIN_STATN	ling station	TK	Tank	WLM	Monitoring well
BF	Backfill	FLOOR Floor FLOOR_SCRP Floor	Rainfa	all station	TMPM	Temperature Monitoring	WLS	Sparge well
BH	Borehole/Soil boring	scrapings	REF Refere		Point	15.11porataro monitoring	WLSG	Soil gas probe/Well
	· ·	FW Faucet/Tap/Spigot	garden/yard		TP	Test Pit	WRP	Waste rock pile
BIN	Roll-off bin	GAGE Gaging station (not USGS)		stream very well	TRANS	Transformer	WSFI	Water system facility intake
BIOL animal)	Biological (plant or	GW Geoprobe well GWTH Groundwater Test	SBAG Soil ba		TUNNEL	Steam tunnel sampling	WT WW	Wetlands Waste water
BLDG	Building (includes	Hold		as Probe	location		****	radio rator
building materials	air and building	HA Hand auger HDPCH Hydropunch	SIDEW Side V		TWP	Temporary well point		
	,	HOLE Hole	SLAG Slag h SND BLST S		UGA	Geophysical anomaly		
BULK	Bulk sample	HP Holding pond/Lagoon	material pile		UNK	Unknown		
BURN	Burn pit	ID Indoors	SP Spring SPT Septic	g/Seep stank	USGS	USGS gauging station		
CB	Concrete boring	IMP Import material		r System	UST	Underground storage		
CENT	Location surveyed at	IN System influent IT Intertidal	-	nd surface	tank			
	er of a UST field	LAGOON Lagoon	STEAM_LN S STKP Stocks		UXO	UXO		
CLGP Point	Canal Level Gauging	LENTIC Freshwater, lentic	STRM_DRN	•	UXO_G	UXO grid		
CPT	Cana nanatramatar	LF Landfarm	Storm STRM MH S		UXO_P	UXO point		
	Cone penetrometer	LGV Landfill Gas Vent	manhole	Norm Grain	VAULT	Vault		
CY	Cryopile	LH Leachate (Landfill) LK Lake/pond/open		nd, sub-surface	VPB	Vertical profile boring		
	Decontamination pad	reservoir	SUBSLAB S SUBT Subtid		WALL	Wall		
DITCH	Channel/Ditch	LOTIC Freshwater, lotic LYS Lysimeter	SUMON S					
DP December 10 cm	Direct	MH Manhole/Catch basin	monument SUMP Sump		WEEP	Weep hole		
Push/Ge	•	MS Sediment e.g., Marine		apor extraction	WF facility	Waste water treatment		
DRN		Sediment NQ Quality Control sample	system		WL	Well		
	Drum/Container	ON Ocean, open water (not						
contents DRW	Brywell	bay) OTHER Other			WLAM	Alluvial Monitoring Well		
Recorde	er:						Date:	
Checker	r:						Date:	
							_	



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SAMPLE NAMING

1.0 PURPOSE

This standard operating procedure (SOP) describes the naming convention to be used for samples collected, analyzed, and reported for the U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) projects. Unique sample identifiers are used to facilitate tracking by laboratory and project personnel and for purposes of storing, sorting, and querying data in the NAVFAC NW NIRIS database.

2.0 PROCEDURES

The contractor is responsible for assigning a unique sample ID to every individual sample collected. The contractor may use his or her own designations as long as the sample ID does not already exist in the NIRIS database. The contractor must also clearly identify which samples are field duplicates. This applies to both historical and planned sampling events. The used sampling identification scheme shall be identified and outlined in the field sampling plan.

3.0 DOCUMENTATION

All sample collection information must be recorded within the field logbook. Each sample collected will be clearly associated with the sample location (installation, site, and well or sample point location), matrix type, sample type (i.e. environmental, field duplicate, equipment rinsate), collection date and time, sampling method, and sampling depth (if appropriate). Only data codes and location IDs associated with NIRIS and NAVFAC NW's electronic deliverables SOP (NAVFAC NW 2015) shall be used.

Any sample submitted for analysis shall be documented using a completed chain-of-custody (COC) form that must accompany the shipment and a copy retained for the project records.

Samples submitted to an EPA laboratory shall also include a completed EPA analysis request form. The COC/analytical request form must be used to track all sample IDs.

4.0 REFERENCES

NAVFAC NW. 2015. Navy Environmental Data Transfer, Version 5.0.

5.0 ATTACHMENTS

None.



Revised March 2015

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MONITORING WELL AND PIEZOMETER INSTALLATION

1.0 PURPOSE

The purpose of this standard operating procedure (SOP) is to outline the methods by which all U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) personnel and their contractors will conduct monitoring well and piezometer installation. This procedure establishes the protocols and necessary equipment for installation of groundwater monitoring wells and piezometers.

2.0 PROCEDURES

2.1 EQUIPMENT

The following is an equipment list:

- Drill rig capable of installing wells to the desired depth in the expected formation material and conditions
- Well casing and well screen
- Bentonite pellets
- Filter pack sand
- Bentonite Grout or Portland Type I or II cement and powdered bentonite for grouting
- Protective well casing with locking cap
- High-pressure steamer/cleaner
- Long-handled bristle brushes
- Wash/rinse tubs
- Appropriate decontamination supplies as specified in the SOP for decontamination procedures
- Location map
- Plastic bags (re-sealable)
- Self-adhesive labels
- Weighted tape measure
- Water level probe
- Deionized water
- Logbook

- Boring log sheets
- Well construction form
- Plastic sheeting
- Drums for containment of cuttings and decontamination and/or development water (if necessary)

2.2 DECONTAMINATION

Before drilling or well installation begins, all drilling and well installation material should be decontaminated according to the protocols in SOP III-I, *Equipment decontamination*. Drilling equipment should be decontaminated between well locations.

2.3 Instrument Calibration

Before going into the field, the sampler should verify that field instruments are operating properly. Calibration times and readings should be recorded in a notebook to be kept by the field sampler. Specific instructions for calibrating the instruments are provided in the respective SOPs.

2.4 DRILLING AND WELL INSTALLATION PROCEDURES

2.4.1 Drilling Technique

If soil sampling is required by project plans, all soil samples should be collected according to the subsurface soil sampling procedures. The hole should be logged according to the methods specified in the project plans.

Boreholes should be advanced via conventional continuous-flight hollow-stem auger, sonic, air rotary, or mud rotary drilling methods and a drill rig capable of completing the monitor well(s) to the depth(s) specified in the project plans. Before drilling begins, well locations should be numbered and staked. The necessary permits and utility clearances shall be obtained in accordance with permits and utility clearance procedures. The permits and clearances will conform to specific Naval installation procedures or SOP 1-A-6 for utility location procedures.

During the drilling operation, the cuttings from the boring shall be placed into 55-gallon drums or roll-off container as specified in the project plans. Disposal of cuttings should be in accordance with the project plans and follow the specific Naval installation procedures or SOP l-A-7 for investigation-derived waste (IDW) management procedures.

2.4.2 Well Bore Drilling Operations

The procedure for well bore drilling is as follows:

- Set up drilling rig at previously staked and borehole location cleared for utilities.
- Record location, date, time, and other pertinent information in the field logbook.
- Drill hole of appropriate size using the project specified drilling method.
- Collect split-spoon samples at the predetermined intervals, if appropriate, for sample description and/or chemical analysis as specified in the project plans.
- Complete the borehole to the depth specified in the project plans.

• Document any difficult drilling conditions and ensures taken in response to such conditions (such as the addition of clean water to control heave).

2.4.3 Well Design Specifications

The general specifications for wells are as follows:

<u>Boring Diameter</u>. The boring should be of sufficient diameter to permit at least 2 inches of annular space between the boring wall and all sides of the centered riser and screen. The boring diameter should be of sufficient size to allow for the accurate placement of the screen, riser, filter pack, seal, and grout.

Well Casing. The well riser should consist of new, flush-threaded, PVC or stainless steel. The well diameter and thickness should be specified in the project plans. The risers should extend approximately 2 feet above the ground surface, except in the case of flush-mount surface casings. The tops of all well casings should be fitted with plugs or caps in locking monuments and locking caps in non-locking monuments.

Well Screens. The screen length for each well should be specified in the project plans. Well screens should consist of new threaded pipe with factory-machine slots or wrapped screen with an inside diameter equal to or greater than that of the well casing. The slot size should be indicated in the project plans and designed to be compatible with aquifer and sand pack material. The schedule thickness of PVC screen should be the same as that of the well casing. All screen bottoms should be fitted with a cap or plug of the same composition as the screen and should be within 0.5 foot of the open part of the screen. Traps may be used.

2.4.4 Well Installation Procedure

The following procedure should be initiated within 12 hours of well bore completion for uncased holes or partially cased holes and within 48 hours for fully cased holes. Once installation has begun, if no unusual conditions are encountered, there should be no breaks in the installation procedure until the well has been completed and the drill casing has been removed.

The procedure for monitoring well installation is as described below.

- 1. Decontaminate all well materials according to the SOP for decontamination procedures. After decontamination, all personnel who handle the casing should put on a clean pair of rubber or surgical gloves.
- 2. Measure each section of casing and screen to nearest 0.10 foot.
- 3. Assemble screen and casing as it is lowered into the open boring or drill casing (augers, when auger drilling is used) the hollow-stem augers.
- 4. Lower screen and casing to about 6 inches above the bottom of the boring.
- 5. Record the level of top of casing and calculate the screened interval. Adjust screen interval by raising assembly to desired interval, if necessary, and add selected filter sand to raise the bottom of the boring.
- 6. Begin adding filter pack sand around the annulus of the screen and casing a few feet at a time while withdrawing the drill casing or augers. Repeated depth soundings should be taken to monitor the level of the sand.

- 7. Allow sufficient time for the filter sand to settle through the water column outside the casing before measuring the sand level.
- 8. Extend the filter pack sand to at least 2 to 5 feet above the top of the well screen.
- 9. After placing the sand filter pack, install a seal at least 3 to 5 feet thick of bentonite pellets or chips. Add the bentonite pellets or chips slowly through the drill casing to avoid bridging. The thickness of the completed bentonite seal should be measured before the pellets have been allowed to swell. The completed bentonite seal should be allowed to hydrate before proceeding with the grouting operations.
- 10. Grout the remaining annulus from the top of the bentonite seal to near the ground surface as measured after the drill casing has been removed. The grout should be tremied into the borehole until the annulus is completely filled. The base of the tremie pipe should be placed approximately 5 feet above the bentonite seal. Bentonite chips or pellets may be used to backfill the well borehole.
- 11. After the grout sets for 24 hours it should be checked for settlement. If necessary, additional grout should be added to top off the annulus. This procedure may not be an option in high traffic or unsecured areas.
- 12. The steel monument, concrete pad and bollards, if required, should be installed according to the specifications in this SOP. The protective casing and posts should be painted a highly visible color.
- 13. Optional: Personnel should affix to the outer steel protective casing of each well a permanent, noncorrosive tag that clearly identifies the well number, the client's name, or the adjusted top of casing elevation. In some states, a state well identification number must be affixed to the monument.

2.4.5 Well Installation Specifications

<u>Filter Pack.</u> The annular space around the well screen should be backfilled with clean, washed silica sand sized to perform as a filter between the formation material and the well screen. The filter pack should extend a minimum 3 feet above the screen and may be tremied into place. The final depth to the top of the filter pack should be measured directly with the use of a weighted tape measure or rod and not by volumetric calculation methods. The grain size of the filter pack should be shown on the well construction log. The filter pack must be selected based on the grain size distribution of the native formation, and should be specified in the project plans.

Bentonite Seal and Grout. A minimum 2-foot-thick bentonite pellet/chip seal should be placed in the annulus above the filter pack. The thickness of the seal may vary slightly based on site conditions. The thickness of the seal should be measured immediately after placement, without allowance for swelling. Bentonite Grout or cement grout should then be placed from the top of the bentonite seal to the ground surface. Bentonite grout is preferred because of potential investigation derived waste issues if too much cement grout is prepared and due to heat generated from cement grout. Bentonite grout shall be "high solids" and prepared in accordance with the manufacturer's instructions. Cement grout should consist of a mixture of Portland cement (ASTM C150) and clean water, with a ratio of no more than 7 gallons of clean water per bag of cement (1 cubic foot or 94 pounds). Additionally, 3 percent by weight of bentonite powder should be added if permitted by state regulations. The grout should be prepared in a rigid

aboveground container by first thoroughly mixing the cement with water, and then mixing in the bentonite powder. Grout mixtures should be placed, by pumping through a tremie pipe. The lower end of the tremie pipe should be kept within 5 feet of the top of the bentonite seal. Grout should be pumped through the tremie pipe until undiluted grout flows from the annular space at the ground surface. The tremie pipe should then be removed and more grout added to compensate for settling. After 24 hours, the drilling contractor should check the site for grout settlement and add more grout to fill any depression. This should be repeated until firm grout remains at the surface.

<u>Protection of Well.</u> Personnel should at all times during the progress of the work take precautions to prevent tampering with the wells or the entry of foreign material into them. Upon completion of a well, a suitable cap should be installed to prevent foreign material from entering the well. The wells should be enclosed in a protective steel casing. Steel casings should be, at a minimum, 6 inches in diameter and should be provided with locking caps and locks. All locks used at a site should be keyed alike. If the well is to be a stickup (i.e., an aboveground monument), as specified in the project plans, a 1/4-inch drainage hole should be drilled in the protective steel casing, centered approximately 1/8-inch above the internal mortar collar for drainage. The well designation should be painted on the protective casing with a brush or paint pen. Painting should be done prior to well development. If specified in the project plans, a concrete pad should be constructed around the protective casing at the final ground level elevation and sloping away from the well. The concrete pad should measure at least 2 by 2 feet, with a thickness of 6 to 8 inches. Three 3-inch-diameter or larger steel posts should be equally spaced around the well and embedded in separate concrete-filled holes just outside the concrete pad. The protective steel posts should extend approximately 1 foot above the well riser. Any well that is to be temporarily removed from service or left incomplete due to a delay in construction should be capped with a watertight cap and equipped with a "vandal-proof" cover, satisfying applicable state or local regulations or recommendations.

3.0 DOCUMENTATION

Observations and data acquired in the field during the drilling and installation of wells should be recorded to establish a permanent record. A boring log should be completed for each well bore.

Additional documentation of well construction in the field logbook will include the following:

- Top of Casing surveyed elevation to 0.01 feet relative to known benchmarks, control points, and coordinate systems as defined in the Survey Specifications of NAVFAC NW SOPs V5.0 (or more current)
- Date
- Time
- Personnel
- Weather
- Subcontractors
- Health and safety monitoring equipment and readings
- Description of well location and triangulation measurements from landmarks, or GPS readings.
- Quantity and composition of grout, seals, and filter pack actually used during construction

- Screen slot size (in inches), slot configuration, outside diameter, nominal inside diameter, schedule/thickness, composition, and manufacturer
- Coupling/joint design and composition
- Protective casing composition and nominal inside diameter
- Start and completion dates
- Discussion of all procedures and any problems encountered during drilling and well construction

In addition, the well installation details should be shown in a diagram drawn in the field logbook. Each well diagram should consist of the following (denoted in order of decreasing depth from the ground surface):

- Reference elevation for all depth measurements
- Project and site names
- Well number
- Date(s) of installation
- Depth at which the hole diameter changes (if appropriate)
- Depth of the static water level and date of measurement(s)
- Total depth of completed well
- Depth of any grouting or sealing
- Nominal hole diameter(s)
- Depth and type of well casing
- Description (to include length, internal diameter, slot size, and well screen material
- Any sealing off of water-bearing strata
- Static water level upon completion of the well and after development
- Drilling date(s)
- Other construction details of monitoring well including grain size of well filter pack material and location of all seals and casing joints

All entries in the field logbook should be printed in black ink and legible.

4.0 REFERENCES

SOP I-A-7, IDW Management

SOP III-I, Equipment Decontamination

5.0 ATTACHMENTS

None.



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MONITORING WELL DEVELOPMENT

1.0 PURPOSE

This section describes the standard operating procedures (SOP) for monitoring well development to be used by all U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) personnel and their contractors.

2.0 PROCEDURE

2.1 Introduction

Well development procedures are crucial in preparing a well for sampling. Development enhances the flow of groundwater from the formation into the well and grades the well filter pack to reduce the movement of fine (clay and silt) particles into the well. The reduction in groundwater sample turbidity achieved by development improves the representation of chemical analyses performed on groundwater samples.

The goal of well development is to restore the area adjacent to a well to its natural condition by correcting damage to the formation during the drilling process. Well development should accomplish the following tasks:

- Remove any filter cake or any drilling fluid within the borehole that affects formation permeability.
- Grade the well filter pack to reduce the intrusion of fine formation particles.

Well development should not be performed sooner than 24 hours after the completion of well installation to allow the annular seal to fully set up.

2.2 FACTORS AFFECTING MONITORING WELL DEVELOPMENT

2.2.1 Type of Geologic Materials

Different types of geologic materials are developed more effectively by using certain development methods. Where permeability is greater, water moves more easily into and out of the formation and development is accomplished more quickly. Highly stratified deposits are effectively developed by methods that concentrate on distinct portions of the formation. If development is performed unevenly, a ground-water sample will likely be more representative of the permeable zones. In uniform deposits, development methods that apply powerful surging forces over the entire screened interval will produce satisfactory results.

2.2.2 Design and Completion of the Well

Because the filter pack reduces the amount of energy reaching the borehole wall, it must be as thin as possible if the development procedures are to be effective in removing fine particulate material from the interface between the filter pack and natural formation. Conversely, the filter pack must be thick enough to ensure a good distribution of the filter-pack material during emplacement and allow effective grading during development. Generally, filter pack material must be at least 2 inches thick. Variances from state agencies may be required for filter pack materials of less than 2 inches thick.

The screen slot size must be appropriate for the geologic material and filter pack material in order for development to be effective. If the slot size is too large, the filter pack and native material will enter the well, causing settlement of overlying materials and sediment accumulation in the casing. If the slot size is too small, full development may not be possible and the well yield will be below the potential of the formation. Additionally, incomplete development coupled with a narrow slot size can lead to blockage of the screen openings.

2.2.3 Drilling Method

The drilling method influences development procedure. Typical problems associated with specific drilling methods include the following:

- If a mud rotary method is used, a mudcake builds up on the borehole wall and must be removed during the development process.
- If drilling fluid additives have been used, the development process must attempt to remove all fluids that have infiltrated into the native formation.
- If driven casing or hollow-stem auger methods have been used, the interface between the casing or auger flights and the natural formation may have been smeared with fine particulate matter that must be removed during the development process.
- If an air rotary method has been used in rock formations, fine particulate matter is likely to build up on the borehole walls and may plug pore spaces, bedding planes, and other permeable zones. These openings must be restored during the development process.

2.3 PREPARATION

In preparing for monitoring well development, development logs for any other monitoring wells in the vicinity should be reviewed to determine the general permeability of the water-bearing formation, the associated likely groundwater yield from the well and the appropriate development method.

Depth to groundwater and information from the well construction log should be used in calculating of the required quantity of water to be removed. The distance between the equilibrated water level and the bottom of screen is the saturated section. The saturated section (feet) multiplied by the unit well volume per foot (gallons/linear foot) equals the gallons required to remove one total well volume of water. The unit well volume is the sum of the casing volume and the filter-pack pore volume, both of which depend upon casing and borehole diameter and the porosity of the filter pack material. Well volume for wells can be calculated using Table I-C-2-1 and Table I-C-2-2.

Table I-C-2-1* Casing Volume

Casing Diameter (inches)	Volume (gallon/linear foot)
2	0.16
4	0.65
6	1.47

Table I-C-2-2*
Filter Pack Pore Volume

Casing Diameter (inches)	Borehole Diameter (inches)	Volume ^a (gallon/linear foot)
2	6	0.52
2	8	0.98
4	10	1.37
4	12	2.09
6	12	1.76

^{*} The above two volumes must be added together to obtain one unit well volume.

2.4 DECONTAMINATION

The purpose of decontamination of development equipment is to prevent cross-contamination between monitoring wells. A steam-cleaner, if available, should be used to decontaminate development equipment. The equipment should be cleaned away from the monitoring well in such a fashion that decontamination effluent can be containerized.

A triple rinse decontamination procedure is acceptable for equipment such as bailers if access to a steam cleaner is not possible. See SOP III-I, *Equipment Decontamination*.

2.5 WELL DEVELOPMENT MONITORING

Throughout the well development process, a development record should be maintained in the field logbook. A well development field form presented in Attachment 1 (or similar) may be filled out in addition to the field logbook. The record should include the following information:

General

Well name/number and location

^a Assumes a porosity of 40% for filter pack.

- Date, time, and weather conditions
- Names of personnel involved

Development volume

- Initial and final water level
- Casing total depth and diameter
- Borehole diameter
- Casing volume, filter pack pore volume, total well volume
- Volume of water to be evacuated
- Method and rate of removal
- Appearance of water before and after development

Monitoring data for each sample point

- Date, time, elapsed time
- Cumulative gallons removed, removal method, removal rate
- Temperature, pH, specific conductance, turbidity, dissolved oxygen, and redox potential

Part of the well development procedure should consist of acquisition and analysis of general water quality parameters at periodic intervals, considering the total quantity of water to be removed and the removal rate. Depending on site conditions, the parameters specific conductance, pH, temperature, dissolved oxygen, turbidity, and redox potential may be measured. At a minimum the temperature, pH and turbidity should be monitored. Parameter measurements should be collected on a periodic basis during development. At a minimum, these parameters should be measured after removal of each well volume. The cumulative water volume of removed, the clock time, and the time elapsed during development should be recorded and a flow rate should be calculated. Development should continue until turbidity stabilizes at or below 10 nephelometric units or at least three well volumes have been removed. If three successive parameter measurements show stable values (values within 10% of each other) and turbidity is low, well development may cease. If stabilization has not been attained, if turbidity remains high, or if the well does not readily yield water, development should continue for a reasonable time as determined in the project plans or by the Project Manager.

The discussion of well development in special situations such as low yield formations is described in Section 2.7.

2.6 METHODS OF MONITORING WELL DEVELOPMENT

The methods available for the development of monitoring wells have been inherited from production well practices. Methods include (1) mechanical surging with a heavy, non-disposable bailer (stainless steel or PVC) surge block or swab, and (2) surge pumping. Development methods using air or jetting of water into the well are discouraged because of the potential for affecting water quality. In some circumstances,

air or water jet development may be necessary and should be conducted under the supervision of a qualified hydrogeologist.

All development water must be containerized and appropriately labeled, unless it is permissible to discharge onsite. Development should generally utilize mechanical surging or surge pumping, followed by bailing or groundwater removal with a pump. More detailed descriptions of appropriate development methods are presented below.

2.6.1 Mechanical Surging and Bailing

For mechanical surging and bailing, a heavy bailer, surge block or swab is operated either manually or by a drill rig. The bailer, surge block, or swab should be of sufficient weight to free-fall through the water in the well and create a vigorous outward surge. The equipment lifting the tool must be strong enough to extract it rapidly. A bailer is then used to remove fine-grained sediment and groundwater from the well.

Methodologies:

- 1. Properly decontaminate all equipment entering well.
- 2. Record the static water level and the total well depth.
- 3. Lower the bailer, surge block or swab to top of the screened interval.
- 4. Operate in a pumping action with a typical stroke of approximately 3 feet.
- 5. Gradually work the surging downward through the screened interval during each cycle.
- 6. Surge for several minutes per cycle.
- 7. Remove surge block and attach bailer in its place.
- 8. Bail to remove fines loosened by surging until water appears clear.
- 9. Repeat the cycle of surging and bailing until turbidity is reduced and stabilization of water quality parameters occurs.
- 10. The surging should initially be gentle and the energy of the action should gradually increase during the development process.

The advantages (+) and disadvantages (-) of this method are listed below:

- + It reverses the direction of flow, reduces bridging between large particles; the inflow then moves the fine material into the well for withdrawal.
- + It affects the entire screened interval.
- + It effectively removes fines from the formation and the filter pack.
- It may cause upward movement of water in the filter pack that could disrupt the seal.
- Potential exists for damaging a screen with a tight-fitting surge block or with long surge strokes.

2.6.2 Surge Pumping

Methodologies:

- 1. Properly decontaminate all equipment entering well.
- 2. Record the static water level and the total well depth.
- 3. Lower a submersible pump or airlift pump without a check valve to a depth within 1 to 2 feet of the bottom of the screened section.
- 4. Start pumping and increase discharge rate causing rapid drawdown of water in the well.
- 5. Periodically stop and start pump, allowing the water in the drop pipe to fall back into the well and surge the formation (backwashing), thus loosening particulates.
- **6.** The pump intake should be moved up the screened interval in increments appropriate to the total screen length.
- 7. At each pump position, the well should be pumped, over-pumped, and backwashed alternately until satisfactory development has been attained as demonstrated by reduction in turbidity and stabilization of water quality parameters.

The advantages (+) and disadvantages (-) of this method are listed below:

- + Reversing the direction of flow reduces bridging between large particles, and the inflow then moves the fine material into the well for withdrawal.
- + It effectively removes fines from the formation and filter pack.
- The pump position or suction line must be changed to cover the entire screen length.
- Submersible pumps suitable to perform these operations may not be available for small diameter (2 inches or less) monitoring wells.
- It is not possible to remove sediment from the well unless particle size is small enough to move through pump.

For additional information on well development, consult the references included in Section 4.0 of this SOP.

2.7 SPECIAL SITUATIONS

2.7.1 Development of Low Yield Wells

Development procedures for monitoring wells in low-yield (<0.25 gpm) water-bearing zones are somewhat limited. Due to the low hydraulic conductivity of the materials, surging of water in and out of the well casing is difficult. Also, when the well is pumped, the entry rate of water is inadequate to remove fines from the well bore and the gravel pack. Additionally, the process may be lengthy because the well can be easily pumped dry and the water level will be very slow to recover.

The procedures for mechanical surging and bailing should be followed for low yield wells. During surging and bailing, wells in low yield formations should be drawn down to total depth twice if possible.

Development can be terminated, however, if the well does not exhibit 80% recovery after 2 hours have passed.

3.0 DOCUMENTATION

Well development information should be documented in field logbooks in accordance with SOP III-D, *Logbooks* using indelible ink. In addition, well development monitoring forms (Attachment I-C-2-1 or similar) may be filled out in addition to the field logbook documentation. Copies of this information should be sent to the Project Manager and to the project files.

4.0 REFERENCES

Driscoll, F.G. 1987. Ground Water and Wells. Published by Johnson Division, St. Paul, Minnesota.

USEPA. 1992. RCRA, Ground Water Monitoring Technical Enforcement Guidance Document. U.S. Environmental Protection Agency/530/R-93/001. November.

U.S. EPA Environmental Response Team. 1988. Response Engineering and Analytical Contract Standard Operating Procedures. U.S. EPA, Research Triangle Park, NC.

SOP III-I, Equipment Decontamination

SOP III-D, Logbooks

5.0 ATTACHMENTS

Attachment I-C-2-1 Well Development Record

Attachment 1-C-2-1 Well Development Record

		PR	ROJECT		WELL NO.	
		JO	OB NO.	SITE	PREPARED BY	
WELL DEV	ELOPMENT LOG					
METHOD OVERPUMPAGE	INITIAL WATE	ER LEVEL		REMARKS:		
BAILER	FINAL WATER	LEVEL				
SURGE BLOCK	CAPACITY OF (GALLONS/LIN		VOLUME BETWEE (GALLONS/LINEA (ASSUMING 40% P			
AIR LIFT	2" = 0.16		2" CASING AND 6"	HOLE - 0.52		
OTHER	4" = 0.65 6" = 1.47		2" CASING AND 8" 4" CASING AND 10 4" CASING AND 12	" HOLE = 1.37		
Hole Diameter $d_h = $	→ d _w ← GROUND SURFACE	WELL VOL	LUME CALCULATION	:		
Well Casing:	SURFACE		(4	$(D)^2$) ²	
Inside Diameter d _w ID =_		CASING	$6 \text{ VOLUME} = V_c = \Pi \left(\frac{a}{a} \right)$	$\left(\frac{\text{wID}}{2}\right)^2 \left(\text{TD - H}\right) = 3.14 \left(\frac{1}{2}\right)$	-) (<u> </u>	_
Outside Diameter $d_wOD = $	TD			[(,)2 (, op	\27	
Depth to Base of	— k k -	FILTER	PACK PORE VOLUMI	$E = V_f = \prod \left[\left(\frac{d_h}{2} \right)^2 - \left(\frac{d_w OD}{2} \right)^2 \right]$	$\left(TD - (S \text{ or } H *)(P) = \frac{1}{2}\right)$	
Seal: S =_	SCREENED			(* if S > H, use	S; if S < H, use H)	
Depth to Base of Well: TD =	d _n			$=3.14\left[\left(\frac{}{2}\right)^2-\left(\frac{}{2}\right)^2\right]$)=
Estimated Filter Pack Porosity: P =		TOTAL	WELL VOLUME = VT	$= Vc + Vf = \underline{\qquad} + \underline{\qquad}$		
			-1		1	
DEVELOPM	ENT LOG:	CUMULATIVE WATER REMOVED		WATER QUALITY	C	OMMENTS
DATE TIME METH BEGIN/END	HOD ELAPSED FLOW TIME RATE (gpm)	GALLONS		CONDUC- D.O.* RED TIVITY	OOX TURBID- ITY	
	ll l		II .		11	

^{* =} Dissolved Oxygen



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LOW-FLOW GROUNDWATER PURGING AND SAMPLING

1.0 PURPOSE

This standard operating procedure (SOP) describes the conventional monitoring well sampling procedures to be used by all U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) personnel and contractors. Conventional monitoring well sampling procedures are provided in SOP I-C-4, *Groundwater Sampling from Temporary Wells (Piezometers)*.

2.0 PROCEDURE

2.1 PURPOSE

This procedure establishes the method for sampling groundwater monitoring wells for water-borne contaminants and general groundwater chemistry. The objective is to obtain groundwater samples with as little alteration of water chemistry as possible.

2.2 PREPARATION

2.2.1 Site Background Information

A thorough understanding of the purposes of the sampling event should be established prior to commencing field activities. A review of available data obtained from the site and pertinent to the water sampling should also be conducted. Copies of well logs or summary tables regarding well construction information should be available on-site if possible.

Previous groundwater development and sampling logs give a good indication of well purging rates and the types of problems that may be encountered during sampling, such as excessive turbidity and low well yield. They may also indicate where dedicated pumps are placed in the water column.

It is highly recommended that the field sampling team is familiar with the U.S. EPA recommended protocols for low-flow sampling outlined in the April 1996 Ground Water Issue *Low-Flow (Minimal Drawdown) Groundwater Sampling Procedures* (U.S. EPA 1996).

2.2.2 Groundwater Analysis Selection

The requisite field and laboratory analyses should be established prior to performing water sampling. The types and numbers of quality assurance/quality control (QA/QC) samples to be collected (refer to SOP III-B, *Field QC Samples (Water, Soil)*) should be specified in the QA plan developed for the site.

2.3 GROUNDWATER SAMPLING PROCEDURES

Groundwater sampling procedures at a site should include: (1) measurement of depth to groundwater and total depth, (2) assessment of the presence or absence of an immiscible phase (if required by the project plan), (3) assessment of purge parameter stabilization, (4) purging of static water within the well and well bore, and (5) obtaining a groundwater sample. Each step is discussed in sequence below. Depending

upon specific field conditions, additional steps may be necessary. As a rule, at least 24 hours should separate well development and well sampling events.

2.3.1 Measurement of Static Water Level Elevation

The depth to water and the total depth of the well should be measured to the nearest 0.01 foot to provide baseline hydrologic data, to calculate the volume of water in the well, and to provide information on the integrity of the well (e.g., identification of siltation problems). Dependent upon individual project requirements, synoptic water level collection may be required prior to groundwater sampling activities. In the event that synoptic water levels **are not** collected prior to sampling activities, total depth measurements should be collected **after** purging and sampling activities to prevent the suspension of fine-grained sediment that may be present at the bottom of the well. Each well should be marked with a permanent, easily identified reference point for water level measurements whose location and elevation have been surveyed.

An electronic water level meter accurate to 0.01 foot should be used to measure the water level surface and depth of the well. The presence of light, non-aqueous phase liquids (LNAPLs) and/or dense, non-aqueous phase liquids (DNAPLs) in a well requires measurement of the elevation of the top and the bottom of the product, generally using an interface probe. Water levels in such wells must then be corrected for density effects to accurately determine the elevation of the water table.

2.3.2 Decontamination of Equipment

Each piece of non-dedicated equipment should be decontaminated prior to entering the well. Decontamination should also be conducted prior to the start of sampling at a site, even if the equipment is known to be decontaminated subsequent to its last usage. This precaution is taken to minimize the potential for cross-contamination. In addition, each piece of equipment used at the site should be decontaminated prior to leaving the site. Dedicated sampling equipment need only be decontaminated prior to installation within the well. Clean sampling equipment should not be placed directly on the ground or other contaminated surfaces prior to insertion into the well. Dedicated sampling equipment that has been certified by the manufacturer as being decontaminated can be placed in the well without onsite decontamination.

Further details are presented in SOP III-I, Equipment Decontamination.

2.3.3 Detection of Immiscible Phase Layers

Unless specified in the project plans, groundwater samples should not be collected from wells with detectable amounts of LNAPL and DNAPL.

2.3.4 Purging Equipment and Use

To help minimize the potential for cross-contamination, well sampling should proceed from the least contaminated to the most contaminated. This order may be changed in the field if conditions warrant, particularly if dedicated sampling equipment is used. If decontamination of tubing is required by the project, Teflon[®] tubing is recommended. All groundwater removed from potentially contaminated wells should be handled in accordance with the investigation-derived waste (IDW) handling procedures described in SOP I-A-7, *IDW Management*.

Purging should be accomplished by removing groundwater from the well at low flow rates using a pump. According to the U.S. EPA (1996), the rate at which groundwater is removed from the well during purging ideally should be between than 0.1 to 0.5 L/min. The pump intake should be placed in the middle

of the calculated saturated screened interval. The purge rate should be low enough that substantial drawdown (>0.3 foot) in the well does not occur during purging. If a stabilized drawdown in the well can't be achieved and the water level is approaching the top of the screened interval, reduce the flow rate or turn the pump off (for 15 minutes) and allow for recovery. It should be noted whether or not the pump has a check valve. A check valve is required if the pump is shut off. *Under no circumstances should the well be pumped dry or otherwise over-purged*. Begin pumping at a lower flow rate, if the water draws down to the top of the screened interval again turn pump off and allow for recovery. If two tubing volumes (including the volume of water in the pump and flow cell) have been removed during purging then sampling can proceed next time the pump is turned on. This information should be noted in the field notebook or groundwater sampling log with a recommendation for a different purging and sampling procedure (USEPA, 2012).

Water level measurements should be collected to assess the water level effects of purging. A low purge rate also will reduce the possibility of stripping VOCs from the water, and will reduce the likelihood of mobilizing colloids in the subsurface that are immobile under natural flow conditions.

Water quality parameters should be collected and recorded on a regular basis (every 3-5 minutes) during well evacuation. Field parameters to be collected may include temperature, pH, specific conductance, salinity, dissolved oxygen, Redox potential, and turbidity. At least seven readings should be taken during the purging process unless the field parameters stabilize more quickly. These parameters are measured to demonstrate that the formation water, not stale well casing water, is being evacuated. Purging should be considered complete when the high and low values between three consecutive field parameter measurements stabilize within 10%. Turbidity may be considered stable if values are less than 10 nephelometric turbidity units (NTUs). The criterion for temperature may not be applicable if a submersible pump is used during purging due to the heating of the water by the pump motor. Field personnel should refer to the project-specific Sampling and Analysis Plan (SAP) for specific measurement requirements and well stabilization criteria.

All information obtained during the purging and sampling process should be entered into the field logbook. In addition to the field logbook, the data may be logged on a groundwater sampling log (Figure I-C-5-1 or equivalent). In special situations where LNAPL has been detected in the monitoring well and a groundwater sample is determined to be necessary by the Project Manager, a stilling tube should be inserted into the well prior to well purging. The stilling tube should be composed of a material that meets the performance guidelines for sampling devices. The stilling tube should be inserted into the well to a depth that allows groundwater from the screened interval to be purged and sampled. The bottom of the tube should be set below the upper portion of the screened interval where the LNAPL is entering the well screen. The goal is to sample the aqueous phase (groundwater) while preventing the LNAPL from entering the sampling device. To achieve this goal, the stilling tube must be inserted into the well in a manner that prevents the LNAPL from entering the stilling tube.

One method of doing this is to cover the end of the stilling tube with a membrane or material that will be ruptured by the weight of the pump. A piece of aluminum foil can be placed over the end of the stilling tube. The stilling tube is lowered slowly into the well to the appropriate depth and then attached firmly to the top of the well casing. When the pump is inserted, the weight of the pump breaks the foil covering the end of the tube, and the well can be purged and sampled from below the LNAPL layer. The membrane or material that is used to cover the end of the stilling tube must be fastened firmly so that it remains attached to the stilling tube when ruptured. Moreover, the membrane or material must retain its integrity after it is ruptured. Pieces of the membrane or material must not fall off of the stilling tube into the well. Although aluminum foil is mentioned in this discussion as an example of a material that can be used to

cover the end of the tube, a more chemically inert material may be required, based on the site-specific situation. Stilling tubes should be thoroughly decontaminated prior to each use. Groundwater removed during purging should be collected and stored onsite until its disposition is determined based upon laboratory analytical results. Storage should be in secured containers such as DOT-approved drums. Containers of purge water should be labeled with NAVFAC NW approved labels or paint pens.

2.3.5 Groundwater Sampling Methodology

The well should be sampled when groundwater within it is representative of aquifer conditions and after it has recovered sufficiently to provide enough volume for the groundwater sampling parameters. A period of no more than 2 hours should elapse between purging and sampling to prevent groundwater interaction with the casing and atmosphere. This may not be possible with a slowly recharging well. The water level should be measured and recorded prior to sampling to demonstrate the degree of recovery of the well. Sampling equipment should never be dropped into the well, because this could cause aeration of the water upon impact. In addition, the sampling methodology utilized should allow for the collection of a groundwater sample in as undisturbed a condition as possible, minimizing the potential for volatilization or aeration. This includes minimizing agitation and aeration during transfer to sample containers.

2.3.6 Sample Handling and Preservation

Many of the chemical constituents and physiochemical parameters to be measured or evaluated during groundwater monitoring programs are chemically unstable; therefore, samples must be preserved. The U.S. Environmental Protection Agency document entitled *Test Methods for Evaluating Solid Waste – Physical/Chemical Methods (SW-846)* (U.S. EPA 1995), includes a discussion of appropriate sample preservation procedures. In addition, SW-846 specifies the sample containers that should be used for each constituent or common set of parameters. In general, check with specific laboratory requirements prior to obtaining field samples. In many cases, the laboratory will supply the necessary sample bottles and required preservatives. In some cases, the field team may add preservatives in the field.

Improper sample handling may alter the analytical results of the sample. Samples should be transferred in the field from the sampling equipment directly into the container that has been prepared specifically for that analysis or set of compatible parameters as described in the Quality Assurance Project Plan.

When sampling for VOCs, water samples should be collected in vials or containers specifically designed to prevent loss of VOCs from the sample. An analytical laboratory should provide these vials, preferably by the laboratory that will perform the analysis. Groundwater from the sampling device should be collected in vials by allowing the groundwater to slowly flow along the sides of the vial. Sampling equipment should not touch the interior of the vial. The vial should be filled above the top of the vial to form a positive meniscus with no overflow. No headspace should be present in the sample container once the container has been capped. The sample can be checked for headspace by inverting the sample bottle and tapping the side of the vial to dislodge air bubbles. Sometimes it is not possible to collect a sample without air bubbles, particularly water that is aerated or naturally carbonated. In these cases, the investigator should note the problem to account for possible error. Field logs and laboratory analysis reports should note any headspace in the sample container(s) at the time of receipt by the laboratory, as well as at the time the sample was first transferred to the sample container at the wellhead.

2.3.6.1 Special Handling Considerations

Samples requiring analysis for organics should not be filtered. Samples should not be transferred from one container to another because this could cause aeration or a loss of organic material onto the walls of the container.

Groundwater samples to be analyzed for total and dissolved metals should be obtained sequentially. The sample to be analyzed for total metals, should be obtained directly from the pump and be unfiltered. The second sample should be filtered through a 0.45-micron membrane in-line filter and transferred to a container to be analyzed for dissolved metals. Allow at least 500 ml of effluent to flow through the filter prior to sampling. Any difference in concentration between the total and dissolved fractions may be attributed to the original metallic ion content of the particles and adsorption of ions onto the particles.

2.3.6.2 Field Sampling Preservation

Samples should be preserved immediately upon collection. Ideally, sample jars contain preservatives of known concentration and volume during the initial filling of the jar to a predetermined final sample volume. For example, metals require storage in aqueous media at pH of 2 or less. Typically, 0.5 ml of 1:1 nitric acid added to 500 ml of groundwater will produce a pH less than 2.0. Certain matrices that have alkaline pH (greater than 7) may require more preservative than is typically required. An early assessment of preservation techniques, such as the use of pH strips after initial preservation, may therefore be appropriate. It should be noted that introduction of preservatives will dilute samples, and may require normalization of results. Guidance for the preservation of environmental samples can be found in the EPA "Handbook for Sampling and Sample Preservation of Water and Wastewater:" (U.S. EPA 1982).

3.0 DOCUMENTATION

Information collected during groundwater sampling should be documented in the field logbook in accordance with SOP III-D, *Logbooks*. In addition, groundwater sampling purge logs may be (Figure I-C-5-1 or equivalent) may be filled out in addition to the field logbook. Copies of this information should be sent to the Project Manager and to the project files.

A groundwater sampling log should be documented in the field logbook and contain the following information:

- Identification of well
- Well depth
- Static water level depth
- Presence of immiscible layers
- Purge volume and pumping rate
- Time that the well was purged
- Collection method for immiscible layers
- Sample IDs
- Well evacuation procedure/equipment
- Date and time of collection
- Parameters requested for analysis
- Field analysis data

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- Field observations on sampling event
- Name of collector

Figure 1-C-5-1 Groundwater Sampling Log

Project Number:			Date:			
Location:			Time:	Time:		
Well Number:			Climatic Condi	tions:		
Initial Measurements	s: Static Wat Total Dept					
Well Purging:	Volume of Linear fee	Saturated Zone: f Water to be Evacuate t of Saturation x Casing Removal: Rate:			gals./line gallons	ar ft. x
Well Purge Data: DATE/ TIME	GALLONS REMOVED	рН	SP. COND.	D.O.	REDOX	TURBIDITY
Sample Withdrawal						
Appearance of Samp	ole:	Turbidity				
Laboratory Analysis	Parameters and Pre	servatives:				
Number and Types of						
Sample ID(s):						
Decontamination Pro	ocedures:					
Notes:						
Sampled by:						
Samples delivered to						
Date/Time:						
Transporters		<u> </u>				

^{*} Capacity of casing (gallons/linear foot): 2"-0.16, 4"-0.65, 6"-1.47, 8"-2.61, 10"-4.08, 12"-5.87

4.0 REFERENCES

SOP I-A-7, IDW Management

SOP *-C-4, Groundwater Sampling from Temporary Wells (Piezometers)

SOP III-I, Equipment Decontamination

SOP III-B, Field QC Samples

SOP III-D, Logbooks

- U.S. EPA. 1982. Handbook for Sampling and Sample Preservation of Water and Wastewater. EPA-600/4-82-029. September 1982.
- U.S. EPA. 1986. RCRA Ground-Water Monitoring Technical Enforcement Guidance Document.
- U.S. EPA. 1996. Ground Water Issue, Low-flow (Minimal Drawdown) Groundwater Sampling Procedures. EPA/540/S-95/504. April 1996
- U.S. EPA. 1995 and as revised. Test Methods for Evaluating Solid Waste–Physical/Chemical Methods (SW-846). January 1995.
- U.S. EPA. 2012. Standard Operating Procedure Low-Stress (Low Flow) / Minimal Drawdown Ground-Water Sample Collection, USEPA, Region 9, Management and Technical Services Division, April 2012.

5.0 ATTACHMENTS

None.



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WATER LEVEL MEASUREMENTS

1.0 PURPOSE

The purpose of this standard operating procedure (SOP) is to establish standard protocols for all U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) field personnel for use in making water level measurements.

2.0 PROCEDURE

2.1 EQUIPMENT

Equipment and materials used during liquid level and well-depth measurements:

- Electronic water level indicator with cable marked with 0.01-foot increments
- Electronic oil-water interface probe
- Engineers measuring tape with 0.01-foot increments may be used for water and petroleum reactive pastes as an alternative to an oil-water interface probe
- Weighted steel tape with 0.01-foot increments and chalk may be used as an alternative to a water level indicator
- Decontamination equipment
- Weatherproof, bound field logbook with numbered pages (see SOP III-D, *Logbooks*)
- Health and safety equipment appropriate for site conditions
- Keys for locked well covers
- Wire cutters if well has a security tag
- Turkey baster or hand pump in case flush-mount manhole is filled with water
- Bolt cutters for cutting "frozen" or rusted locks. HWD-40 is used to lubricate a rusted lock, but extreme care should be taken to avoid possible contamination to the well and equipment.
- Extra locks to replace cut locks

2.2 PRELIMINARY STEPS

Follow these steps prior to disturbing the liquid level in the well:

1. Locate the well and, confirm its label (if marked), and verify its position relative to other site features on the site map. Gain access to the top of the well casing.

- 2. Locate the permanent reference mark at the top of the well casing. This reference point shall be scribed, notched, or otherwise noted on the top of the casing. If no such marks are present, measure depth relative to the top of the highest point of the well casing and note this fact in the field logbook. Determine from the records and record the elevation of the permanent reference point and record it in the logbook.
- 3. Record any observations and remarks regarding the characteristics and condition of the well, such as evidence of cracked casing or surface seals, security of the well (locked cap), evidence of tampering, missing well cap, surface water entering the well casing, etc.

2.3 OPERATION

Follow these steps when taking depth to liquid level measurements in well suspected to have NAPL present.

- 1. Sample the air in the wellhead for gross organic vapors if required.
- 2. If non-aqueous phase liquid (NAPL) contamination is suspected, use an oil-water interface probe to determine the existence and thickness of the NAPL.
- 3. Open interface probe housing, turn probe on, and test the alarm. Ground the probe, because the slight electric charge from the probe could set off an explosion of highly flammable vapors. Slowly lower the probe into the well until the alarm sounds. A continuous alarm indicates light non-aqueous phase liquid (LNAPL), while an intermittent alarm indicates water. If LNAPL is detected, record depth of the initial (first) alarm. Mark the spot by grasping the cable with the thumb and forefingers at the top of the casing. Determine the depth to liquid relative to the permanent reference point on the well casing. Withdraw cable sufficiently to record the depth from the scale on the interface probe cable.
- 4. Continue to slowly lower the probe until it passes into the water phase (intermittent alarm). Slowly retract the probe until the NAPL continuous alarm sounds and record that level in the same manner as described above.
- 5. Record the depth to NAPL and the depth to water readings independently in the logbook. The thickness of the LNAPL can be calculated by subtracting depth to LNAPL reading from depth to water measurement.
- 6. Continue to slowly lower the interface probe through the water column to check for the presence of dense non-aqueous phase liquid (DNAPL) if suspected.
- 7. Measure and record the depths of the DNAPL layer (if any) as described above.
- 8. Slowly raise the interface probe, recording the depth to each interface as the probe is withdrawn. If there is a discrepancy in depths, clean the probe sensor and recheck the depth measurements.
- 9. Always lower and raise the interface probe slowly to minimize mixing of media.
- 10. Always perform a NAPL check in wells installed in areas with suspected NAPL contamination. Always perform a NAPL check if headspace test reveals presence of volatiles. Always perform a NAPL check the first time depth to liquid is measured in a well. If a well has been measured previously, with no NAPLs present, and none of the preceding conditions are met, the NAPL check may be omitted.

11. Decontaminate interface probe as appropriate.

For wells where NAPL is not suspected to be present, an electronic water level indicator or steel tape can be used as described below:

- 1. Remove the water level indicator probe from the case, turn on the sounder, and test the battery and sensitivity scale by pushing the test button. Adjust the sensitivity scale until you can hear the alarm.
- 2. Slowly lower the probe and cable into the well, allowing the cable reel to unwind. Continue lowering the probe until the alarm sounds. Very slowly raise and lower the probe until the point is reached where the meter just beeps. Mark the spot by grasping the cable with thumb and forefingers at the top of the casing. Record the depth to water relative to the permanent reference point. If no mark is present, use the highest point on the casing as a reference point. Withdraw the cable and record the depth.
- 3. Alternately, use a steel tape with an attached weight if the aquifer gradients are lower than 0.05 ft./ft. Due to the possibility of adding unknown contaminants from chalk colorants, only white chalk is permitted as a level indicator.
- 4. Rub chalk onto the end (first 1 foot) of the steel tape and slowly lower the chalked end into the well until the weighted end is below the water surface. (A small splash can be heard when the weighted end hits the water surface.)
- 5. Mark the spot on the tape by grasping the tape with the thumb and forefingers at the top of the casing as described in the subsection (2) above. Record this spot on the tape in the logbook as the "HOLD". Ensure not to retract the tape from the well until after the depth measurement (HOLD) is recorded.
- 6. Remove the steel tape from the well. The chalk will be wet or absent where the tape was below the water surface. Locate, read, and record this length in the logbook as the "CUT". Subtract the "CUT" length from the "HOLD" length and record the difference in the logbook. This is the depth to water table.
- 7. Decontaminate water level indicator or steel tape as appropriate

2.4 PRECAUTIONS

- Depending on the device used, correction factors may be required for some measurements. For example, if the water level indicator has been shortened during its repair.
- Check instrument batteries prior to each use.
- Exercise care not to break the seals at the top of the electric water level indicator probe.
- It is important to note that when measuring total well depth (bottom of casing), using an interface probe or water level indicator, the increments of measure are ticked off from the alarm sensor on the probe. On some meters there is a portion of the probe that sticks out beyond the alarm sensor. This needs to be accounted for when reading the bottom of casing measurement (i.e., added onto the reading). A potential problem arises if it is unknown whether this has been done on previous readings or not.

3.0 DOCUMENTATION

This section describes the documentation necessary for depth to liquid and well-depth measurements. All information shall be recorded in the field logbook using indelible ink in accordance with SOP III-D, *Logbooks*. At a minimum, the following information must be recorded:

- Date
- Time
- Weather
- Field personnel
- Well location and label
- Well condition
- Monitoring equipment type and readings
- Depth to Liquid measurements obtained
- Any other observations

All entries in the field logbook must be printed in black ink and legible. The actual readings measured should be recorded directly in the logbook. If calculations are necessary to determine the depth to liquid or liquid elevation, they should be performed using direct readings documented in the logbook.

Water level measurements must also be submitted electronically using the appropriate Naval Electronic Data Deliverable (NEDD) format for loading into NIRIS as defined in the NAVFAC NW SOPs (V5.0 or more current).

4.0 REFERENCES

SOP III-D, Logbooks

Thornhill, Jerry T. 1989. "Accuracy of Depth to Groundwater Measurements." In *EPA Superfund Groundwater Issue*. EPA/504/4-89/002.

5.0 ATTACHMENTS

None.



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FIELD PARAMETER MEASUREMENTS

1.0 PURPOSE

This standard operating procedure (SOP) provides instructions for the calibration, use, and checking of instruments and equipment for field measurements.

2.0 PROCEDURES

2.1 WATER QUALITY MEASUREMENTS

All field water quality meters shall be calibrated daily following the manufacturers' specifications. Calibration shall be performed prior to using the instrument for collecting parameters. In addition, the meter's calibration should be checked at mid-day and the end of the day to determine if measurements have drifted from the original calibration numbers. These checks are not intended to be a recalibration of the instrument. All calibration and measurement data shall be recorded in the project logbook. Fluids used for calibration shall be changed at regular intervals to ensure its integrity. Since different fluids have different shelf lives and tolerances, manufacturers' specifications should be checked as appropriate.

Most multi-probe water quality meters utilize a flow-through cell. If the unit being used does not have a flow-through cell, a large enough vessel (i.e. polypropylene beaker) in which the probes will be submerged shall be used. The water to be measured will be pumped continuously through the beaker from the bottom, overflowing the top. The flow-through cells will usually allow for quicker stabilization of dissolved oxygen and oxidation-reduction potential readings.

Water shall be allowed to flow continuously through the cell or beaker with water quality measurements being collected at regular intervals, every three to five minutes, until stabilization of the parameters has occurred. A minimum number of seven sets of readings should be collected or as otherwise outlined in the field sampling plan. Stabilization is considered to have occurred when three consecutive readings meet the following guidelines:

рН	+ 0.2 Scientific Units
Specific Conductance	+ 3 % mS/cm
Turbidity	+ 10% or < 10 NTUs
Dissolved Oxygen	+ 10% mg/cm
Salinity	+ 10%
Oxidation-Reduction Potential	+ 10 mV
Temperature	+ 10% °C

In addition to recording the above listed parameters the following information shall also be documented: date, time of measurement, flow rates, purge volumes, total volume purged, and other relative information (i.e. odors, sheen, comments on turbidity, water color)

2.2 ORGANIC VAPORS

Various organic vapor monitors have differing requirements for equipment warm-up and operation. Ensure that all organic vapor monitors are calibrated and operated according to the manufacturer's specification.

For measuring vapors present in soils, expose the monitor to a sample of soil by collecting a sample in sealable plastic baggy and placing the probe tip into the closed bag. In cold weather, the soil may need to be warmed prior to testing.

For measuring breathing zone vapors, hold the probe tip in the area of the breathing zone while field activities are being conducted. Take representative measurements from each different work or sampling area.

For monitoring well head space, place the probe tip just inside of the monitoring well casing immediately after removing the cap.

All readings including calibration information shall be recorded in the field logbook.

3.0 DOCUMENTATION

Record all observations and analysis in the field logbook as defined in SOP III-D, *Logbooks*. If required by the SAP, also complete the Field Measurement Data Form.

Field measurements must also be submitted electronically using the appropriate Naval Electronic Data Deliverable (NEDD) format for loading into NIRIS as defined in the NAVFAC NW SOPs (V5.0 or more current).

4.0 REFERENCES

ASTM International. 2003. D6771-02 Standard Practice for Low-flow Purging and Sampling Wells and Devices Used for Groundwater Quality Investigations

SOP III-D, Logbooks

5.0 ATTACHMENTS

Attachment I-D-7-1 Example Field Measurement Data form

		FII	FC ELD MEA	ORM 11-3 SUREME		ГА			
Installation ID :		Contract ID :				Prime	e Contractor Name	<u>}</u> :	
Site Name:				DO/CTO:		hase:		Measured Date:	
Location Name	Sample	Name	Sample Matrix	Measured Time	Samplin	g Depth	Analyte Test Type	Field Result	Units of Measure
			-						
		An	alyte Test Ty	•		ure			
AIRVE Air Velocity - M_SE ALK Alkalinity - MG_L BAL_GAS % balance gas in BARO PRESS Barometric BG_NIMOC Background no CACRB Calcium Carbonate CASING VOLUME Casing CHI Methane - PCT CO Carbon Monoxide - CO2 Carbon Dioxide - P COLOR Calcon Dioxide	air space by portable gas of Pressure - IN_HG n-methane organic compose - MG_L Volume Removed - GAL PPM CT ITS MG_L by Colometrics - MG_L by Titration - MG_L	meter - PCT FI HX unds - PPM LE M M N N N N O C O C	B Ferric Iro D Flame Io B Hydroger EL Lower ex	plosive limit - P se - MG_L se, lon (MN2+) m Persulfate - I nethane VOCs IG_L MG_L PCT NU Reduction Pote or SU	on - PPM L CT - MG_L - PPM	PU RE SA SC SG SG ST SU TD TE TU VO	3_PRESS Soil gas pre VA Sulfate - MC AGE Stage eleva IL Sulfide - MC IS Total Dissoil MP Temperature RB Turbidity - MC DL_PURGED Actual	- GAL_MIN Free Product - GAL_XT nductivity - MS_CM essure - IN_H2O 3_L biton for surface water - FT S_L ved Solids - MG_L - DEG_C or DEG_F TU Volume Purged [Prior to Sa olume Removed - GAL	impling] - GAL
Recorder:							Date:		
Checker:							Date:		



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LAND SURVEYING

1.0 PURPOSE

This standard operating procedure (SOP) sets forth protocols for acquiring land surveying data to facilitate the location and mapping of geologic, hydrologic, geotechnical data, and analytical sampling points and to establish topographic control over project sites.

2.0 PROCEDURES

The procedures listed below shall be followed during land surveying conducted for NAVFAC Northwest.

- All surveying work shall be performed under the direct supervision of a land surveyor registered in the state or territory in which the work is being performed (i.e. a Professional Land Surveyor, PLS).
- Survey instruments shall be calibrated in accordance with the manufacturer's specifications regarding procedures and frequencies. At a minimum, instruments shall have been calibrated no more than 6 months prior to the start of the survey work.
- Standards for all survey work shall be in accordance with National Oceanic and Atmospheric Administration (NOAA) standards and at the minimum accuracy standards set forth below. The horizontal accuracy for location of all grid intersection and planimetric features shall be (±) 0.1 feet. The horizontal accuracy for boundary surveys shall be one in ten thousand feet (1:10,000). The vertical accuracy for ground surface elevations shall be (±) 0.1 feet. Benchmark elevation accuracy and elevation of other permanent features, including monitoring wellheads, shall be (±) 0.01 feet.
- Surveys shall be referenced to the local established coordinate systems and all elevations and benchmarks established shall be based on North American Vertical Datum of 1988.
- Surveyed points shall be referenced to Mean Sea Level (Mean Lower Low Water Level).
- Appropriate horizontal and vertical control points shall be jointly determined prior to the start of
 survey activities. If discrepancies in the survey (e.g., anomalous water level elevations) are
 observed, the surveyor may be required to verify the survey by comparison to a known survey
 mark. If necessary, a verification survey may be conducted by a qualified third party.
- All field notes, sketches and drawings shall clearly identify the horizontal and vertical control
 points by number designation, description, coordinates and elevations. All surveyed locations
 shall be mapped using a base map or other site mapping specified by the Project Manager.
- All surveys shall begin and end at the designated horizontal and vertical control points to determine the degree of accuracy of the surveys.

- Iron pins used to mark control points shall be made of reinforcement steel or an equivalent material and shall be 18 inches long with a minimum diameter of 5/8 inch. Pins shall be driven to a depth of 18 inches into the soil.
- Stakes used to mark survey lines and points shall be made from 3-foot lengths of 2-inch by 2-inch lumber and pointed at one end. They shall be clearly marked with brightly colored weatherproof flagging and paint.
- The point on a monitoring well casing that is surveyed shall be clearly marked by filing grooves into the casing on either side of the surveyed point.

3.0 DOCUMENTATION

Using generally accepted practices, field notes shall be recorded daily by the surveyor in paper or electronic format. The data shall be neat, legible and easily reproducible. Copies of the surveyor's field notes and calculation forms generated during the work shall be obtained and submitted to the Navy or designee.

Surveyor's field notes shall, at a minimum, clearly indicate:

- The date of the survey
- General weather conditions
- The name of the surveying firm
- The names and job titles of personnel performing the survey work
- Equipment used, including serial numbers
- Field book designations, including page numbers.

Drawings and calculations submitted by the surveyor shall be signed, sealed and certified by a land surveyor registered (PLS stamped) in the state or territory in which the work was done.

Dated records of land surveying equipment calibration shall be provided by the surveyor along with equipment serial numbers and calibration records.

4.0 REFERENCES

The detailed requirements in the Geographic Data, Survey Specifications subsection of the parent compendium (NAVFAC Northwest SOPs V5.0) also apply and are not repeated here in this field procedure. These should be consulted as part of any Land Surveying effort. In addition, NAVFAC Northwest Cadastral Team, Record of Survey or other requirements may apply to the project, an example of their requirements can be found with the Survey Specifications referenced above.

5.0 ATTACHMENTS

None.



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FIELD QC SAMPLES (WATER, SOIL, SEDIMENT, TISSUE)

1.0 PURPOSE

This standard operating procedure (SOP) describes the number and types of field Quality Control (QC) samples that will be collected during U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) site field work. Quality control samples are controlled samples introduced into the analysis stream, whose results are used to review data quality and to calculate the accuracy and precision of the chemical analysis program. The purpose of each type of QC sample collection is described in this procedure. Collection and analysis frequency for quality control samples vary by project and are found in the project QA plan. Note that project-specific or contract requirements may supersede the requirements presented in this SOP.

2.0 PROCEDURES

The equipment required for the collection of QC samples is identical to the equipment required for the collection of environmental samples.

Field QC checks may include submission of trip blank, equipment rinsate, field blank, duplicate, and reference samples to the laboratory. Suggested frequency and types of QC check samples are discussed in the following guidance documents: *RCRA Technical Enforcement Guidance Document*, Section 4.6.1 (EPA 1986); the use and frequency of these field QC samples should be incorporated as appropriate. Types of field QC samples are discussed in general below. The frequency at which field QC samples should be collected for each QC level is provided in Table III-B-1.

The use of performance evaluation (PE) samples is discussed in SOP III-H, *Performance Evaluation Sample Procedures*.

2.1 TRIP BLANK

One trip blank is prepared off site by the laboratory using ASTM Type I organic-free water and included in each shipping container with samples scheduled for analysis of VOCs, regardless of the environmental medium. Trip blanks are placed in sample coolers by the laboratory prior to transport to the site so that they accompany the samples throughout the sample collection/ handling/ transport process. Once prepared, trip blanks remain unopened throughout the transportation and storage processes and are analyzed along with the associated environmental samples. Trip blanks are analyzed for VOCs and reported as water samples, even though the associated environmental samples may be from a matrix such as soil, tissue, or product.

One set of two 40 milliliter vials will constitute a trip blank and will accompany each cooler containing samples to be analyzed for volatile organics (VOCs) by methods such as CLP VOCs, 8010/601, 8020/602, 8240/624, modified 8015 (only if purge and trap analysis is performed, e.g., for gasoline, not

for extraction and analysis for diesel fuel), and equivalent state-specific methods. Trip blanks will be analyzed for VOCs only (EPA 1987).

Trip banks are not typically analyzed in association with tissue samples and are therefore not required for tissue sampling programs.

Table III-B-1
Field QC Samples per Sampling Event

	Lev	rel C2	Lev	el D2	I	Level E2
Type of Sample	Metal	Organic	Metal	Organic	Metal	Organic
Trip blank	NA1	1/cooler	NA1	1/	NA ¹	1/cooler
(for volatiles only)				cooler		
Equipment rinsate ³	1/day	1/day	1/day	1/day	1/day	1/day
Field blank	1/de	contamination	n water sou	rce/event/for	all QC levels	and all analytes
Field duplicates ⁴	10%	10%	10%	10%	5%	5%

Background samples at least 1/sample media/sample event⁵

Notes:

Source: NFESC. 1999. Navy Installation and Restoration Chemical Data Quality Manual.

2.2 EQUIPMENT RINSATE SAMPLES

Equipment rinsate samples are collected by pumping organic-free, analyte-free water over and/or through the sampling equipment (such as a bailer, sampling pump, or mixing bowl) following its final decontamination rinse. This rinse water is collected into the sample containers directly or with the use of a funnel if necessary. The rinse water may be poured by use of an electric or hand submersible pump by tipping the jug of water upside down, or by use of a stopcock.

Equipment rinsate samples are collected daily for sampling equipment used repetitively to collect environmental samples. One equipment rinsate sample shall be collected per day per sampling technique utilized that day (NFESC 1999 and EPA 1986). At least one equipment rinsate sample is analyzed for

¹NA means not applicable.

²QC levels are discussed in Section 2.8, Quality Control (QC) Levels.

³Samples are collected daily; however, only samples from every other day are analyzed. Other samples are held and analyzed only if evidence of contamination exists.

⁴The duplicate must be taken from the same sample that will become the laboratory matrix/spike duplicate for organics or for the sample used as a duplicate in inorganic analysis.

⁵Sample event is defined from the time sampling personnel arrive at the site until they leave the site for more than a period of one week; the use of controlled-lot source water makes one sample per lot rather than per event an option.

each group of 20 samples of a similar matrix type and concentration. Equipment rinsate samples are preserved, handled, and analyzed in the same manner as all environmental samples. Analytical results of equipment rinsate samples are used to access equipment cleanliness and the effectiveness of the decontamination process.

When disposable or dedicated sampling equipment is utilized, only one equipment rinsate sample will be collected per equipment lot or project phase. Disposable and/or dedicated sampling equipment may include stainless steel bowls or trowels that will be used for collection of only one soil sample, disposable bailers for ground-water sampling, dedicated submersible pumps for ground-water sampling, or other such equipment. This disposable and/or dedicated sampling equipment is typically pre-cleaned and individually wrapped by the manufacturer prior to delivery to the site. In this case, the equipment rinsate sample is used to provide verification that contaminants are not being introduced to the samples via sampling equipment.

Sampling devices (e.g., gloved hands, dip nets, or traps) for collection of tissue samples are generally non-intrusive into the organisms collected, so equipment rinsate samples will not be collected as long as the devices have been properly cleaned following SOP III-I, *Equipment Decontamination*, and the devices appear clean.

2.3 FIELD BLANKS

Field blanks are generally prepared on site during the sampling event by pouring American Society for Testing and Materials (ASTM) Type I organic-free water into randomly selected sample containers. Commercially available distilled water may be a satisfactory substitute for the ASTM organic-free water depending specific project requirement. At least one field blank is analyzed for each group of 20 samples of a similar matrix type and concentration.

Field blanks, consisting of samples of the source water used as the final decontamination rinse water, will be analyzed to assess whether the wash or rinse water contained contaminants that may have been carried over into the site samples.

The final decontamination rinse water source, the field blank source water, and equipment rinsate source water should all be from the same purified water source. Tap water used for steam cleaning augers or used in the initial decontamination buckets need not be collected and analyzed as a field blank, because augers typically do not touch the actual samples and because the final decontamination rinse water should be from a purified source.

Field blanks are collected at a frequency of one per sampling event per each source of water for all levels of QC. A sampling event is considered to be from the time sampling personnel arrive at a site until they leave for more than a week. Field blanks will be analyzed for the same analyses as the samples collected during the period that the water sources are being used for decontamination. If the same lot of the water source is used, a field blank needs to be collected only once per lot.

2.4 FIELD DUPLICATE

At least one duplicate sample is analyzed from each group of 10 samples of a similar matrix type and concentration. Field duplicate samples should be collected from areas most likely to be contaminated and

are preserved, handled, and analyzed in the same manner as all environmental samples. Field duplicates have the same location identification, sampling date and time, and depth interval as the associated environmental sample, but are assigned a unique sample number that is associated with the environmental sample number by virtue of the identical timestamp and location information.

Field duplicates for groundwater and surface water samples will generally consist of replicates. Field duplicates for soil samples will consist primarily of collocates. Soil field duplicates that are to be analyzed for volatile constituents will consist only of collocates; no soil samples that are to be analyzed for volatiles will be replicated (i.e., homogenized or otherwise processed or split) in the field. A separate sample will be collected to provide duplicates for non-volatile analyses. The sample may be homogenized and split in the field to form an original and duplicate (replicate) sample, or an additional volume into a separate sample container may be collected to form a duplicate (collocate) sample. Alternatively, replicates may be formed by homogenization in the laboratory. Duplicates will be analyzed for the same analytical parameters as their associated original sample.

Field duplicates for biological tissue samples will consist of splits of the original sample. Twice the required volume of organisms for one sample will be collected and placed into one food-grade self-sealing bag. The sample will later be homogenized in the laboratory and split, producing an original and a replicate sample. Replicates will be analyzed for the same analytical parameters as their associated original samples.

2.5 REFERENCE SAMPLES

There are two types of background levels of chemicals:

- Naturally occurring levels, which are concentrations of chemicals present in the environment that have not been influenced by humans (e.g., iron, aluminum)
- Anthropogenic levels, which are concentrations of chemicals that are present in the environment due to human-made, non-site sources (e.g., industry, automobiles)

Reference samples are samples taken from media similar to site media, but that are collected outside the zone of contamination, usually offsite.

Reference samples will be collected for each medium sampled at a site. Site-specific conditions will dictate the number of reference samples necessary to characterize background concentrations of contaminants of concern. However, at least one reference sample from each medium will be collected during each sampling event at a site. The samples will be analyzed for all the analytes for which site samples of that medium are analyzed. Background analysis, especially for metals, should be performed to assess the typical naturally occurring levels.

At least one reference sample will be collected for each biological species collected at a site. It may be difficult to find a nearby offsite location similar enough to the project site that has the same biological species available for offsite reference sample collection. Therefore, reference sample locations may need to be more distant from the site than for soil or water offsite reference samples. Collection methods will be identical for site and reference samples.

State-specific procedures may be required to establish background conditions for the site. This SOP is not intended to address such procedures and they should be consulted as necessary.

2.6 TEMPERATURE BLANKS

Temperature blanks are used to measure cooler temperatures upon receipt of the coolers at the laboratory. One temperature blank will be prepared and submitted to the project laboratory with each cooler. The temperature blank will consist of a sample jar containing water, which will be packed in the cooler in the same manner as the rest of the samples and labeled "temperature blank."

2.7 LABORATORY QUALITY CONTROL SAMPLES

The analytical laboratory uses a series of QC samples specified in each standard analytical method and laboratory SOP to assess laboratory performance. Analyses of laboratory QC samples are performed for samples of similar matrix type and concentration and for each sample batch. The types of laboratory QC samples are matrix spike/matrix spike duplicates, laboratory control standards, laboratory duplicates, method blanks, and surrogates. In addition, there may be other project-specific technical QC requirements.

2.7.1 Matrix Spike/matrix Spike Duplicate

Matrix spike/matrix spike duplicates (MS/MSDs) are used to assess sample matrix interferences and analytical errors, as well as to measure the accuracy and precision of the analysis. For MS or MSD samples, known concentrations of analytes are added to the environmental samples; the samples are then processed through the entire analytical procedure and the recovery of the analytes is calculated. Results are expressed as percent recovery of the known spiked amount for matrix spikes and the relative percent difference (RPD) for MS/MSDs. The MS/MSDs will be collected and analyzed at a rate of 5 percent of the field samples for each matrix and analytical method or at least one for each analytical batch, whichever frequency is greater.

Generally, a specific sampling location is used to collect field QC samples; however, it may not be possible to collect MS/MSD samples for all analyses at the same sampling location because of a limited volume of available material. In those instances, MS/MSD samples designated for various analyses will be collected from different locations (for example a MS/MSD for metals is collected at location X and an MS/MSD for PCBs is collected at location Y). Additionally, samples designated for MS/MSD analyses will not be collected from locations with potentially high concentrations of target analytes that may mask the added spike compounds. MS/MSD samples have the same location identification, sampling time, depth interval, and sample number as the associated environmental sample.

2.8 QUALITY CONTROL (QC) LEVELS

NAVFAC NW QC Levels III, IV are defined in SOP I-A-8 and Data Validation Procedure SOPs II-A through II-O. Level IV QC is appropriate to use for laboratory analysis for sites where cleanup decisions will be based on risk assessment. Sites on or eligible for the National Priorities List (NPL) will also have laboratory analyses conducted at Level IV QC. The QC level selected for laboratory analyses for many sites, therefore, will be NAVFAC NW Level IV. Other QC levels may be appropriate for certain types of

samples or analyses; criteria for selection of the appropriate QC level for individual projects and field work activities are discussed in SOP I-A-8, *Data Validation Planning and Coordination*.

3.0 DOCUMENTATION

Records of the collection of field QC samples should be kept in the sample logbook by the methods discussed in SOPs III-E, *Record Keeping, Sample Labeling, and Chain-of-Custody* and III-D, *Logbooks*.

4.0 REFERENCES

EPA. 1987. Data Quality Objectives for Remedial Response Activities: Development Process

NFESC. 1999. Navy Installation and Restoration Chemical Data Quality Manual.

EPA. 1992. RCRA Technical Enforcement Guidance Document.

SOP III-I, Equipment Decontamination

SOP, III-D, Logbooks

5.0 ATTACHMENTS

None.



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LOGBOOKS

1.0 PURPOSE

This standard operating procedure (SOP) describes the activities and responsibilities of U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) personnel and/or their contractors pertaining to the identification, use, and control of logbooks and associated field data records. This SOP establishes a standard format for recording field observations and describes the methods for use and maintenance of field logbooks.

2.0 PROCEDURE

2.1 EQUIPMENT

- Waterproof hardbound field logbook (typically 4-inch by 7-inch to 8-inch by 10.5-inch) with numbered pages
- Waterproof/indelible marking pen
- Ruler/straight edge
- Clipboard

2.2 LOGBOOK MAINTENANCE

Prior to commencement of field work, logbooks will be assigned to field personnel by the Project Manager. If personnel changes must be made during a project, the successor may use the same logbook. In this case, the logbook cover page will indicate all persons who have made entries and the dates. This may be inappropriate if there are a large number of people involved.

The logbook user is responsible for recording pertinent data into the logbook to satisfy project requirements and for attesting to the accuracy of the entries by dated signature. The logbook user is also responsible for safeguard of the logbook while having custody of it.

Individuals performing specific tasks associated with a field project may keep a separate logbook; however, these logbooks must conform to this procedure and will become a permanent part of the central project file. The Project Manager is responsible for reviewing and signing all field logbooks associated with the project.

2.3 RECORDING FIELD ACTIVITIES

The field team provides a permanent record of daily activities, observations, and measurements through the use of a field logbook. All logbook entries will be made in indelible black or blue ink. No erasures are permitted. If an incorrect entry is made, the data will be crossed out with a single line and initialed and dated by the originator. Entries can be organized into easily understood tables if possible.

All logbook pages will be signed and dated at the bottom. Times will be recorded next to each entry. If a full page is not used during the course of a workday, a diagonal line will be drawn through the unused portion of the page and signed (in this case, it would not be necessary to sign the bottom of the page). If the project is completed and the logbook has not been completely filled, a diagonal line will be drawn across the first blank page after the last entry, and "no further entries" written before the page is signed and dated.

Daily entries will be made during field activities by, at a minimum, one field team member to provide daily records of all significant events, observations, and measurements during field operations. Notes will start at the beginning of the first blank page and extend through as many pages as necessary. All page numbers will be consecutively numbered as the logbook is filled.

The inside cover page of each logbook will contain the following information:

- Book number
- Project name
- Contract number
- Project number
- Navy Activity/Installation
- Site name
- Start date
- End date
- Person to whom the logbook is assigned
- Agency/Company name
- Agency/Company address
- Agency/Company phone number

The field logbook serves as the primary record of field activities. When possible, the field book should be dedicated to a singular Navy Activity/Installation to facilitate long-term records archiving. Entries shall be made chronologically and in sufficient detail to allow the writer or a knowledgeable reviewer to reconstruct the applicable events. Individual data forms may be generated to provide systematic data collection documentation. Entries on these forms shall meet the same requirements as entries in the logbook and shall be referenced in the applicable logbook entry. Individual data forms shall reference the applicable logbook and page number. At a minimum, names of all samples collected shall be included in the logbook even if recorded elsewhere.

All field descriptions and observations are entered into the logbook, as described in Attachment III-D-1.

Typical information to be entered includes, but is not limited to, the following:

- Date and time of all onsite activities
- Site location and description
- Weather conditions
- Field work documentation
- Descriptions of and rationale for approved deviations from the Work Plan or Field Sampling Plan
- Field instrumentation readings
- Personnel present
- Photograph references
- Sample locations
- Sample identifications, as described in SOP I-A-11, Sample Naming
- Field QC sample information
- Field descriptions, equipment used, and field activities accomplished to reconstruct field operations
- Meeting information
- Daily health and safety meeting notes
- Important times and dates of telephone conversations, correspondence, or deliverables
- Field calculations
- PPE level
- Calibration records
- Subcontractors present
- Equipment decontamination procedures and effectiveness
- Procedures used for containerization of investigative-derived waste

Logbook page numbers shall appear on each page to facilitate identification of photocopies.

If a person's initials are used for identification, or if uncommon acronyms are used, these should be identified on a page at the beginning of the logbook.

At least weekly and preferably daily, the preparer shall photocopy and retain the pages completed during that session for backup. This will prevent loss of a large amount of information if the logbook is lost.

A technical review of each logbook shall be performed by a knowledgeable individual such as the Project Manager.

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3.0 DOCUMENTATION

The field logbook shall be retained as a permanent project record. If a particular Task Order requires submittal of photocopies of logbooks, this shall be performed as required.

4.0 REFERENCES

SOP I-A-11, Sample Naming

5.0 ATTACHMENTS

Attachment III-D-1 Description of Logbook Entries

Attachment 1 Description of Logbook Entries

Logbook entries shall contain the following information, as applicable, for each activity recorded. Some of these details may be entered on data forms as described previously.

Name of Activity For example, Asbestos Bulk Sampling, Charcoal Canister Sampling, Aquifer

Testing.

Task Team Members and

Equipment

Name all members on the field team involved in the specified activity. List equipment used by serial number or other unique identification, including

calibration information.

Activity Location Indicate location of sampling area as specified in the Field Sampling Plan. Record

valid Navy Installation/Active and Site, at a minimum.

Weather Indicate general weather and precipitation conditions.

Level of Personal Protective Equipment The level of personal protective equipment (PPE), e.g., Level D, should be

recorded.

Methods Indicate method or procedure number employed for the activity.

Sample IDs Indicate the unique identifier associated with the physical samples. Identify QC

samples. Value can be numeric or alphanumeric and must not already exist in the

database.

Sample Type and Volume

Indicate the medium, container type, preservative, and the volume for each sample.

Sample Collection Information Indicate the location of sample, date and time of collection, sample matrix, sample depth interval, sample methods, sample handling, including filtration and preservation, analysis required and packaging and shipping information.

Time and Date Record the time and date when the activity was performed (e.g., 0830/08/OCT/89).

Use the 24-hour clock for recording the time and two digits for recording the day of

the month and the year.

Analyses Indicate the appropriate code for analyses to be performed on each sample, as

specified in the Field Sampling Plan.

Field Measurements Indicate measurements and field instrument readings taken during the activity.

Chain of Custody and Distribution

Indicate chain-of-custody for each sample collected and indicate to whom samples

are transferred and the destination.

References If appropriate, indicate references to other logs or forms, drawings or photographs

employed in the activity.

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Recorded by

Narrative (including time and location)

Create a factual, chronological record of the team's activities throughout the day, including the time and location of each activity. Include descriptions of any general problems encountered and their resolution. Provide the names and affiliations of non-field team personnel who visit the site, request changes in activity, impact to the work schedule, requested information, or observe team activities. Record any visual or other observations relevant to the activity, the contamination source, or the sample itself.

It should be emphasized that logbook entries are for recording data and chronologies of events. The logbook author must include observations and descriptive notations, taking care to be objective and recording no opinions or subjective comments unless appropriate.

Include the signature of the individual responsible for the entries contained in the

logbook and referenced forms.

Checked by Include the signature of the individual who performs the review of the completed

entries.



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RECORD KEEPING, SAMPLE LABELING, AND CHAIN-OF-CUSTODY PROCEDURES

1.0 PURPOSE

The purpose of this standard operating procedure (SOP) is to establish standard protocols for all U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) field personnel and their contractors for use in maintaining field and sampling activity records, writing sample logs, labeling samples, ensuring that proper sample custody procedures are utilized, and completing chain-of-custody/analytical request forms.

2.0 PROCEDURES

Standards for documenting field activities, labeling the samples, documenting sample custody, and completing chain-of-custody and analytical request forms are provided in this procedure. The standards presented in this section shall be followed to ensure that samples collected are maintained for their intended purpose and that the conditions encountered during field activities are documented.

2.1 RECORD KEEPING

The field logbook serves as the primary record of field activities. Entries shall be made chronologically and in sufficient detail to allow the writer or a knowledgeable reviewer to reconstruct each day's events. Field logs such as soil boring logs and ground-water sampling logs will also be used. These procedures are described in SOP III-D, *Logbooks*.

2.2 SAMPLE LABELING

A sample label with adhesive backing shall be affixed to each individual sample container. Clear tape shall be placed over each label (preferably prior to sampling) to prevent the labels from tearing off, falling off, or being smeared, and to prevent loss of information on the label. The following information shall be recorded with a waterproof marker on each label:

- Project name or number (optional)
- Sample ID
- Date and time of collection
- Sampler's initials
- Matrix (optional)
- Sample preservatives (if applicable)
- Analysis to be performed on sample. This shall be identified by the method number or name identified in the subcontract with the laboratory. For water samples, a separate container is

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typically used for each separate test method, whereas with soil samples, multiple analyses can be performed on the soil obtained from one sample container. In order to avoid lengthy lists on each container and confusion, soil sample containers may not list every analysis to be performed.

These labels may be obtained from the analytical laboratory or printed from a computer file onto adhesive labels. The adhesive glue used on the labels must be such that it does not contaminate the sample.

2.3 CUSTODY PROCEDURES

For samples intended for chemical analysis, sample custody procedures shall be followed through collection, transfer, analysis, and disposal to ensure that the integrity of the samples is maintained. Custody of samples shall be maintained in accordance with EPA chain-of-custody guidelines as prescribed in EPA's NEIC Policies and Procedures, National Enforcement Investigations Center, Denver, Colorado, revised May 1986; EPA RCRA Ground Water Monitoring Technical Enforcement Guidance Document (TEGD), Guidance for Conducting Remedial Investigations and Feasibility Studies Under CERCLA (EPA OSWER Directive 9355 3-01), Appendix 2 of the Technical Guidance Manual for Solid Waste Water Quality Assessment Test (SWAT) Proposals and Reports, and Test Methods for Evaluating Solid Waste (EPA SW-846). A description of sample custody procedures is provided below.

2.3.1 Sample Collection Custody Procedures

According to EPA's NEIC Policies and Procedures, a sample is considered to be in custody if:

- It is in one's actual physical possession or view
- It is in one's physical possession and has not been tampered with (i.e., it is under lock or official seal)
- It is retained in a secured area with restricted access
- It is placed in a container and secured with an official seal such that the sample cannot be reached without breaking the seal

Custody seals shall be placed on sample containers immediately after sample collection and on shipping coolers if the cooler is to be removed from the sampler's custody. Custody seals will be placed in such a manner that they must be broken to open the containers or coolers. The custody seals shall be labeled with the following information:

- Sampler's name or initials
- Date and time that the sample/cooler was sealed.

These seals are designed to enable detection of sample tampering. An example of a custody seal is shown in Attachment III-E-1.

Field personnel shall also log individual samples onto carbon copy chain-of-custody forms when a sample is collected. These forms may also serve as the request for analyses. Procedures for completing these forms are discussed in Section 2.4 indicating sample number, matrix, date and time of collection, number of containers, analytical methods to be performed on the sample, and preservatives added (if any). The samplers will also sign the COC form signifying that they were the personnel who collected the samples. The COC form shall accompany the samples from the field to the laboratory. When a cooler is ready for shipment to the analytical laboratory, the person delivering the samples for transport will sign and

indicate the date and time on the accompanying COC form. One copy of the COC form will be retained by the sampler and the remaining copies of the COC form shall be placed inside a self-sealing bag and taped to the inside of the cooler. Each cooler must be associated with a unique COC form. Whenever a transfer of custody takes place, both parties shall sign and date the accompanying carbon copy COC forms, and the individual relinquishing the samples shall retain a copy of each form. One exception is when the samples are shipped; the delivery service personnel will not sign or receive a copy because they do not open the coolers. The laboratory shall attach copies of the completed COC forms to the reports containing the results of the analytical tests. An example COC form is provided in Attachment III-E-2. An example of a completed COC form is provided in Attachment III-E-3 and described in Section 2.4.

2.3.2 Laboratory Custody Procedures

The following are custody procedures to be followed by an independent laboratory receiving samples for chemical analysis; the procedures in their Laboratory Quality Assurance Plan (LQAP) must follow these same procedures. A designated sample custodian shall take custody of all samples upon their arrival at the analytical laboratory. The custodian shall inspect all sample labels and COC forms to ensure that the information is consistent, and that each is properly completed. The custodian will also measure the temperature of the samples in the coolers upon arrival. The custodian shall also note the condition of the samples including:

- If the samples show signs of damage or tampering.
- If the containers are broken or leaking.
- If headspace is present in sample vials.
- Proper preservation of samples (made by pH measurement, except VOCs and purgeable TPH). The pH of these samples will be checked by the laboratory analyst, after the sample aliquot has been removed from the vial for analysis.
- If any sample holding times have been exceeded.

All of the above information shall be documented on a sample receipt sheet by the custodian.

Any discrepancy or improper preservation shall be noted by the laboratory as an out-of-control event and shall be documented on an out-of-control form with corrective action taken. The out-of-control form shall be signed and dated by the sample control custodian and any other persons responsible for corrective action. An example of an out-of-control form is included as Attachment III-E-4.

The custodian shall then assign a unique laboratory number to each sample and distribute the samples to secured storage areas maintained at 4°C. The unique laboratory number for each sample, contractor sample ID, client name, date and time received, analysis due date, and storage details shall also be manually logged onto a sample receipt record and later entered into the laboratory's computerized data management system. The custodian shall also sign the shipping bill and maintain a copy.

Laboratory personnel shall be responsible for the care and custody of samples from the time of their receipt at the laboratory through their exhaustion or disposal. Samples should be logged in and out on internal laboratory COC forms each time they are removed from storage for extraction or analysis.

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2.4 COMPLETING CHAIN-OF-CUSTODY/ANALYTICAL REQUEST FORMS

COC form/analytical request completion procedures are crucial in properly transferring the custody and responsibility of samples from field personnel to the laboratory. This form also is important for accurately and concisely requesting analyses for each sample; it is essentially a release order from the analysis subcontract.

Attachment III-E-2 is an example of a generic COC/analytical request form that may be used by field personnel. Multiple copies may be tailored to each project so that much of the information described below need not be handwritten each time. Attachment III-E-3 is an example of a completed site-specific COC/analytical request form, with box numbers identified and discussed in text below.

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Box 1 Project Manager: This name shall be the name that will appear on the report. Do not write the name of the Project Coordinator or point of contact for the project instead of the Project Manager.

Project Name: Write it, as it is to appear on the report.

Project Number: Write it as it is to appear on the report. It shall include the project number, task number, and general ledger section code. The laboratory subcontract number should also be included.

- Box 2 Bill to: List the name and address of the person/company to bill only if it is not in the subcontract with the laboratory.
- Box 3 Sample Disposal Instructions: These instructions will be stated in the Basic Ordering Agreement (BOA) or each Task Order statement of work with each laboratory.

Shipment Method: State the method of shipment, e.g., hand carry; air courier via FEDEX, AIRBORNE, DHL or equivalent.

Comment: This area shall be used by the field team to communicate observations, potential hazards, or limitations that may have occurred in the field or additional information regarding analysis. For example: a specific metals list, explanation of Mod 8015, Mod 8015 + Kerosene, samples expected to contain high analyte concentrations.

Box 4 Cooler Number: This will be written somewhere on the inside or outside of the cooler and shall be included on the COC. Some laboratories attach this number to the trip blank identification, which helps track VOC samples. If a number is not on the cooler, field personnel shall assign a number, write it on the cooler, and write it on the COC.

QC Level: Enter the reporting/QC requirements, e.g., NAVFAC NW QC Level C, D, or E.

Turnaround time (TAT): TAT for contract work will be determined by a sample delivery group (SDG), which may be formed over a 14-day period, not to exceed 20 samples. Standard turnaround time once the SDG has been completed is 35 calendar days from receipt of the last sample in the SDG. Entering NORMAL or STANDARD in this field will be acceptable. If quicker TAT is required, it shall be in the subcontract with the laboratory and reiterated on each COC to remind the laboratory.

Box 5 Type of containers: The type of container used, e.g., 1-liter glass amber, for a given parameter in that column.

Preservatives: Field personnel must indicate on the COC the correct preservative used for the analysis requested. Indicate the pH of the sample (if tested) in case there are buffering conditions found in the sample matrix.

Box 6 Sample number: Five-character alpha-numeric identifier to be used by the laboratory to identify samples. The use of this identifier is important since the labs are restricted to the number of characters they are able to use. See SOP I-A-11, Sample Naming.

Description (sample identification): This name will be determined by the location and description of the sample, as described in SOP I-A-11, Sample Naming. This sample identification should not be submitted to the laboratory, but should be left blank. If a computer COC version is used, the sample identification can be input but printed with this block black. A cross-referenced list of sample number and sample identification must be maintained separately.

Date Collected: Collection date must be recorded in order to track the holding time of the sample. Note: For trip blanks, record the date it was placed in company with samples.

Time Collected: When collecting samples, record the time the sample is first collected. Use of the 24-hour military clock will avoid a.m. or p.m. designations; e.g., 1815 instead of 6:15 p.m. Record local time; the laboratory is responsible for calculating holding times to local time.

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Lab Identification: This is for laboratory use only.

- Box 7 Matrix and QC: Identify the matrix: e.g., water, soil, air, tissue, fresh water sediment, marine sediment, or product. If a sample is expected to contain high analyte concentrations, e.g., a tank bottom sludge or distinct product layer, notify the laboratory in the comment section. Mark an "X" for the sample(s) that have extra volume for laboratory QC matrix spike/matrix spike duplicate (MS/MSD) purposes. The sample provided for MS/MSD purposes is usually a field duplicate.
- Box 8 Analytical Parameters: Enter the parameter by descriptor and the method number desired. When requesting metals that are modifications of the standard lists, define the list in the comment section. This would not be necessary when requesting standard list metals such as priority pollutant metals (PPM), target compound list from ILM03.0, and Title 22 metals which are groups of metals commonly requested and should not cause any confusion as to what metals are being analyzed. Whenever possible, list the parameters as they appear in the laboratory subcontract to maintain consistency and avoid confusion.

In the boxes below the analytical parameter, indicate the number of containers collected for each parameter by marking an "X". If more than one container is used for a sample, write a number in the desired box to indicate a request for analysis and to indicate the number of containers sent for that analysis.

Box 9 Sampler's Signature: The person who collected samples must sign here.

Relinquished By: This space shall contain the signature of the person who turned over the custody of the samples to a second party other than an express mail carrier such as FEDEX, DHL or Air Borne Express.

Received By: Typically, this is a written signature by a representative of the receiving laboratory, or a field crewmember who delivered the samples in person from the field to the laboratory. A courier such as FedEx or DHL does not sign because they do not open the coolers. It must also be used by the prime contracting laboratory when samples are sent to a subcontractor.

Relinquished By: In the case of subcontracting, the primary laboratory will sign the Relinquished By space and fill out an additional COC to accompany the samples being subcontracted.

Received By (Laboratory): This space is for the final destination (e.g., at a subcontracted laboratory).

- Box 10 Lab Number and Questions: This box is to be filled in by the laboratory only.
- Box 11 Control Number: This number is the "COC" followed by the first sample number in a cooler, or contained on a COC. This control number must be unique and never used twice. Record the date the COC is completed. It should be the same date the samples are collected.
- Box 12 Total No. of Containers/row: Sum the number of containers in that row.
- Box 13 Total No. of Containers/column: Sum the number of containers in that column.

Because COC forms contain different formats based upon who produced the form, not all of the information listed in items 1 to 13 may be recorded. However, as much of this information as possible shall be included.

COC forms tailored to each Task Order can be drafted and printed onto multi-ply forms. This eliminates the need to rewrite the analytical methods column headers each time. It also eliminates the need to write the project manager, name, and number; QC Level; TAT; and the same general comments each time.

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Complete one COC form per cooler. Whenever possible, reduce the number of trip blanks by placing all samples to be analyzed for VOA, gasoline, and BTEX compounds into one cooler. Complete all sections and be sure to sign and date the COC form. One copy of the COC form must remain with the field personnel.

3.0 DOCUMENTATION

The COC/analytical request form shall be faxed daily, if possible, to the Task Order Laboratory Coordinator for accuracy verification. Following the completion of sampling activities, the sample logbook and COC forms will be transmitted to the Project Manager for storage in project files. The Project Manager shall review COC forms on a monthly basis at a minimum. The data validators shall also receive a copy. Along with the data delivered, the original COC/analytical request form shall be submitted by the laboratory. Any changes to the analytical requests that are required shall be made in writing to the laboratory. A copy of this written change shall be sent to the data validators and placed in the project files. The reason for the change shall be included in the project files so that recurring problems can be easily identified.

4.0 REFERENCES

SOP I-A-11, Sample Naming

SOP III-D, Logbooks

State of California Water Resources Control Board. 1988. Technical Guidance Manual for Solid Waste Water Quality Assessment Test (SWAT) Proposals and Reports.

USEPA. 1986. EPA NEIC Policies and Procedures, National Enforcement Investigations Center, Denver, Colorado.

USEPA. 1988. Guidance for Conducting Remedial Investigations and Feasibility Studies Under CERCLA (EPA OSWER Directive 9355 3-01).

USEPA. 1992. RCRA Ground Water Monitoring Technical Enforcement Guidance Document (TEGD).

USEPA. 1995 and as updated. Test Methods for Evaluating Solid Waste (SW-846), Third edition.

5.0 ATTACHMENTS

Attachment III-E-1 Chain-of-Custody Seal

Attachment III-E-2 Generic Chain-of-Custody/Analytical Request Form

Attachment III-E-3 Sample Completed Chain-of-Custody/Analytical Request Form

Attachment III-E-4 Sample Out-of-Control Form

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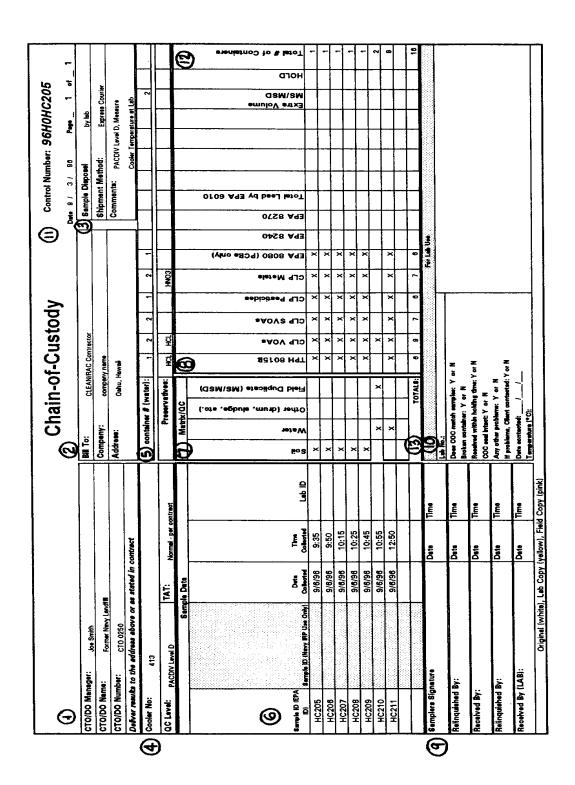
Attachment III-E-1 Chain-of-Custody Seal

	SAMPLE NO.	DATE	SEAL BROKEN BY
[LABORATORY]	SIGNATURE		DATE
	PRINT NAME AND TIT	LE (Inspector, Analyst or T	echnician

Attachment III-E-2 Generic Chain-of-Custody/Analytical Request Form

						Cha	Chain-of-Custody	્યુ	ısto	þ				Control Number: 94H0	Vumber: Deta	94H0		-	
CTO/DO Menager:	inager:				<u>ت</u> ة			:						Semple Dispose	-				
CTO/DO Neme:	30:				Company:	. <u>.</u>							1	Shipment					
CTO/DO Number	mber:				Address:								_	Comments:					
Defiver resul	Defiver results to the address above or as stated in contract	stated in co	ontract										\dashv	Ì	ŀ	İ	ŀ		T
Cooler No:						# of containers:	tainers:		\dashv	4		_	4		\dashv		\dashv		
QC Level:	AT	TAT:				Preser	Preservatives:		\vdash			Н	H		Н		Н		
	Sample Date	Deta				Matrix/OC				L		-	_						
Comple D (CPA	(And on the state of the state	Defense Collected	Thre	Qi qe'i	į os	Water Other (drum, eludge, etc.)	(MSM\8M) etacilidad bleif	83108 H 9 T	ON GID	CLP SVOA=	CLP Metals	EPA 8080 (PCBs only)	EPA 8240	Total Lead by EPA 6010	-		Extra Volume	HOLD MS/MSD	anenistnoo to & latoT
	L								-			\dashv	\dashv	1	4		+		
						+			\dashv	_	1	+	\downarrow	1	4		+		T
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							TOTALS:		\dashv		-	-	_	1	-		-		
Samplers Signature	gnature		Date	Tine								For Last Use							
Refinquished By:	l By:		Dete Dete	Time	Doe CO	Does COC match samples:	Does COC metch semples: Y or N	2											
Received By:	e		Dete	Time	A COC	Received within holding to COC seel intset: Y or N	Received within holding time: Y or N COC seed intset: Y or N	z											
Relinquished By:	I By:		Dete	Time	Any other	probleme.	Any other problems: Y or N H problems, Clent contacted: Y or N	z ŏ											-
Received By (LAB)	(LAB):		Date	Time	Date contacted: Temperature (°C):	anted: ure (°C):													
	Original (white), Leb Copy (yellow), Field Copy (pink)	ab Copy ()	rellow), Fiek	d Copy (pink)									1						

Attachment III-E-3 Sample Completed Chain-Of-Custody/ Analytical Request Form



Attachment III-E-4 Sample Out-Of-Control Form

			Status	Date		Initial
			Noted OOC			
	OUT OF CONTROL FOR	M	Submit for CA*			
			Resubmit for CA*			
			Completed			
Date F	Recognized:	By:			Sample	es Affected
Dated	Occurred:	Matrix			(List by	Accession
Param	eter (Test Code):	Method:			AND S	ample No.)
Analy	st:	Superviso	or:			
1. Ty	oe of Event	2. Correc	ctive Action (CA)*			
	(Check all that apply)		(Check all that apply)			
	Calibration Corr. Coefficient <0.995		Repeat calibration			
	%RSD>20%		Made new standards			
	Blank >MDL		Reran analysis			
	Does not meet criteria:		Sample(s) redigested and rerun			
	Spike		Sample(s) reextracted and rerun			
	Duplicate		Recalculated			
	LCS		Cleaned system			
	Calibration Verification		Ran standard additions			
	Standard Additions		Notified			·
	MS/MSD		Other (please explain)			
	BS/BSD					
	Surrogate Recovery					
	Calculations Error					
	Holding Times Missed					
	Other (Please explain	Commen	ts:			
3. Res	sults of Corrective Action					
	Return to Control (indicated with)					
	•					
	Corrective Actions Not Successful - DATA	A IS TO BE FLA	GGED with			
Analy	st:	Date:				
Super	visor:	Date:				
OA D	anartment:	Date				



Page 1 of 7

SAMPLE CONTAINERS AND PRESERVATION

1.0 PURPOSE

This standard operating procedure (SOP) describes the conventional containers used for sample collection and delivery to a laboratory for analysis. Additionally it will discuss sample preservation and holding times.

2.0 PROCEDURES

The purpose of sample preservation is to prevent or retard the degradation and modification of chemicals or to retard biological activity in samples during transit and storage. Efforts to preserve the integrity of the samples must be initiated as soon as possible after the time of sampling and continue until analyses are performed. Preservatives must be added to the sample container as soon as possible after the time of sample collection. The recommended procedure is to request that bottles be provided by the analytical laboratory and be pre-preserved.

Complete and unequivocal preservation of samples, domestic sewage, industrial wastes, or natural waters, is impossible in practice. Regardless of the nature of the sample, complete stability for every constituent is not likely to be achieved. At best, preservation techniques can retard the chemical and biological changes that inevitably continue after the sample is removed from the parent source. Degradation of the sample ceases only if it is preserved at a temperature of absolute zero (-273°C). However, freezing of a sample to extend hold times is not permitted. Therefore, as a general rule, it is best to analyze the samples as soon as possible after collection. This is especially true when the analyte concentration is expected to be in the low microgram per liter (mg/l) range.

Methods of preservation are relatively limited and are intended generally to perform the following:

- Retard biological action
- Retard hydrolysis of chemical compounds and complexes
- Reduce volatility of constituents
- Reduce absorption effects

Preservation methods are generally:

- pH control
- Chemical addition
- Refrigeration and/or chilling using ice

The recommended preservative for various constituents is given in the Exhibits at the end of this SOP. Preservation techniques for some analyses requiring more than simple refrigeration or filtering are discussed in Section 2.2. The exhibits also provide the estimated volume of sample required for the analysis, the suggested type of container, and the maximum recommended holding times for samples to be properly preserved.

When selecting preservation techniques and sample container type, always refer to the guidance provided in the documentation of the analytical methods to be used.

2.1 SAMPLE CONTAINERS

Select sample containers based on the analytical parameters of interest. Use containers made of materials that are non-reactive. Glass and polyethylene containers are the most commonly accepted, and both are used when sampling many constituents. When metals are the analytes of interest, however, polyethylene containers with Teflon-lined caps are preferred. When organics are the analytes of interest, use glass containers with Teflon-lined caps.

2.2 SAMPLE PRESERVATION

Utilize pre-preserved sample bottles whenever possible. If this is not possible or practical, perform appropriate chemical preservation in the field for various analytical parameters as soon as possible after the time of sample collection. Cool samples after collection and during shipment. All samples should be kept out of direct sunlight as much as possible and stored in the dark (e.g., in a cooler). Regardless of the method of preservation, analyses should be performed as soon after sampling as possible.

In some instances, the optimal method for sample preservation may be inappropriate due to the restrictions placed on the transport of certain chemicals by shippers. When shipping restrictions prevent the use of some reagents for sample preservation, use the most appropriate and permissible technique. The project chemist or laboratory should be able to assist in deciding the best alternative method of preservation.

2.3 MAXIMUM HOLDING TIME

Complete and unequivocal preservation of a sample for an extended period of time is a practical impossibility. Regardless of the nature of the sample, complete stability for every constituent is not likely to be achieved. Maximum holding times are assigned to each analyte and are designed for quality assurance purposes to minimize degradation effects on the analysis. Therefore, as a rule, it is better to analyze the sample as soon as possible after collection. This is especially true when low contaminant concentrations are expected.

2.4 REVIEW

The Field Manager or an approved designee shall check all sample control documentation to ensure that the samples, transport, and analysis events have met the criteria outlined in this SOP and the field sampling plan. Any discrepancies shall be noted and the documentation will be returned to the originator for correction or explanation. The reviewer will acknowledge that corrections have been incorporated by signing and dating each reviewed document.

3.0 DOCUMENTATION

All sample collection information must be recorded within the field logbook. Each sample collected will be clearly associated with a sample type (i.e. normal, field duplicate, equipment blank) sample location, matrix type, collection time, collection date, analysis and sampling depth if appropriate.

With every sample submitted for analysis, a completed chain of custody (COC) must accompany the shipment and a copy retained for the project records. The COC/analytical request form must be used to track all sample identifiers.

4.0 REFERENCES

None.

5.0 ATTACHMENTS

Attachment III-F-1 Example Sample Collection Form

Attachment III-F-1 Example Sample Collection Form

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	SA	MPLE CO	FORM 11-2 LLECTION INI	FOR	RMATIC)N			
Installation ID:		Establishing	Contract ID:		Prime (Contractor N	lame:		
Site Name:		DO/CTO:	Establishing Phas	se:			Collection D	ate:	
Location Name	Sample Name	Depth R	ange (feet bgs)		ollection Time	Sample Matrix	Sample Type	Sampling Equipment	Composite (Y/N)
		Start Depth	End Depth		Tillie	IVIAUIA	Туре	Ечиринен	(1/14)
2V Double Van Veen Grab AC Air Canister AP Pump - Air Lift (bladder) AS Ashing B Bailer BR Brass (California) Ring BS Beach Seine C Continuous Flight Auger CC Continuous Core Sampler CH Charcoal Sampling Tube CL Clover Leaf Dredge Sampler CN Cone Penetrometer CO Core Sampler CP Pump - Centrifugal DF Diffusion Bag Samplers	DG Drill Rig DS Dredge Sampler (brass, etc.) DT Driver Tube (geoprobe, direct push, CPT rig) E2 Pump - Electric Submersible EC Encore Soil Sampler FC Cassette Filter G Grab GP Gas-Operated, Double Acting Piston Pump H Hollow Stem Auger HA Hand Auger HB Bucket Auger HC Hand Collected HD Hand Drill - Portable Powered HK Hook and Line HP Hydropunch	HU Air Samp HV Air Samp HX Air Samp IF Isolation LV Air Samp LY Lysimeter MPPS Micro Pt NC Nickel Co NX NX Rock PP Pump - P PS Passive S PU Pump - S RS Hollow G SC Scoop/Tr	ush Point Sampler pated Brass Bomb Sample Coring teristaltic Soil Gas Sampling Probe standard, Type not Recorde lass Sampling Rod	esin Resin nus	SK SS ST SY T TB TL TR TS UNN VC VS VV W WB	Syringe Shelby Tube Tedlar Bag Trawl Animal Trap Thief Sampl (Unknown Vacuum (ga Van Dorn Se Van Veen Swab or Wi	er and/or Thief T is) ampler	ype Sampler	
Recorder:							Da	te:	
Checker							Da	te:	

			Sample Matrix		
۸۸	Ambient Air	MA	Sample Matrix Mastic	WI	Cround Water Influent (into avetem)
AA AC	Composite Air Sample	MO	Mortar	WL	Ground Water Influent (into system) Leachate
ACS	Air - Crawlspace	MR	Marine Sediment	WM	Marine Water
AD	Air - Crawispace Air - Drilling	MS	Metal Shavings	WN	Porewater
Al	Air - Indoor	NS	Near-Surface Soil	WO	Ocean Water
AIN	Integrated Air Sample	PA	Paper	WP	Drinking Water
AIR-ABS	Activity Based Sampling (ABS) Air	PC	Paint Chips	WQ	Water for QC Samples
AIIT-ABO	Sample		r anti Onips	WQ	water for QO campies
AO	Air - Outdoor	PP	Precipitate	WR	Ground Water Effluent (from system)
AQ	Air Quality Control Matrix	RE	Residue	WS	Surface Water
AQS	Aqueous	RK	Rock	WT	Composite Ground Water Sample
ASB	Asbestos	SB	Bentonite	WU	Storm Water
ASBF	Asbestos-Fibrous	SBS	Sub-Surface Soil (>6")	WW	Waste Water
ASBNF	Asbestos-Non-Fibrous	SC	Cement/Concrete	XR	XRF Data
AVE	Air-Vapor Extraction, Effluent	SD	Drill Cuttings - Solid Matrix		
AX	Air Sample from Unknown Origin	SE	Sediment		Sample Type
BK	Brick	SEEP	Seep	AB	Ambient Condition Blank
BS	Brackish Sediment	SF	Filter Sandpack	BIOCON	Bioassay Control Sample
CA	Cinder Ash	SJ	Sand	BS	Blank Spike
CK	Caulk	SK	Asphalt	BSD	Blank Spike Duplicate
CN	Container	SL	Sludge	EB	Equipment Blank
CR	Carbon (usually for a remediation system)	SM	Water Filter (solid material used to filter water)	EBD	Equipment Blank/Rinsate Duplicate
DF	Dust/Fallout	SN	Miscellaneous Solid Materials - Building Materials	FB	Field Blank
DR	Debris/Rubble	SO	Soil	FD	Field Duplicate
DS	Storm Drain Sediment	SP	Casing (PVC, stainless steel, cast iron, iron pipe)	FR	Field Replicate
DT	Trapped Debris	SQ	Soil/Solid Quality Control Matrix	FS	Field Spike
EF	Emissions Flux	SS	Scrapings	IDW	Purge and Rinseate Water
EW	Elutriate Water	SSD	Subsurface Sediment	LB	Lab Blank
FB	Fibers	STKG	Stack Gas	LR	Lab Replicate
FL	Forest Litter	STPM	Stripper Tower Packing Media	MB	Material Blank
GE	Soil Gas Effluent - Stack Gas (from system)	SU	Surface Soil (less than 6 inches)	MIS	Multi-Incremental Sample
GI	Soil Gas Influent (into system)	SW	Swab or Wipe	MS	Matrix Spike
GL	Headspace of Liquid Sample	SZ	Wood	N	Normal (Regular)
GQ	Gaseous or Headspace QC	TA	Animal Tissue	PE	Performance Evaluation
GR	Gravel	TP	Plant Tissue	PURGE	Purge Water Sample
GS	Soil Gas	TQ	Tissue QC	RD	Regulatory Duplicate
GSS	Soil Gas - Subslab	TX	Tissue	SB	Source Blank
GT	Grit	UNK	Unknown	SBD	Source Blank Duplicate
IC	IDW Concrete	W	Water (not groundwater, unspecified)	SCREEN	Screening Sample
IDD	IDW Solid	WA	Drill Cuttings - Aqueous Mix	SD	Matrix Spike Duplicate
IDS	IDW Soil	WB	Brackish Water	SPLIT	Sample Split
IDW	IDW Water	WC	Drilling Water (used for well construction)	SRM	Standard Reference Material
IW	Interstitial Water	WD	Well Development Water	TB	Trip Blank
LA	Aqueous Phase of Multiphase Liquid/Soil	WF	Freshwater (not groundwater)	TBD	Trip Blank Duplicate
LF	Product (floating or free)	WG	Ground Water	TBR	Trip Blank Replicate
LQ	Organic Liquid Quality Control	WH	Equipment Wash Water (i.e. water		
	Matrix		used for washing equipment)		

Instructions

Form 11-2 (Sample Collection Information)

The purpose of this form is to collate sample collection information for data entry to serve as a quick reference for sample information. Every sample that is collected should be recorded on one of these forms. The information recorded on this form must come from the field logbook, which is the official record. This form must be filled out in its entirety; if a value or piece of information is unknown or not applicable, a horizontal line should be drawn through that field.

The information on this form must be checked against the field logbook for accuracy and completeness by a field staff member before the form is submitted for data entry. Data from this form will not be entered without the signature of the individual who checked the form for accuracy and completeness.

Installation ID: Unique identifier for installation associated with the location (example: WHIDBEY)

Establishing Contract ID: Unique contract ID assigned by Division Contraction Office (example: D459559365800)

Prime Contractor Name: Name of company that established location (example: URS)

Site Name: Site name associated with the location (example: Site 11)

DO/CTO: Contract Task Order (CTO) or Delivery Order (DO) number assigned by the Navy. The format is

NNNN (example: 0012)

Establishing Phase: Task Phase, Subtask Number or Annual Quarter (example: 1)

Collection Date: Date samples were collected

Location Name: Unique name used for the location (example: MW-2R)

Sample Name: Unique sample name assigned by the contractor and/or derived from historical data submittal

(example: MW-1-11/02/98)

Depth Range (feet bgs): Start and end depth of sample collection, if applicable.

Collection Time: Time at which sample was collected

Sample Matrix: Matrix type code from options at the bottom of form (example: MR)

Sample Type: Sample type code from options at bottom of form (example: N)

Sampling Equipment: Sampling equipment code from options at bottom of form (example: G)

Composite: A Y/N field indicating whether or not the sample is a composite

Recorder: Signature of individual who completed form and date completed

Checker: Signature of individual who checked the data against the field logbook and date checked



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SAMPLE HANDLING, STORAGE, AND SHIPPING

1.0 PURPOSE

This standard operating procedure (SOP) sets forth the methods for use by U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) field personnel and their contractors engaged in handling, storing, and transporting water, soil and/or sediment samples.

2.0 PROCEDURE

2.1 HANDLING AND STORAGE

Immediately following collection, all samples will be labeled according to the procedures in SOP III-E, *Record Keeping, Sample Labeling, and Chain-of-Custody Procedures*. The lids of the containers shall not be sealed with duct tape, but may be covered with custody seals or placed directly into sealed plastic bags. The sample containers shall be placed in an insulated cooler with frozen gel packs (such as "blue ice") or ice in double, self-sealing bags. Samples should occupy the lower portion of the cooler, while the ice should occupy the upper portion. An absorbent material (e.g., proper absorbent cloth material) may be placed on the bottom of the cooler to contain liquids in case of spillage. All empty space between sample containers shall be filled with bubble wrap, Styrofoam "peanuts," or other appropriate material. Prior to shipping, glass sample containers should be wrapped on the sides, tops, and bottoms with bubble wrap or other appropriate padding and/or surrounded by packing material to prevent breakage during transport. Prior to shipment, the ice or cold packs in the coolers may require replacement to maintain samples as close to 4°C as possible during transport of the samples to the analytical laboratory. Samples shall be shipped as soon as possible to allow the laboratory to meet holding times for analyses. The procedures for maintaining sample temperatures at 4°C, pertains to all water, soil, and sediment field samples.

2.2 SHIPPING

All appropriate U.S. Department of Transportation (DOT) regulations (e.g., 49 Code of Federal Regulations (CFR), Parts 171-179) shall be followed in shipment of air, soil, water, and other samples.

2.2.1 Hazardous Materials Shipment

Field personnel must state whether any sample is suspected to be a hazardous material. A sample should be assumed to be hazardous unless enough evidence exists to indicate it is nonhazardous. If not suspected to be hazardous, shipments may be made as described in the Section 2.2.2 for non-hazardous materials. If hazardous, the procedures summarized below must be followed.

Any substance or material that is capable of posing an unreasonable risk to life, health, or property when transported is classified as hazardous. Hazardous materials identification should be performed by checking the list of dangerous goods for that particular mode of transportation. If not on that list,

materials can be classified by checking the Hazardous Materials Table (49 CFR 172.102 including Appendix A) or by determining if the material meets the definition of any hazard class or division (49 CFR Part 173), as listed in Attachment III-G-2.

All persons offering for shipment any hazardous material <u>must</u> be properly trained in the appropriate regulations, as required by HM-126F, Training for Safe Transportation of Hazardous Materials. The training covers loading, unloading, handling, storing, and transporting of hazardous materials, as well as emergency preparedness in the case of accidents. Carriers such as commercial couriers must also be trained.

When shipping hazardous materials, including bulk chemicals or samples suspected of being hazardous, the proper shipping papers (49 CFR 172 Subpart C), package marking (49 CFR 172 Subpart D), labeling (49 CFR 172 Subpart E), placarding (49 CFR 172 Subpart F, generally for carriers), and packaging must be used. Attachment III-G-1 shows an example of proper package markings. A copy of 49 CFR should be referred to each time a hazardous material or potentially hazardous samples are shipped.

According to Section 2.7 of the International Air Transport Association (IATA) Dangerous Goods Regulations publication, very small quantities of certain dangerous goods may be transported without certain marking and documentation requirements as described in 49 CFR Part 172. However, other labeling and packing requirements must still be followed. Attachment III-G-2 shows the volume or weight for different classes of substances. A "Dangerous Goods in Excepted Quantities" label must be completed and attached to the associated shipping cooler (Attachment III-G-3). Certain dangerous goods are not allowed on certain airlines in any quantity.

As stated in item 4 of Attachment III-G-4, the Hazardous Materials Regulations do not apply to hydrochloric acid (HCl), nitric acid (HNO₃), sulfuric acid (H₂SO₄), and sodium hydroxide (NaOH) added to water samples if their pH or percentages by weight criteria are met. These samples may be shipped as non-hazardous materials as discussed below.

2.2.2 Nonhazardous Materials Shipment

If the samples are suspected to be nonhazardous, based on previous site sample results, field screening results, or visual observations, if applicable, then samples may be shipped as nonhazardous.

When a cooler is ready for shipment to the laboratory, copies of the chain-of-custody form shall be placed inside a sealed plastic bag and placed inside of an insulated cooler. The coolers will then be sealed with waterproof tape and labeled "Fragile," "This-End-Up" (or directional arrows pointing up), or other appropriate notices. Custody seals will be placed on the coolers as discussed in SOP III-E, *Record Keeping, Sample Labeling, and Chain-of-Custody Procedures*.

2.2.3 Shipments from Outside the Continental United States

Shipment of sample coolers to the U.S. from locations outside the continental U.S. is controlled by the USDA and is subject to their inspection and regulation. Documentation is required to prove that the analytical laboratory receiving samples is certified. The laboratory must have certification by USDA to receive and properly dispose of soil; this is called a "USDA Soil Import Permit." In addition, all sample coolers must be inspected by a USDA representative, affixed with a label indicating that the coolers contain environmental samples, and shipping forms stamped by the USDA inspector prior to shipment. In addition, samples shipped from U.S. territorial possessions or foreign countries, must be cleared by the

U.S. Customs Service upon entry into the United States. As long as the commercial invoice is properly completed (see below), shipments typically pass through U.S. Customs without the need to open coolers for inspection.

Completion and use of proper paperwork will, in most cases, minimize or eliminate the need of the USDA and U.S. Customs to inspect the contents. Attachment III-G-5 shows an example of how paperwork may be placed on the outside of coolers for nonhazardous materials. For hazardous materials, refer to Section 2.2.1.

In summary, the paperwork listed below should be taped to the outside of the coolers to assist sample shipments. If a shipment is made up of multiple pieces (e.g., more than one cooler), the paperwork need be attached only to one cooler, provided that the courier agrees. All other coolers in the shipment need only be taped and have address and chain-of-custody seals affixed.

- 1. **Courier Shipping Form & Commercial Invoice** See Attachments III-G-6, III-G-7, and III-G-8 for examples of the information to be included on these forms. Both forms should be placed inside a clear plastic adhesive-backed pouch, which adheres to the package (typically supplied by the courier) and placed on the cooler lid as shown in Attachment 5.
- 2. **Soil Import Permit and USDA Letter** (soil only) See Attachments III-G-9 and III-G-10 for examples. The laboratory shall supply these documents prior to mobilization. The USDA in Hawaii often <u>does</u> stop shipments of soil without these documents. The 2" x 2" USDA label (described below), the USDA letter, and soil impact permit should be stapled together and placed inside a clear plastic pouch. Clear plastic and adhesive-backed pouches are typically supplied by the mailing courier.
- 3. The analytical laboratory should supply the Soil Import Permit. Although original labels are preferred, copies of this label, which are cut out to the 2" x 2" dimensions, are acceptable. Placing one label (as shown in Attachment III-G-5) covered with clear packing tape and one stapled to the actual permit is suggested.
- 4. The USDA does not control water samples, thus the requirements for soils listed above do not apply.
- 5. **Custody Seals**. Task Order personnel must sign and date custody seals. At least two seals should be placed in such a manner that they stick to both the cooler lid and body. The seals shall be placed so the cooler/container cannot be opened without breaking the seal. The custody seals are then covered with clear packing tape. This prevents the seal from coming loose and enables detection of tampering.
- 6. **Address Label**. A label stating the destination (the sending and laboratory, company, or location address) should be affixed to each cooler. The label should also include both telephone numbers.
- 7. Special Requirements for Hazardous Materials see Section 2.2.1.

Upon receipt of sample coolers at the laboratory, the sample custodian shall inspect the sample containers as discussed in SOP III-E, *Record Keeping, Sample Labeling, and Chain-of-Custody Procedures*. The samples shall then be immediately extracted and/or analyzed, or stored in a refrigerated storage area until they are removed for extraction and/or analysis. Whenever the samples are not being extracted or analyzed, they shall be returned to refrigerated storage.

3.0 DOCUMENTATION

Records shall be maintained as required by implementing these procedures.

4.0 REFERENCES

HM-126F, Training for Safe Transportation of Hazardous Materials

SOP III-E, Record Keeping, Sample Labeling, and Chain-of-Custody Procedures

5.0 ATTACHMENTS

Attachment III-G-1 Example Package Marking

Attachment III-G-2 Packing Groups

Attachment III-G-3 Label for Dangerous Goods in Excepted Quantities

Attachment III-G-4 SW-846 Preservative Exception

Attachment III-G-5 Sample Cooler Marking Figure

Attachment III-G-6 Example Courier Form

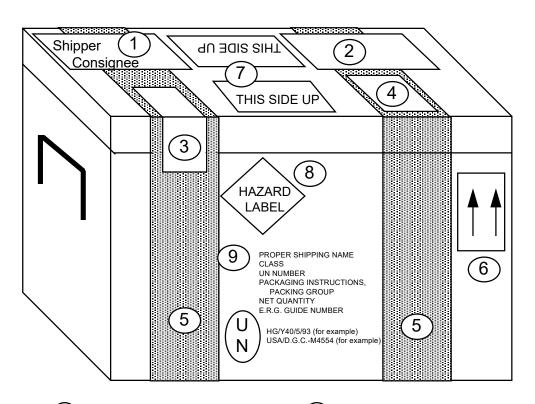
Attachment III-G-7 Commercial Invoice - Soil

Attachment III-G-8 Commercial Invoice - Water

Attachment III-G-9 Soil Import Permit

Attachment III-G-10 Soil Samples Restricted Entry Labels

Attachment III-G-1 Example Hazardous Material Package Marking



- AIR BILL/COMMERCIAL INVOICE
- **USDA PERMIT (Letter to** Laboratory from USDA)
- **CUSTODY SEAL**
- USDA 2" X 2" SOIL IMPORT PERMIT (9
- WATERPROOF STRAPPING TAPE **PACKAGE SPECIFICATIONS**
- **TWO REQUIRED**

DIRECTION ARROWS STICKER -

- THIS SIDE UP STICKERS
- **HAZARD LABEL**
 - HAZARDOUS MATERIAL INFORMATION

Attachment III-G-2 Packing Groups

Packing	g Group of the Substance	Packing	Group I	Packing	Group II	Packing	Group III
	S or DIVISION of PRIMARY or DIARY RISK	Packa	ngings	Packa	ngings	Packa	gings
		Inner	Outer	Inner	Outer	Inner	Outer
1:	Explosives			Forbidd	len (Note A)		
2.1:	Flammable Gas			Forbidd	len (Note B)		
2.2:	Non-Flammable, non-toxic gas			See Note	s A and B		
2.3:	Toxic gas			Forbidd	len (Note A)		
3.	Flammable liquid	30 mL	300 mL	30 mL	500 mL	30 mL	1 L
4.1	Self-reactive substances	Forb	idden	Forb	idden	Forb	idden
4.1:	Other flammable solids	Forb	idden	30 g	500 g	30 g	1 kg
4.2:	Pyrophoric substances	Forb	idden	Not Ap	plicable	Not Ap	plicable
4.2	Spontaneously combustible substances	Not Ap	plicable	30 g	500 g	30 g	1 kg
4.3:	Water reactive substances	Forbidden		30 g or 30 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
5.1:	Oxidizers	Forb	idden	30 g or 30 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
5.2:	Organic peroxides (Note C)	See N	lote A	30 g or 30 mL	500 g or 250 mL	Not Ap	plicable
6.1:	Poisons - Inhalation toxicity	Forb	idden	1 g or 1 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
6.1:	Poisons - oral toxicity	1 g or 1 mL	300 g or 300 mL	1 g or 1 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
6.1:	Poisons - dermal toxicity	1 g or 1 mL	300 g or 300 mL	1 g or 1 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
6.2:	Infectious substances			Forbidd	len (Note A)		
7:	Radioactive material (Note D)			Forbidd	len (Note A)		
8:	Corrosive materials	Forb	idden	30 g or 30 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
9:	Magnetized materials			Forbidd	len (Note A)		
9:	Other miscellaneous materials (Note E)	Forb	idden	30 g or 30 mL	500 g or 500 mL	30 g or 30 mL	1 kg or 1 L
				l		1	

Note A: Packing groups are not used for this class or division.

Note B: For inner packagings, the quantity contained in receptacle with a water capacity of 30 mL. For outer packagings, the sum of the water capacities of all the inner packagings contained must not exceed 1 L.

Note C: Applies only to Organic Peroxides when contained in a chemical kit, first aid kit or polyester resin kit.

Note D: See 6.1.4.1, 6.1.4.2 and 6.2.1.1 through 6.2.1.7, radioactive material in excepted packages.

Note E: For substances in Class 9 for which no packing group is indicated in the List of Dangerous Goods, Packing Group II quantities must be used.

Attachment III-G-3 Label For Dangerous Goods In Excepted Quantities

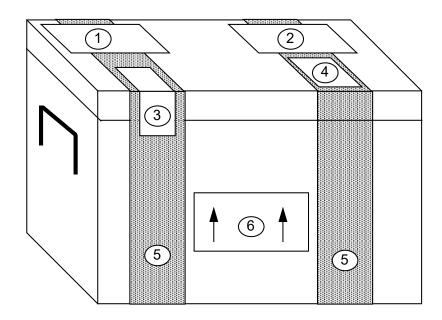
	nce with t		ole internati				s in all respects in ulations and the
Signatui	e of Ship	per					
Title				Date			
This pac	kage con	s of Shippe tains substate box(es))	r ance(s) in C	lass(es)			
This pac	kage con	tains substa		lass(es)	6	8	9
This pac	kage con pplicable	tains substate box(es))	ance(s) in C		6	8	9
This pac (check a Class:	ekage con applicable 2	tains substate box(es))	ance(s) in C 4	5			

ATTACHMENT III-G-4 Preservative Exception

Measurement	Vol. Req. (mL)	Container ²	Preservative ^{3,4}	Holding Time ⁵
MBAS	50	P,G	Cool, 4°C	48 Hours
NTA	0	P,G	Cool, 4°C	24 Hours

- 1. More specific instructions for preservation and sampling are found with each procedure as detailed in this manual. A general discussion on sampling water and industrial wastewater may be found in ASTM, Part 31, p. 72-82 (1976) Method D-3370.
- 2. Plastic (P) or Glass (G). For metals, polyethylene with a polypropylene cap (no liner) is preferred.
- 3. Sample preservation should be performed immediately upon sample collection. For composite samples each aliquot should be preserved at the time of collection. When use of an automated sampler makes it impossible to preserve each aliquot, then samples may be preserved by maintaining at 4°C until compositing and sample splitting is completed.
- 4. When any sample is to be shipped by common carrier or sent through the United States Mail, it must comply with the Department of Transportation Hazardous Materials Regulations (49 CFR Part 172). The person offering such material for transportation is responsible for ensuring such compliance. for the preservation requirements of Table 1, the Office of Hazardous Materials, Materials Transportation Bureau, Department of Transportation has determined that the Hazardous Materials regulations do not apply to the following materials: Hydrochloric acid (HCl) in water solutions at concentration of 0.04% by weight or less (pH about 1.96 or greater); Nitric acid (HNO₃) in water solutions at concentrations of 0.15% by weight or less (pH about 1.62 or greater); Sulfuric acid (H₂SO₄) in water solutions at concentrations of 0.35% by weight or less (pH about 1.15 or grater); Sodium hydroxide (NaOH) in water solutions at concentrations of 0.080% by weight or less (pH about 12.30 or less).
- 5. Samples should be analyzed as soon as possible after collection. The times listed are the maximum times that samples may be held before analysis and still considered valid. Samples may be held for longer periods only if the permittee, or monitoring laboratory, has data on file to show that the specific types of sample under study are stable for the longer time, and has received a variance from the Regional Administrator. Some samples may not be stable for the maximum time period given in the table. A permittee, or monitoring laboratory, is obligated to hold the sample for a shorter time if knowledge exists to show this is necessary to maintain sample stability.
- 6. Should only be used in the presence of residual chlorine.

Attachment III-G-5 Non-Hazardous Material Cooler Marking Figure For Shipment From Outside the Continental United States



- 1 AIR BILL/COMMERCIAL INVOICE
- (2) USDA PERMIT (Letter to Laboratory from USDA)
- (3) CUSTODY SEAL
- 4 USDA 2" X 2" SOIL IMPORT PERMIT
- (5) WATERPROOF STRAPPING TAPE
- 6 DIRECTION ARROWS STICKER TWO REQUIRED

Attachment III-G-6 Example Courier Form

FedEx. USA Airbill Tracking Number 801704855619	O200 form Sender's Copy
1 From (please print and press hard) Account Number DateSender's FedEx Account Number	FedEx Priority Overnight [Next business morning] FedEx Standard Overnight [Next business aftermoon] FedEx ZDay*
Sender's Name Phone (808) 545-2462	FedEx First Overnight (Third business day)
Company OGDEN ENVIRONMENTAL/CRC ACCT	4b Express Freight Service Packages over 150 lbs. Delivery commitment may be later in some areas.
Address 680 IWILEI RD STE 660 Dept/Floor/Suite/Room	FedEx Overnight Freight FedEx 2Day Freight FedEx Express Saver Freight (Second business day) (Up to 3 business days) (Call for delivery schedule. See back for detailed descriptions of freight services.)
City HONOLULU State HI ZIP 96817	Packaging
Your Internal Billing Reference Information (Optional) (First 24 characters will appear on invoice)	6 Special Handling As per attached Special Handling
3 To (please print and press hard) Recipient's Sample Receipt Name Phone () Lab Phone #	FedEx FedEx FedEx FedEx FedEx Dther Pkg. 6 Special Handling Solution Pkg. Support Pkg. 6 Special Handling Solution Pkg. Pkg. Pkg. 6 Special Handling Solution Pkg. Pkg. Pkg. 6 Special Handling Solution Pkg. Pkg. Pkg. Pkg. 6 Special Handling Pkg. Pkg. Pkg. Pkg. Pkg. Pkg. 6 Special Handling Pkg. Pkg.
Lab Name Company	Bill Sender Recipient Third Party Credit Card Cash/ Check
Lab Address Address (Check here if residence (Extra charge applies for Felfx. Express Sever) (For 1901): at Felfx location. (We Cannot Deliver to P.D. Roxes or P.D. 7!P Codes) Deat/RoomSuite/Room	section 1 will be billed) (Enter FedEx account no. or Credit Card no. below) FedEx Account No.
(To 'H0LD' at FedEx location, (We Cannot Deliver to P.O. Boxes or P.O. ZIP Codes) Dept/Floor/Suite/Room print FedEx address here)	Credit Exp. Oate.
City State ZIP For HOLD at FedEx Location check here For Saturday Delivery check here	Total Packages Total Weight Total Declared Value Total Charges
Hold Weekday Not available with Gaturday (Not available at all locations)	*When declaring a value higher than \$100 per shipment, you pay an additional charge. See SERVICE CONDITIONS, DECLARED VALUE, AND LIMIT OF LUABILITY section for further information.
Service Conditions, Declared Value, and Limit of Liability – By using this Airbill, you agree to the service conditions in our current Service Guide or U.S. Government Service Guide. Both are available on request. SEE BACK OF SENDER'S COPY OF THIS AIRBILL FOR INFORMATION AND ADDITIONAL TERMS.	8 Release Signature Sign to authorize delivery without obtaining signature.
SENDER'S COPY OF THIS AIRBILL FOR INFORMATION AND ADDITIONAL TERMS. We will not be responsible for any claim in excess of \$100 per package whether the result of loss, damage, or delay, non-delivery, misdelivery, or misinformation, unless you declare a higher value, pay an additional charge, and document your "See the FedEx Service Guide for further details." See the FedEx Service Guide for further details.	Your signature authorizes Federal Express to deliver this ship- ment without obtaining a signature and agrees to indemnify and hold harmless Federal Express from any resulting claims. WCSL 0897
Questions? Call 1-800-Go-FedEx (800)463-3339 The World On Time	Rev. Date 5/97 Part 1/50054 0/1994-97 Fode: PRINTED IN U.S.A.

Name/Title

Signature

Attachment III-G-7 Commercial Invoice - Soil

DATE OF EXI	PORTATION	V		EXPO		ENCES (i.e., ord	er no., invoic	e no., etc.)	
SHIPPER/EXI Joe Smith Ogden c/o <hc< td=""><td>PORTER (controlled the controlled th</td><td>mplete name and a</td><td>ddress)</td><td>CONS Samp <lab< td=""><td>SIGNEE lle Receipt Name> Address></td><td></td><td></td><td></td><td></td></lab<></td></hc<>	PORTER (controlled the controlled th	mplete name and a	ddress)	CONS Samp <lab< td=""><td>SIGNEE lle Receipt Name> Address></td><td></td><td></td><td></td><td></td></lab<>	SIGNEE lle Receipt Name> Address>				
COUNTRY Of Guam, USA	F EXPORT			IMPC	ORTER - IF C	OTHER THAN C	CONSIGNEE		
COUNTRY Of Guam, USA	F ORIGIN O	F GOODS							
COUNTRY O	F ULTIMAT	E DESTINATION							
INTERNATIO AIR WAYBIL						(NOTE: All Federal Exp	l shipments n ress Internati	nust be accom onal Air Way	npanied by a bill)
MARKS/ NOS	NO. OF PKGS	TYPE OF PACKAGING	FULL DESCRIPT OF GOODS		QTY	UNIT OF MEASURE	WEIGH T	UNIT VALUE	TOTAL VALUE
NOS	3 coolers Soil samples for laboratory analyst							\$1.00	\$3.00
	TOTAL NO. OF PKGS.		L				TOTAL WEIGH T		TOTAL INVOICE VALUE
	3								\$3.00
									Check one F.O.B. C&F C.I.F.
THESE CO	MMODIT	TES ARE LICE	ENSED FOR TH	E UL	ГІМАТЕ І	DESTINATIO	ON SHOW	N.	
DIVERSIO	N CONTR	CARY TO UNI	TED STATES L	AW IS	S PROHIB	SITED.			
I DECLAR	E ALL TH	IE INFORMAT	TION CONTAIN	ED IN	N THIS IN	VOICE TO B	BE TRUE A	AND COR	RECT
SIGNATUF	RE OF SH	IPPER/EXPOR	TER (Type name	e and	title and si	gn)			
Joe Smith, 0	Ogden	Joe Smit	h			1/1/94			

Date

ATTACHMENT III-G-8 **Commercial Invoice - Water**

DATE OF EXI	PORTATIO	ON				ENCES (i.e., ord	er no., invoic	e no., etc.)	
1/1/94				<cto #=""></cto>	>				
SHIPPER/EXF	PORTER (c	omplete name and	address)	CONSIG	NEE				
Joe Smith		-		Sample F					
Ogden				<lab na<="" td=""><td></td><td></td><td></td><td></td><td></td></lab>					
	tel name> tel address	_		<lab ad<="" td=""><td>dress></td><td></td><td></td><td></td><td></td></lab>	dress>				
\no	nei address								
COUNTRY O	F EXPORT			IMPORT	ER - IF C	OTHER THAN C	CONSIGNEE		
Guam, USA									
COUNTRY OF	F ORIGIN (OF GOODS							
Guam, USA									
COLINTRY OF	FIIITIMA	TE DESTINATIO	N .						
USA	CLIIWIA	TE DESTINATIO	11						
INTERNATIO	NAL						l shipments n		
AIR WAYBIL	L NO.					Federal Exp	ress Internati	onal Air Way	bill)
MARKS/	NO.	TYPE OF	FULL DESCRIPTION	ON OF	QTY	UNIT OF	WEIGH	UNIT	TOTAL
NOS	OF	PACKAGING	GOODS			MEASURE	T	VALUE	VALUE
	PKGS								
	3	coolers	Water samples for la	aboratory				\$1.00	\$3.00
			analysis only						
	TOTA				I.	1	TOTAL		TOTAL
	L NO.						WEIGH		INVOICE
	OF						T		VALUE
	PKGS.								
	3	1							\$3.00
	_	_							
									Check one
									☐ F.O.B. ☐ C&F
									C.I.F.
THESE COMN	MODITIES	ARE LICENSED	FOR THE ULTIMAT	E DESTIN	ATION S	SHOWN.			
DIVERSION	CONTRAR	Y TO UNITED ST	TATES LAW IS PROF	HIRITED					
			ONTAINED IN THIS		TO DE T	DITE AND COR	DECT		
					IO BE I	KUE AND CUK	KEU I		
SIGNATURE	OF SHIPPE	EK/EXPORTER (Type name and title and	a sign)					

Joe Smith, Ogden	Joe Smith	1/1/94
Name/Title	Signature	Date

Attachment III-G-9 Soil Import Permit

UNITED STATES DEPARTMENT OF AGRICULTURE ANIMAL AND PLANT HEALTH INSPECTION SERVICE PLANT PROTECTION AND QUARANTINE PROGRAMS

COMPLIANCE AGREEMENT

1. NAME AND MAILING ADDRESS OF PERSON OR FIRM
Ogden Environmental & Energy Service Co.
680 Iwilei Road, Suite 660
Honolulu, HI 96817

Telephone: 545-2462
Fax: 528-5379

3. REGULATED ARTICLE(S)
Foreign soil samples destined to approved laboratories in the Continental United States transiting through Honolulu International Airport and military facilities on Oahu, Hawaii.

4. APPLICABLE FEDERAL QUARANTINE(S) OR REGULATIONS
7 CFR 330.300

6. I/We agree to the following:

See the attached Addendum, Foreign Soil Samples Destined To Approved Laboratories In The Continental United States Transiting Through Honolulu International Airport And Military Facilities On Oahu, Hawaii

THIS COMPLIANCE AGREEMENT IS VALID FOR 2 YEARS FROM THE DATE OF ISSUANCE. For renewal, call our office at 861-8446 or Fax 861-8450.

EXPIRATION DATE: SEPTEMBER 30, 2000

7. SIGNATURE Betsy D. aispauge	8. TITLE AIR & HAZARDAS WASE GROUP MANAGER	9. DATE SIGNED 9/9/98	
The affixing of the signatures below will validate this agreement which shall remain in effect until canceled, but may be revised as necessary or revoked for noncompliance.		10. AGREEMENT NO. OAHU-ST-002	
		11. DATE OF AGREEMENT September 2, 1998	
12. PPQ OFFICIAL (Name and Title) Michael M. Jodoi, Supervisor, Satellite Operations	USDA, APHIS, PPQ 3375 Koapaka Street, Suite G330 Honolulu, HI 96819		
14. SIGNATURE			
15. STATE AGENCY OFFICIAL (Name and Title) N/A	16. ADDRESS N/A		
17. SIGNATURE N/A			
PPQ FORM 519 REPLACES PPQ 274, 519, 560, AI AUG 1977	REPLACES PPQ 274, 519, 560, AND AQI 83, WHICH ARE OBSOLETE		

Attachment III-G-10 Soil Samples Restricted Entry Labels

U.S. DEPARTMENT OF AGRICULTURE ANIMAL AND PLANT HEALTH INSPECTION SERVICE PLANT PROTECTION AND QUARANTINE HYATTSVILLE, MARYLAND 20782

soil samples

restricted entry

The material contained in this package is imported under authority of the Federal Plant Pest Act of May 23, 1957.

For release without treatment if addressee is currently listed as approved by Plant Protection and Quarantine.

PPQ FORM 550

Edition of 12/77 may be used

(JAN 83)

U.S. DEPARTMENT OF AGRICULTURE ANIMAL AND PLANT HEALTH INSPECTION SERVICE PLANT PROTECTION AND QUARANTINE HYATTSVILLE, MARYLAND 20782

soil samples

restricted entry

The material contained in this package is imported under authority of the Federal Plant Pest Act of May 23, 1957.

For release without treatment if addressee is currently listed as approved by Plant Protection and Quarantine.

PPQ FORM 550

Edition of 12/77 may be used

(JAN 83)

U.S. DEPARTMENT OF AGRICULTURE ANIMAL AND PLANT HEALTH INSPECTION SERVICE PLANT PROTECTION AND QUARANTINE HYATTSVILLE, MARYLAND 20782

soil samples

restricted entry

The material contained in this package is imported under authority of the Federal Plant Pest Act of May 23, 1957.

For release without treatment if addressee is currently listed as approved by Plant Protection and Quarantine.

PPQ FORM 550

Edition of 12/77 may be used

(JAN 83)



Page 1 of 3

EQUIPMENT DECONTAMINATION

1.0 PURPOSE

The standard operating procedure (SOP) describes general methods of equipment decontamination (decon) for use by U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) field personnel and their contractors during field sampling activities. Some sites may require additional steps (e.g. nitric rinses for metals, hexane for chlorinated pesticides) to insure equipment is properly deconned. These should be identified and addressed in the Work Plans and/or the Quality Assurance Project Plans (QAPPs)

2.0 PROCEDURES

Decontamination of equipment is necessary to prevent cross-contamination and to maintain the highest integrity possible in collected samples. Planning a decontamination program should include consideration of the following factors:

- The location where the decon procedures will be conducted
- The types of equipment requiring decon
- The frequency of equipment decontamination
- The cleaning technique and types of cleaning solutions appropriate to the contaminants of concern
- The method for containing the residual contaminants and wash water from the deconning process
- The use of a quality control measure to determine the effectiveness of the decontamination procedure (e.g. equipment rinsate samples)

This subsection describes standards for decontamination, including the techniques to be used, frequency of decontamination, cleaning solutions, and effectiveness.

2.1 DECONTAMINATION AREA

An appropriate location for the decontamination area at a site shall be selected on the basis of the ability to control access to the area, control residual material removed from equipment, the need to store dirty and clean equipment, and the ability to restrict access to the area being investigated. The decontamination area shall be located an adequate distance away and upwind from potential contaminant sources to avoid contamination of clean equipment.

2.2 TYPES OF EQUIPMENT

Examples of drilling equipment that must be deconned includes drill bits, auger sections, split spoon samplers, and hand tools. Decontamination of monitoring well development and ground-water sampling equipment includes submersible pumps, non-disposable bailers, interface probes, water level meters,

bladder pumps, airlift pumps, and lysimeters. Other sampling equipment that may require decontamination includes, but is not limited to, hand trowels, hand augers, slide hammer samplers, shovels, stainless steel spoons and bowls, soil sample liners and caps, wipe sampling templates, COLIWASA samplers, and dippers. Equipment with a porous surface, such as rope, cloth hoses, and wooden blocks, cannot be thoroughly decontaminated and should be properly disposed of after one use.

2.3 FREQUENCY OF EQUIPMENT DECONTAMINATION

Down-hole drilling equipment and equipment used in monitoring well development and purging shall be decontaminated prior to initial use and between each borehole or well. However, down hole drilling equipment may require more frequent cleaning to prevent cross-contamination between vertical zones within a single borehole. When drilling through a shallow contaminated zone and installing a surface casing to seal off the contaminated zone, the drilling tools shall be decontaminated prior to drilling deeper. Groundwater sampling should be initiated by sampling ground water from the monitoring well where the least contamination is suspected. This is more important when not using disposable equipment. All groundwater, surface water, and soil sampling devices shall be decontaminated prior to initial use and between collection of each sample to prevent the possible introduction of contaminants into successive samples.

2.4 CLEANING SOLUTIONS AND TECHNIQUES

Decontamination can be accomplished using a variety of techniques and fluids. The preferred method of decontaminating major equipment such as drill bits, augers, drill string, pump drop-pipe, etc., is steam cleaning. Steam cleaning is accomplished using a portable, high-pressure steam cleaner equipped with a pressure hose and fittings. For this method, equipment shall be thoroughly steam washed and rinsed with potable tap water to remove particulates and contaminants.

A rinse decontamination procedure is acceptable for equipment such as bailers, water level meters, new and re-used soil sample liners, and hand tools. The decontamination procedure shall consist of the following: (1) wash with a non-phosphate detergent (Citrinox®, Liquinox®, or other suitable phosphate free detergent) and potable water solution, (2) rinse with potable water, and (3) rinses with deionized or distilled water. Equipment shall be disassembled as much as is practical, prior to cleaning. An initial gross wash scrub down and quick rinse should be completed at the beginning of the process if equipment is heavily soiled. After decontamination, care needs to be taken that the cleaned equipment does not become contaminated. This may require wrapping items in uncoated foil or plastic and storing the equipment in a specified "clean" area.

Decontaminating submersible pumps requires additional effort because internal surfaces become contaminated during usage. The pumps shall be decontaminated by circulating fluids through the pump while it is operating. This circulation can be done using a clean 4-inch or greater diameter pipe equipped with an end cap. The pipe shall be filled with enough decon fluid to submerge the pump, the pump placed within the capped pipe, and the pump operated while circulating the fluids within the pipe. The decontamination sequence shall include (1) detergent and potable water, (2) potable water rinse, and (3) deionized or distilled water rinse. The decontamination fluids shall be changed after each cycle. Changing of the fluids may include dumping of the detergent water, mixing detergent in the potable water rinse, using the deionized water as the potable rinse and renewing the distilled/deionized water. All decon water shall be disposed of as outlined in the field work plans.

Decontamination solvent(s) to be used during field activities will be specified in Project Work Plans or QAPPs. If solvents are used, sufficient time must be allowed to insure the solvent has evaporated from the equipment prior to reuse.

Equipment used for measuring field parameters such as pH, temperature, specific conductivity, and turbidity shall be rinsed with deionized or distilled water. New, unused soil sample liners and caps will be cleaned using the three step process, outlined above, to remove any dirt or cutting oils that may be on them prior to use.

2.5 CONTAINMENT OF RESIDUAL CONTAMINANTS AND CLEANING SOLUTIONS

Decontamination program for equipment exposed to potentially hazardous materials requires a provision for catchment and disposal of the contaminated material, cleaning solution, and wash water. This may require setting up a containment area with a system for pumping the water generated decontamination water into proper containers.

Clean equipment should be stored in a separate location to prevent recontamination. Decontamination fluids contained within the bermed area shall be collected and disposed of as outlined in the field sampling plan.

Containment of fluids from the decontamination of lighter-weight drilling equipment and hand-held sampling devices shall be accomplished using wash buckets or tubs. The decontamination fluids shall be collected and disposed of as outlined in the field sampling plan.

2.6 EFFECTIVENESS OF DECONTAMINATION PROCEDURES

A decontamination program must incorporate quality control measures to determine the effectiveness of cleaning methods. Quality control measures typically include collection of equipment rinsate samples or wipe testing. Equipment rinsates consist of analyte-free water that has been poured over or through the sample collection equipment after its final decontamination rinse. Wipe testing is performed by wiping a cloth over the surface of the equipment after cleaning. Further descriptions of these samples and their required frequency of collection are provided in SOP III-B, *Field QC Samples (Water, Soil)*. These quality control measures provide "after-the fact" information that may be useful in determining whether or not cleaning methods were effective in removing the contaminants of concern.

3.0 DOCUMENTATION

The decontamination process shall be recorded in the field logbook.

4.0 REFERENCES

SOP III-B, Field QC Samples (Water, Soil).

5.0 ATTACHMENTS

None.



Revised April 2015

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EQUIPMENT CALIBRATION, OPERATION, AND MAINTENANCE

1.0 PURPOSE

This standard operating procedure (SOP) describes the activities and responsibilities of the U.S. Naval Facilities Engineering Command Northwest (NAVFAC NW) personnel pertaining to the operating, calibration, and maintenance of equipment used to collect environmental data. Reliable measurements of data required by the field sampling plan are necessary because the information recorded may be the basis for development of remedial action and responses.

2.0 PROCEDURES

2.1 EQUIPMENT CALIBRATION

All water quality monitoring equipment will be calibrated and adjusted to operate within the manufacturers' specifications. Water quality instruments and equipment that require calibration are to be calibrated to specifications prior to field use. In addition, a one-point calibration check is made at midday and at intervals outlined in the field sampling plan. A final check is conducted at the end of each field day. This is not a recalibration of the meter but a check of the calibration to ensure the continued accuracy of the meter. All calibration information shall be recorded in the project logbook.

Special attention shall be paid to instruments that may be affected by the change in the ambient temperature or humidity. Calibration checks should also be performed when sampling conditions change significantly, a change of sample matrix, and/or readings are unstable or there is a change of parameter measurements that appear unusual.

2.2 EQUIPMENT MAINTENANCE

All field monitoring equipment, field sampling equipment, and accessories are to be maintained in accordance with the manufacturer's recommendations and specifications and/or established field practices. All maintenance will be performed by qualified personnel and documented in the field logbook.

Equipment requiring battery charging shall be charged as recommended by the manufacturer. Backup batteries for meters requiring them shall be included as part of the meters accessories. Care must be taken to protect meters from adverse elements. This may involve placing the meter in a large plastic bag to shield it from the weather.

3.0 DOCUMENTATION

All field equipment calibration, maintenance, and operation information shall be recorded within the field logbook. This is to document that appropriate procedures have been followed and to track the equipment operation. All entries in the field logbook must be written accurately and legibly as outlined in the SOP III-D, *Logbooks*.

Logbook entries shall contain, but are not necessarily limited to, the following:

Revised April 2015

- Equipment model and serial numbers
- Date and time of calibration or maintenance performed
- Calibration standard used
- Calibration lot number and expiration date if listed on bottle
- Calibration procedure used if there are multiple options
- Calibration and calibration check readings including units used
- Problems and solutions regarding use, calibration or maintenance of the equipment
- And other pertinent information

4.0 REFERENCES

SOP III-D, Logbooks

5.0 ATTACHMENTS

None.

Appendix C
Department of Defense Environmental
Laboratory Accreditation Program
Accreditation Letter



PERRY JOHNSON LABORATORY ACCREDITATION, INC.

Certificate of Accreditation

Perry Johnson Laboratory Accreditation, Inc. has assessed the Laboratory of:

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061

(Hereinafter called the Organization) and hereby declares that Organization has met the requirements of ISO/IEC 17025:2017) General Requirements for the competence of Testing and Calibration Laboratories and the United States Department of Defense Environmental Laboratory Accreditation Program (DoD-ELAP) requirements identified within the DoD/DOE Quality Systems Manual (DoD/DOE QSM) Version 5.3 May 2019 and is accredited is accordance with the:

United States Department of Defense Environmental Laboratory Accreditation Program (DoD-ELAP)

This accreditation demonstrates technical competence for a defined scope and the operation of a laboratory quality management system (as outlined by the joint ISO-ILAC-IAF Communiqué dated April 2017):

Environmental Testing (As detailed in the supplement)

Accreditation claims for such testing and/or calibration services shall only be made from addresses referenced within this certificate. This Accreditation is granted subject to the system rules governing the Accreditation referred to above, and the Organization hereby covenants with the Accreditation body's duty to observe and comply with the said rules.

For PJLA:

Tracy Szerszen

President

Initial Accreditation Date:

Issue Date:

Expiration Date:

November 17, 2016

February 22, 2021

March 31, 2023

Accreditation No:

Certificate No.:

91667

L21-123

The validity of this certificate is maintained through ongoing assessments based on a continuous accreditation cycle. The validity of this certificate should be confirmed through the PJLA website: www.pjlabs.com

Perry Johnson Laboratory Accreditation, Inc. (PJLA) 755 W. Big Beaver, Suite 1325 Troy, Michigan 48084



Certificate of Accreditation: Supplement

ISO/IEC 17025:2017 and DoD-ELAP

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Matrix	Standard/Method	Technology	Analyte
Drinking Water	EPA 533	LC/MS/MS	11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11Cl-PF3OUdS)
Drinking Water	EPA 533	LC/MS/MS	9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF3ONS)
Drinking Water	EPA 533	LC/MS/MS	4,8-dioxa-3H-perfluorononanoic acid (Adona)
Drinking Water	EPA 533	LC/MS/MS	Hexafluoropropylene oxide dimer acid (HFPO-DA)
Drinking Water	EPA 533	LC/MS/MS	Nonafluoro-3,6-Dioxaheptanoic Acid (NFDHA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-butanoic Acid (PFBA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-1-butanesulfonic Acid (PFBS)
Drinking Water	EPA 533	LC/MS/MS	1H,1H,2H,2H-Perfluorodecane sulfonate (8:2FTS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-decanoic Acid (PFDA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-dodecanoic acid (PFDoA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro(2-ethoxyethane)sulfonic Acid (PFEESA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-1-heptanesulfonate (PFHpS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-heptanoic Acid (PFHpA)
Drinking Water	EPA 533	LC/MS/MS	1H,1H,2H,2H-Perfluorohexane sulfonate (4:2FTS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-1-hexanesulfonic Acid (PFHxS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-hexanoic acid (PFHxA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-3-Methoxypropanoic Acid (PFMPA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-4-Methoxybutanoic Acid (PFMBA)
Drinking Water	EPA 533	LC/MS/MS	Perfluorononanoic acid (PFNA)
Drinking Water	EPA 533	LC/MS/MS	1H,1H,2H,2H-Perfluorooctane sulfonate (6:2FTS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-1-octanesulphonic Acid (PFOS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-octanoic Acid (PFOA)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-pentanoic acid (PFPeA)
Drinking Water	EPA 533	LC/MS/MS	Sodium perfluoro-1-pentanesulfonate (PFPeS)
Drinking Water	EPA 533	LC/MS/MS	Perfluoro-n-undecanoic acid (PFUnA)
Drinking Water	EPA 537.1.1	LC/MS/MS	4,8-dioxa-3H-perfluorononanoic acid (ADONA)
Drinking Water	EPA 537.1.1	LC/MS/MS	9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF3ONS)
Drinking Water	EPA 537.1.1	LC/MS/MS	11-chloroeicosafluoro-3-oxaundecane-1- sulfonic acid (11Cl-PF3OUdS)
Drinking Water	EPA 537.1.1	LC/MS/MS	Hexafluoropropylene oxide dimer acid (HFPO-DA)



Certificate of Accreditation: Supplement

ISO/IEC 17025:2017 and DoD-ELAP

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Matrix	Standard/Method	Technology	Analyte
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-hexanoic acid (PFHxA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-heptanoic Acid (PFHpA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-octanoic Acid (PFOA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluorononanoic acid (PFNA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-decanoic Acid (PFDA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-undecanoic acid (PFUnA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-dodecanoic acid (PFDoA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-tridecanoic acid (PFTrDA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-n-tetradecanoic acid (PFTeDA)
Drinking Water	EPA 537.1.1	LC/MS/MS	N-methylperfluoro-1-octanesulfonamidoacetic acid (NMeFOSAA)
Drinking Water	EPA 537.1.1	LC/MS/MS	N-ethylperfluoro-octanesulfonamidoacetic acid (NEtFOSAA)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-1-butanesulfonic Acid (PFBS)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-1-hexanesulfonic Acid (PFHxS)
Drinking Water	EPA 537.1.1	LC/MS/MS	Perfluoro-1-octanesulphonic Acid (PFOS)
Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	3-Perfluoropropyl propanoic Acid (3:3 FTCA)
Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	3-Perfluoropropyl propanoic acid (5:3 FTCA)
Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	3-Perfluoropropyl propanoic acid (7:3 FTCA)
Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-octanesulfonamide (PFOSA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	4,8-dioxa-3H-perfluorononanoic acid (Adona)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	9-chlorohexadecafluoro-3-oxanonane-1-sulfonic acid (9CI-PF3ONS)



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ISO/IEC 17025:2017 and DoD-ELAP

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141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Matrix	Standard/Method	Technology	Analyte
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11CI-PF3OUdS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Hexafluoropropylene oxide dimer acid (HFPODA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Sodium perfluoro-1-pentanesulfonate (PFPeS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-nonanesulfonate (PFNS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-heptanesulfonate (PFHpS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	N-ethylperfluoro-octanesulfonamidoacetic acid (NEtFOSAA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	1H,1H,2H,2H-Perfluorohexane sulfonate (4:2FTS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	1H,1H,2H,2H-Perfluorooctane sulfonate (6:2FTS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	1H,1H,2H,2H-Perfluorodecane sulfonate (8:2FTS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-butanoic Acid (PFBA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-pentanoic acid (PFPeA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-hexanoic acid (PFHxA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-heptanoic Acid (PFHpA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-octanoic Acid (PFOA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluorononanoic acid (PFNA)



ISO/IEC 17025:2017 and DoD-ELAP

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Accreditation is granted to the facility to perform the following testing:

Matrix	Standard/Method	Technology	Analyte
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-decanoic Acid (PFDA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-undecanoic acid (PFUnA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-dodecanoic acid (PFDoA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-tridecanoic acid (PFTrDA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-n-tetradecanoic acid (PFTeDA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	N-methylperfluoro-1-octanesulfonamidoacetic acid (NMeFOSAA)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-butanesulfonic Acid (PFBS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-hexanesulfonic Acid (PFHxS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-octanesulphonic Acid (PFOS)
Aqueous/Solids/Tissues	PFAS by LCMSMS Compliant with QSM 5.3 Table B-15	LC/MS/MS	Perfluoro-1-decanesulfonate (PFDS)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,3',4,4'-Hexachlorobiphenyl (BZ 128)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,3',4,5-Hexachlorobiphenyl (BZ 129)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4,4',5,5'-Heptachlorobiphenyl (BZ 180)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4,4',5',6-Heptachlorobiphenyl (BZ 183)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4,4',5'-Hexachlorobiphenyl (BZ 138)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4,4',6,6'-Heptachlorobiphen yl (BZ 184)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4',5,5',6-Heptachlorobiphenyl (BZ 187)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,4,5'-Pentachlorobiphenyl (BZ 87)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',3,5'-Tetrachlorobiphenyl (BZ 44)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',4,4',5,5'-Hexachlorobiphenyl (BZ 153)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',4,5,5'-Pentachlorobiphenyl (BZ 101)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',4,5'-Tetrachlorobiphenyl (BZ 49)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',5,5'-Tetrachlorobiphenyl (BZ 52)

Issue: 02/2021 This supplement is in conjunction with certificate #L21-123

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ISO/IEC 17025:2017 and DoD-ELAP

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Accreditation is granted to the facility to perform the following testing:

Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,2',5-Trichlorobiphenyl (BZ 18)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,3,3',4,4'-Pentachlorobiphenyl (BZ 105)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,3,3',4',6-Pentachlorobiphenyl (BZ 110)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,3',4,4',5-Pentachlorobiphenyl (BZ 118)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,3',4,4'-Tetrachlorobiphenyl (BZ 66)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,4,4'-Trichlorobiphenyl (BZ 28)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,4'-DDD
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,4'-DDE
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,4'-DDT
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	2,4'-Dichlorobiphenyl (BZ 8)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	3,3',4,4',5,5'-Hexachlorobiphenyl (BZ 169)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	3,3',4,4',5-Pentachlorobiphenyl (BZ 126)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	3,3',4,4'-Tetrachlorobiphenyl (BZ 77)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	4,4'-DDD
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	4,4'-DDE
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	4,4'-DDT
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Aldrin
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	alpha-BHC (alpha-Hexachlorocyclohexane)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	alpha-Chlordane
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	beta-BHC (beta-Hexachlorocyclohexane)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Chlorpyrifos
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	cis-Nonachlor
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Decachlorobiphenyl (BZ 209)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	delta-BHC
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Dieldrin
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endosulfan II
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endosulfan I
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endosulfan sulfate
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endrin
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endrin aldehyde
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Endrin ketone
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	gamma-BHC (Lindane, gamma-Hexachlorocyclohexane)
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	gamma-Chlordane
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Heptachlor
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Heptachlor epoxide
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Hexachlorobenzene
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Certificate of Accreditation: Supplement

ISO/IEC 17025:2017 and DoD-ELAP

Battelle

141 Longwater Drive, Suite 202, Norwell, MA 02061 Contact Name: Jonathan Thorn Phone: 781-681-5565

Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Methoxychlor
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Mirex
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	Oxychlordane
Aqueous/Solid/Tissue	EPA 8081 MOD	GC-ECD	trans-Nonachlor
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	1,4-Dichlorobenzene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	1-Methylnaphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	1-Methylphenanthrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',5,5',6-Nonachlorobiphenyl (BZ 206)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',5,5'-Octachlorobiphenyl (BZ 194)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',5,6,6'-Nonachlorobiphenyl (BZ 207)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',5,6-Octachlorobiphenyl (BZ 195)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',5-Heptachlorobiphenyl (BZ 170)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',6,6'-Octachlorobiphenyl (BZ 197)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4',6-Heptachlorobiphenyl (BZ 171)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,4'-Hexachlorobiphenyl (BZ 128)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,5',6,6'-Nonachlorobiphenyl (BZ 208)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,5',6-Octachlorobiphenyl (BZ 198)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,5',6'-Octachlorobiphenyl (BZ 199)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,5'-Heptachlorobiphenyl (BZ 172)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,6,6'-Octachlorobiphenyl (BZ 200)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5',6,6'-Octachlorobiphenyl (BZ 201)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,6-Heptachlorobiphenyl (BZ 173)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5,6'-Heptachlorobiphenyl (BZ 174)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5',6-Heptachlorobiphenyl (BZ 175)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5',6'-Heptachlorobiphenyl (BZ 177)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,5'-Hexachlorobiphenyl (BZ 130)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,6,6'-Heptachlorobiphenyl (BZ 176)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4,6-Hexachlorobiphenyl (BZ 131)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',4-Pentachlorobiphenyl (BZ 82)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5,5',6,6'-Octachlorobiphenyl (BZ 202)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5,5',6-Heptachlorobiphenyl (BZ 178)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5,6,6'-Heptachlorobiphenyl (BZ 179)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5,6-Hexachlorobiphenyl (BZ 134)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5,6'-Hexachlorobiphenyl (BZ 135)



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Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',5-Pentachlorobiphenyl (BZ 83)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',6,6'-Hexachlorobiphenyl (BZ 136)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3',6-Pentachlorobiphenyl (BZ 84)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,3'-Tetrachlorobiphenyl (BZ 40)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',5,5',6-Octachlorobiphenyl (BZ 203)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',5,5'-Heptachlorobiphenyl (BZ 180)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',5',6-Heptachlorobiphenyl (BZ 183)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',5-Hexachlorobiphenyl (BZ 137)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',5'-Hexachlorobiphenyl (BZ 138)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',6,6'-Heptachlorobiphenyl (BZ 184)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',6-Hexachlorobiphenyl (BZ 139)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4',6'-Hexachlorobiphenyl (BZ 140)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,4'-Pentachlorobiphenyl (BZ 85)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,5,5',6-Heptachlorobiphenyl (BZ 185)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',5,5',6-Heptachlorobiphenyl (BZ 187)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,5,5'-Hexachlorobiphenyl (BZ 141)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',5,5'-Hexachlorobiphenyl (BZ 146)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',5,6,6'-Heptachlorobiphenyl (BZ 188)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',5',6-Hexachlorobiphenyl (BZ 149)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,5',6-Hexachlorobiphenyl (BZ 144)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4,5'-Pentachlorobiphenyl (BZ 87)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',5'-Pentachlorobiphenyl (BZ 97)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4',6-Pentachlorobiphenyl (BZ 91)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4-Tetrachlorobiphenyl (BZ 41)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,4'-Tetrachlorobiphenyl (BZ 42)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,5,5',6-Hexachlorobiphenyl (BZ 151)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,5,5'-Pentachlorobiphenyl (BZ 92)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,5',6-Pentachlorobiphenyl (BZ 95)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,5-Tetrachlorobiphenyl (BZ 43)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,5'-Tetrachlorobiphenyl (BZ 44)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,6'-Tetrachlorobiphenyl (BZ 46)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3,6-Tetrachlorobiphenyl (BZ 45)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',3-Trichlorobiphenyl (BZ 16)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4',5,5'-Hexachlorobiphenyl (BZ 153)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4',5,6'-Hexachlorobiphenyl (BZ 154)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4',5-Pentachlorobiphenyl (BZ 99)



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Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4',6,6'-Hexachlorobiphenyl (BZ 155)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4',6-Pentachlorobiphenyl (BZ 100)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,4'-Tetrachlorobiphenyl (BZ 47)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,5,5'-Pentachlorobiphenyl (BZ 101)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,5-Tetrachlorobiphenyl (BZ 48)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,5'-Tetrachlorobiphenyl (BZ 49)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,6,6'-Pentachlorobiphenyl (BZ 104)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,6'-Tetrachlorobiphenyl (BZ 51)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4,6-Tetrachlorobiphenyl (BZ 50)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',4-Trichlorobiphenyl (BZ 17)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',5,5'-Tetrachlorobiphenyl (BZ 52)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',5,6'-Tetrachlorobiphenyl (BZ 53)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',5-Trichlorobiphenyl (BZ 18)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',6,6'-Tetrachlorobiphenyl (BZ 54)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2',6-Trichlorobiphenyl (BZ 19)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,2'-Dichlorobiphenyl (BZ 4)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5,5',6-Octachlorobiphenyl (BZ 205)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5,5'-Heptachlorobiphenyl (BZ 189)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5,6-Heptachlorobiphenyl (BZ 190)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5',6-Heptachlorobiphenyl (BZ 191)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5-Hexachlorobiphenyl (BZ 156)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',5'-Hexachlorobiphenyl (BZ 157)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4',6-Hexachlorobiphenyl (BZ 158)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4,4'-Pentachlorobiphenyl (BZ 105)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4',5,5',6-Heptachlorobiphenyl (BZ 193)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4',5,6-Hexachlorobiphenyl (BZ 163)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4',5',6-Hexachlorobiphenyl (BZ 164)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4',6-Pentachlorobiphenyl (BZ 110)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,3',4'-Tetrachlorobiphenyl (BZ 56)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4,4',5,5'-Hexachlorobiphenyl (BZ 167)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4,4',5,6-Hexachlorobiphenyl (BZ 166)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4,4',5-Pentachlorobiphenyl (BZ 114)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4,4',5-Pentachlorobiphenyl (BZ 118)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4,4',5'-Pentachlorobiphenyl (BZ 123)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4,4',6-Pentachlorobiphenyl (BZ 115)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4,4'-Tetrachlorobiphenyl (BZ 60)



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Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4,4'-Tetrachlorobiphenyl (BZ 66)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4',5,5'-Pentachlorobiphenyl (BZ 124)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4',5',6-Pentachlorobiphenyl (BZ 125)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4',5-Tetrachlorobiphenyl (BZ 63)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4,5-Tetrachlorobiphenyl (BZ 67)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4',5-Tetrachlorobiphenyl (BZ 70)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4',6-Tetrachlorobiphenyl (BZ 64)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4',6-Tetrachlorobiphenyl (BZ 71)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,4'-Trichlorobiphenyl (BZ 22)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4-Trichlorobiphenyl (BZ 25)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',4'-Trichlorobiphenyl (BZ 33)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',5-Trichlorobiphenyl (BZ 26)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,5-Trimethylnaphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3,6-Trichlorobiphenyl (BZ 24)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3',6-Trichlorobiphenyl (BZ 27)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3-Dichlorobiphenyl (BZ 5)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,3'-Dichlorobiphenyl (BZ 6)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4,4',5-Tetrachlorobiphenyl (BZ 74)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4,4',6-Tetrachlorobiphenyl (BZ 75)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4,4'-Trichlorobiphenyl (BZ 28)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4,5-Trichlorobiphenyl (BZ 29)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4',5-Trichlorobiphenyl (BZ 31)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4,6-Trichlorobiphenyl (BZ 30)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4',6-Trichlorobiphenyl (BZ 32)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4'-DDD
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4'-DDE
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4'-DDT
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4-Dichlorobiphenyl (BZ 7)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,4'-Dichlorobiphenyl (BZ 8)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,5-Dichlorobiphenyl (BZ 9)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2,6-Dimethylnaphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2-Chlorobiphenyl (BZ 1)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2-Chloronaphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2-Methylnaphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	2-Methylphenanthrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3',4,4',5,5'-Hexachlorobiphenyl (BZ 169)



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Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3',4,4',5-Pentachlorobiphenyl (BZ 126)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3',4,4'-Tetrachlorobiphenyl (BZ 77)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3',4,5,5'-Pentachlorobiphenyl (BZ 127)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3',5,5'-Tetrachlorobiphenyl (BZ 80)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,3'-Dichlorobiphenyl (BZ 11)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,4,4',5-Tetrachlorobiphenyl (BZ 81)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,4,4'-Trichlorobiphenyl (BZ 37)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,4-Dichlorobiphenyl (BZ 12)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,4'-Dichlorobiphenyl (BZ 13)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	3,6-Dimethylphenanthrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	4,4'-Dichlorobiphenyl (BZ 15)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	4-Chlorobiphenyl (BZ 3)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Acenaphthene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Acenaphthylene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Anthracene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(a)anthracene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(a)pyrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(b)fluoranthene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(b)thiophene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(e)pyrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(g,h,i)perylene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Benzo(k)fluoranthene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Biphenyl
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Chrysene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	cis-Decalin
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Decachlorobiphenyl (BZ 209)
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Dibenz(a,h)anthracene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Dibenzofuran
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Dibenzothiophene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Fluoranthene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Fluorene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Indeno(1,2,3-cd)pyrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Naphthalene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Perylene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Phenanthrene
Aqueous/Solid/Tissue	EPA 8270E MOD	GC-MS	Pyrene

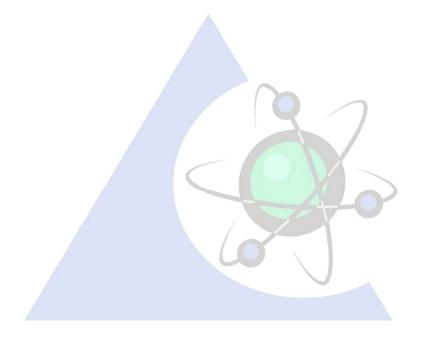


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Matrix	Standard/Method	Technology	Analyte
Aqueous/Solid/Tissue	EPA 8270DE MOD	GC-MS	trans-Decalin
Aqueous	EPA 3510 C	Separatory Funnel	Prep
Aqueous/Solid/Tissue	EPA 3640A MOD	Gel-permeation chromatography (GPC)	Cleanup
Aqueous/Solid/Tissue	EPA 3660B MOD	Sulfur Cleanup	Cleanup
Solids/Tissues	NOAA NOS ORCA 71	Orbital Shaker	Prep
Tissue	NOAA NOS ORCA 71	Tissuemizer	Prep



Appendix D Laboratory Standard Operating Procedures







Sampling and Analysis Plan
Per- and Polyfluoroalkyl Substances Remedial Investigation
Outlying Landing Field Coupeville
Naval Air Station Whidbey Island
Oak Harbor, Washington

NOTIFICATION: APPENDIX D CONTAINS SENSITIVE BUT UNCLASSIFIED INFORMATION WHICH IS PROTECTED BY THE FREEDOM OF INFORMATION ACT

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